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<p>(21) International Application Number: PCT/US98/26852</p> <p>(22) International Filing Date: 17 December 1998 (17.12.98)</p> <p>(30) Priority Data:</p> <table style="width: 100%; border: none;"> <tr> <td style="width: 30%;">08/996,441</td> <td style="width: 40%;">18 December 1997 (18.12.97)</td> <td style="width: 30%;">US</td> </tr> <tr> <td>08/993,722</td> <td>18 December 1997 (18.12.97)</td> <td>US</td> </tr> <tr> <td>08/993,775</td> <td>18 December 1997 (18.12.97)</td> <td>US</td> </tr> <tr> <td>08/993,170</td> <td>18 December 1997 (18.12.97)</td> <td>US</td> </tr> </table> <p>(63) Related by Continuation (CON) or Continuation-in-Part (CIP) to Earlier Applications</p> <table style="width: 100%; border: none;"> <tr> <td style="width: 30%;">US</td> <td style="width: 40%;">08/993,722 (CON)</td> <td style="width: 30%;"></td> </tr> <tr> <td>Filed on</td> <td>18 December 1997 (18.12.97)</td> <td></td> </tr> <tr> <td>US</td> <td>08/993,170 (CON)</td> <td></td> </tr> <tr> <td>Filed on</td> <td>18 December 1997 (18.12.97)</td> <td></td> </tr> <tr> <td>US</td> <td>08/993,775 (CON)</td> <td></td> </tr> <tr> <td>Filed on</td> <td>18 December 1997 (18.12.97)</td> <td></td> </tr> <tr> <td>US</td> <td>08/996,441 (CON)</td> <td></td> </tr> <tr> <td>Filed on</td> <td>18 December 1997 (18.12.97)</td> <td></td> </tr> </table> <p>(71) Applicants (for all designated States except US): ECOGEN, INC. [US/US]; 2005 Cabot Boulevard West, Langhorne, PA 19407-3023 (US). MONSANTO COMPANY [US/US]; 700 Chesterfield Parkway North, St. Louis, MO 63198 (US).</p>	08/996,441	18 December 1997 (18.12.97)	US	08/993,722	18 December 1997 (18.12.97)	US	08/993,775	18 December 1997 (18.12.97)	US	08/993,170	18 December 1997 (18.12.97)	US	US	08/993,722 (CON)		Filed on	18 December 1997 (18.12.97)		US	08/993,170 (CON)		Filed on	18 December 1997 (18.12.97)		US	08/993,775 (CON)		Filed on	18 December 1997 (18.12.97)		US	08/996,441 (CON)		Filed on	18 December 1997 (18.12.97)		<p>(72) Inventors; and (75) Inventors/Applicants (for US only): ENGLISH, Leigh [US/US]; 1415 Carriage Crossing, Chesterfield, MO 63005 (US). BRUSSOCK, Susan, M. [US/US]; 7 Hillside Lane, New Hope, PA 18938 (US). MALVAR, Thomas, M. [US/US]; 12046 Charter House Lane, St. Louis, MO 63146 (US). BRYSON, James, W. [US/US]; 87 Wood Stream Drive, Langhorne, PA 19053 (US). KULESZA, Caroline, A. [US/US]; 211 Raymond Avenue, Charlottesville, VA 22903 (US). WALTERS, Frederick, S. [US/US]; 3413 6th Avenue, Beaver Falls, PA 15010 (US). SLATIN, Stephen, L. [US/US]; 3823 Leslie Place, Fair Lawn, NJ 07410 (US). VON TERSCH, Michael, A. [US/US]; 14 Rutledge Avenue, Trenton, NJ 08618 (US). ROMANO, Charles [US/US]; 2402 Maple Crossing Drive, Wildwood, MO 63011 (US).</p> <p>(74) Agent: MOORE, Mark, D.; Arnold White & Durkee, P.O. Box 4433, Houston, TX 77210 (US).</p> <p>(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).</p> <p>Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i></p>
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<p>(54) Title: INSECT-RESISTANT TRANSGENIC PLANTS AND METHODS FOR IMPROVING δ-ENDOTOXIN ACTIVITY AGAINST TARGET INSECTS</p>																																					
<p>(57) Abstract</p> <p>Disclosed are methods for increasing the activity of <i>B. thuringiensis</i> δ-endotoxins against Coleopteran insect pests. Also disclosed are methods for mutagenizing nucleic acid sequences encoding these polypeptides, and increasing insect resistance in transgenic plants expressing these genes.</p>																																					

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DESCRIPTION

INSECT-RESISTANT TRANSGENIC PLANTS AND METHODS FOR IMPROVING δ -ENDOTOXIN ACTIVITY AGAINST TARGET INSECTS

5

1.0 BACKGROUND OF THE INVENTION

1.1 FIELD OF THE INVENTION

This invention relates to methods for producing genetically-engineered, re-combinant δ -endotoxins derived from *Bacillus thuringiensis* that are useful in the control of southern corn rootworm (*Diabrotica undecimpunctata howardi* Barber) and western corn rootworm (*Diabrotica virgifera virgifera* LeConte).

1.2 DESCRIPTION OF THE RELATED ART

Almost all field crops, plants, and commercial farming areas are susceptible to attack by one or more insect pests. Particularly problematic are Coleopteran and Lepidoptern pests. For example, vegetable and cole crops such as artichokes, kohlrabi, arugula, leeks, asparagus, lentils, beans, lettuce (*e.g.*, head, leaf, romaine), beets, bok choy, malanga, broccoli, melons (*e.g.*, muskmelon, watermelon, crenshaw, honeydew, cantaloupe), brussels sprouts, cabbage, cardoni, carrots, napa, cauliflower, okra, onions, celery, parsley, chick peas, parsnips, chicory, peas, chinese cabbage, peppers, collards, potatoes, cucumber, pumpkins, cucurbits, radishes, dry bulb onions, rutabaga, eggplant, salsify, escarole, shallots, endive, soybean, garlic, spinach, green onions, squash, greens, sugar beets, sweet potatoes, turnip, swiss chard, horseradish, tomatoes, kale, turnips, and a variety of spices are sensitive to infestation by one or more of the following insect pests: alfalfa looper, armyworm, beet armyworm, artichoke plume moth, cabbage budworm, cabbage looper, cabbage webworm, corn earworm, celery leafeater, cross-striped cabbageworm, european corn borer, diamondback moth, green cloverworm, imported cabbageworm, melonworm, omnivorous leafroller, pickleworm, rindworm complex, saltmarsh caterpillar, soybean looper, tobacco budworm, tomato fruitworm, tomato

hornworm, tomato pinworm, velvetbean caterpillar, and yellowstriped armyworm. Likewise, pasture and hay crops such as alfalfa, pasture grasses and silage are often attacked by such pests as armyworm, beef armyworm, alfalfa caterpillar, European skipper, a variety of loopers and webworms, as well as yellowstriped armyworms.

5 Fruit and vine crops such as apples, apricots, cherries, nectarines, peaches, pears, plums, prunes, quince almonds, chestnuts, filberts, pecans, pistachios, walnuts, citrus, blackberries, blueberries, boysenberries, cranberries, currants, loganberries, raspberries, strawberries, grapes, avocados, bananas, kiwi, persimmons, pomegranate, pineapple, tropical fruits are often susceptible to attack and defolia-
10 tion by achema sphinx moth, amorbia, armyworm, citrus cutworm, banana skipper, blackheaded fireworm, blueberry leafroller, cankerworm, cherry fruitworm, citrus cutworm, cranberry girdler, eastern tent caterpillar, fall webworm, fall webworm, filbert leafroller, filbert webworm, fruit tree leafroller, grape berry moth, grape leafroller, grapeleaf skeletonizer, green fruitworm, gummos-batrachedra com-
15 mosae, gypsy moth, hickory shuckworm, hornworms, loopers, navel orangeworm, obliquebanded leafroller, omnivorous leafroller. omnivorous looper, orange tortrix, orangedog, oriental fruit moth, pandemis leafroller, peach twig borer, pecan nut casebearer, redbanded leafroller, redhumped caterpillar, roughskinned cutworm, saltmarsh caterpillar, spanworm, tent caterpillar, thecla-thecla basillides, tobacco
20 budworm, tortrix moth, tufted apple budmoth, variegated leafroller, walnut caterpillar, western tent caterpillar, and yellowstriped armyworm.

 Field crops such as canola/rape seed, evening primrose, meadow foam, corn (field, sweet, popcorn), cotton, hops, jojoba, peanuts, rice, safflower, small grains (barley, oats, rye, wheat, *etc.*), sorghum, soybeans, sunflowers, and tobacco are of-
25 ten targets for infestation by insects including armyworm, asian and other corn borers; banded sunflower moth, beet armyworm, bollworm, cabbage looper, corn rootworm (including southern and western varieties), cotton leaf perforator, diamondback moth, european corn borer, green cloverworm, headmoth, headworm, imported cabbageworm, loopers (including *Anacamptodes* spp.), obliquebanded
30 leafroller, omnivorous leafier, podworm, podworm, saltmarsh caterpillar, south-

western corn borer, soybean looper, spotted cutworm, sunflower moth, tobacco budworm, tobacco hornworm, velvetbean caterpillar,

Bedding plants, flowers, ornamentals, vegetables and container stock are frequently fed upon by a host of insect pests such as armyworm, azalea moth, beet
5 armyworm, diamondback moth, ello moth (hornworm), Florida fern caterpillar, Io moth, loopers, oleander moth, omnivorous leafroller, omnivorous looper, and tobacco budworm.

Forests, fruit, ornamental, and nut-bearing trees, as well as shrubs and other nursery stock are often susceptible to attack from diverse insects such as bagworm,
10 blackheaded budworm, browntail moth, california oakworm, douglas fir tussock moth, elm spanworm, fall webworm, fruittree leafroller, greenstriped mapleworm, gypsy moth, jack pine budworm, mimosa webworm, pine butterfly, redhumped caterpillar, saddleback caterpillar, saddle prominent caterpillar, spring and fall cankerworm, spruce budworm, tent caterpillar, tortrix, and western tussock moth.
15 Likewise, turf grasses are often attacked by pests such as armyworm, sod webworm, and tropical sod webworm.

Because crops of commercial interest are often the target of insect attack, environmentally-sensitive methods for controlling or eradicating insect infestation are desirable in many instances. This is particularly true for farmers, nurserymen,
20 growers, and commercial and residential areas which seek to control insect populations using eco-friendly compositions.

The most widely used environmentally-sensitive insecticidal formulations developed in recent years have been composed of microbial pesticides derived from the bacterium *Bacillus thuringiensis*. *B. thuringiensis* is a Gram-positive
25 bacterium that produces crystal proteins or inclusion bodies which are specifically toxic to certain orders and species of insects. Many different strains of *B. thuringiensis* have been shown to produce insecticidal crystal proteins. Compositions including *B. thuringiensis* strains which produce insecticidal proteins have been commercially-available and used as environmentally-acceptable insecticides because they are quite toxic to the specific target insect, but are harmless to
30 plants and other non-targeted organisms.

1.2.1 δ -ENDOTOXINS

δ -endotoxins are used to control a wide range of leaf-eating caterpillars and beetles, as well as mosquitoes. These proteinaceous parasporal crystals, also referred to as insecticidal crystal proteins, crystal proteins, Bt inclusions, crystalline inclusions, inclusion bodies, and Bt toxins, are a large collection of insecticidal proteins produced by *B. thuringiensis* that are toxic upon ingestion by a susceptible insect host. Over the past decade research on the structure and function of *B. thuringiensis* toxins has covered all of the major toxin categories, and while these toxins differ in specific structure and function, general similarities in the structure and function are assumed. Based on the accumulated knowledge of *B. thuringiensis* toxins, a generalized mode of action for *B. thuringiensis* toxins has been created and includes: ingestion by the insect, solubilization in the insect midgut (a combination stomach and small intestine), resistance to digestive enzymes sometimes with partial digestion actually "activating" the toxin, binding to the midgut cells, formation of a pore in the insect cells and the disruption of cellular homeostasis (English and Slatin, 1992).

1.2.2 GENES ENCODING CRYSTAL PROTEINS

Many of the δ -endotoxins are related to various degrees by similarities in their amino acid sequences. Historically, the proteins and the genes which encode them were classified based largely upon their spectrum of insecticidal activity. The review by Höfte and Whiteley (1989) discusses the genes and proteins that were identified in *B. thuringiensis* prior to 1990, and sets forth the nomenclature and classification scheme which has traditionally been applied to *B. thuringiensis* genes and proteins. *cryI* genes encode lepidopteran-toxic CryI proteins. *cryII* genes encode CryII proteins that are toxic to both lepidopterans and dipterans. *cryIII* genes encode coleopteran-toxic CryIII proteins, while *cryIV* genes encode dipteran-toxic CryIV proteins.

Based on the degree of sequence similarity, the proteins were further classified into subfamilies; more highly related proteins within each family were as-

signed divisional letters such as CryIA, CryIB, CryIC, *etc.* Even more closely related proteins within each division were given names such as CryIC1, CryIC2, *etc.*

Recently a new nomenclature was developed which systematically classified the Cry proteins based upon amino acid sequence homology rather than upon insect target specificities. The classification scheme for many known toxins, not including allelic variations in individual proteins, is summarized in Table 1.

TABLE 1

KNOWN *B. THURINGIENSIS* δ -ENDOTOXINS, GENBANK ACCESSION NUMBERS,
AND REVISED NOMENCLATURE^A

New	Old	GenBank Accession #
Cry1Aa1	CryIA(a)	M11250
Cry1Aa2	CryIA(a)	M10917
Cry1Aa3	CryIA(a)	D00348
Cry1Aa4	CryIA(a)	X13535
Cry1Aa5	CryIA(a)	D175182
Cry1Aa6	CryIA(a)	U43605
Cry1Ab1	CryIA(b)	M13898
Cry1Ab2	CryIA(b)	M12661
Cry1Ab3	CryIA(b)	M15271
Cry1Ab4	CryIA(b)	D00117
Cry1Ab5	CryIA(b)	X04698
Cry1Ab6	CryIA(b)	M37263
Cry1Ab7	CryIA(b)	X13233
Cry1Ab8	CryIA(b)	M16463
Cry1Ab9	CryIA(b)	X54939
Cry1Ab10	CryIA(b)	A29125
Cry1Ac1	CryIA(c)	M11068
Cry1Ac2	CryIA(c)	M35524
Cry1Ac3	CryIA(c)	X54159
Cry1Ac4	CryIA(c)	M73249

TABLE 1 (CONT'D)

New	Old	GenBank Accession #
Cry1Ac5	CryIA(c)	M73248
Cry1Ac6	CryIA(c)	U43606
Cry1Ac7	CryIA(c)	U87793
Cry1Ac8	CryIA(c)	U87397
Cry1Ac9	CryIA(c)	U89872
Cry1Ac10	CryIA(c)	AJ002514
Cry1Ad1	CryIA(d)	M73250
Cry1Ae1	CryIA(e)	M65252
Cry1Ba1	CryIB	X06711
Cry1Ba2		X95704
Cry1Bb1	ET5	L32020
Cry1Bc1	CryIb(c)	Z46442
Cry1Bd1	CryE1	U70726
Cry1Ca1	CryIC	X07518
Cry1Ca2	CryIC	X13620
Cry1Ca3	CryIC	M73251
Cry1Ca4	CryIC	A27642
Cry1Ca5	CryIC	X96682
Cry1Ca6	CryIC	X96683
Cry1Ca7	CryIC	X96684
Cry1Cb1	CryIC(b)	M97880
Cry1Da1	CryID	X54160
Cry1Db1	PrtB	Z22511
Cry1Ea1	CryIE	X53985
Cry1Ea2	CryIE	X56144
Cry1Ea3	CryIE	M73252
Cry1Ea4		U94323
Cry1Eb1	CryIE(b)	M73253
Cry1Fa1	CryIF	M63897

TABLE 1 (CONT'D)

New	Old	GenBank Accession #
Cry1Fa2	CryIF	M63897
Cry1Fb1	PrtD	Z22512
Cry1Ga1	PrtA	Z22510
Cry1Ga2	CryIM	Y09326
Cry1Gb1	CryH2	U70725
Cry1Ha1	PrtC	Z22513
Cry1Hb1		U35780
Cry1Ia1	CryV	X62821
Cry1Ia2	CryV	M98544
Cry1Ia3	CryV	L36338
Cry1Ia4	CryV	L49391
Cry1Ia5	CryV	Y08920
Cry1Ib1	CryV	U07642
Cry1Ja1	ET4	L32019
Cry1Jb1	ET1	U31527
Cry1Ka1		U28801
Cry2Aa1	CryIIA	M31738
Cry2Aa2	CryIIA	M23723
Cry2Aa3		D86084
Cry2Ab1	CryIIB	M23724
Cry2Ab2	CryIIB	X55416
Cry2Ac1	CryIIC	X57252
Cry3Aa1	CryIIIA	M22472
Cry3Aa2	CryIIIA	J02978
Cry3Aa3	CryIIIA	Y00420
Cry3Aa4	CryIIIA	M30503
Cry3Aa5	CryIIIA	M37207
Cry3Aa6	CryIIIA	U10985
Cry3Ba1	CryIIIB	X17123

TABLE 1 (CONT'D)

New	Old	GenBank Accession #
Cry3Ba2	CryIIIB	A07234
Cry3Bb1	CryIIIB2	M89794
Cry3Bb2	CryIIIC(b)	U31633
Cry3Ca1	CryIIID	X59797
Cry4Aa1	CryIVA	Y00423
Cry4Aa2	CryIVA	D00248
Cry4Ba1	CryIVB	X07423
Cry4Ba2	CryIVB	X07082
Cry4Ba3	CryIVB	M20242
Cry4Ba4	CryIVB	D00247
Cry5Aa1	CryVA(a)	L07025
Cry5Ab1	CryVA(b)	L07026
Cry5Ba1	PS86Q3	U19725
Cry6Aa1	CryVIA	L07022
Cry6Ba1	CryVIB	L07024
Cry7Aa1	CryIIIC	M64478
Cry7Ab1	CryIIICb	U04367
Cry8Aa1	CryIIIE	U04364
Cry8Ba1	CryIIIG	U04365
Cry8Ca1	CryIIIF	U04366
Cry9Aa1	CryIG	X58120
Cry9Aa2	CryIG	X58534
Cry9Ba1	CryIX	X75019
Cry9Ca1	CryIH	Z37527
Cry9Da1	N141	D85560
Cry10Aa1	CryIVC	M12662
Cry11Aa1	CryIVD	M31737
Cry11Aa2	CryIVD	M22860
Cry11Ba1	Jeg80	X86902

TABLE 1 (CONT'D)

New	Old	GenBank Accession #
Cry12Aa1	CryVB	L07027
Cry13Aa1	CryVC	L07023
Cry14Aa1	CryVD	U13955
Cry15Aa1	34kDa	M76442
Cry16Aa1	cbm71	X94146
Cry17Aa1	cbm71	X99478
Cry18Aa1	CryBP1	X99049
Cry19Aa1	Jeg65	Y08920
Cry20Aa1		U82518
Cry21Aa1		I32932
Cry22Aa1		I34547
Cyt1Aa1	CytA	X03182
Cyt1Aa2	CytA	X04338
Cyt1Aa3	CytA	Y00135
Cyt1Aa4	CytA	M35968
Cyt1Ab1	CytM	X98793
Cyt1Ba1		U37196
Cyt2Aa1	CytB	Z14147
Cyt2Ba1	"CytB"	U52043
Cyt2Ba2	"CytB"	AF020789
Cyt2Ba3	"CytB"	AF022884
Cyt2Ba4	"CytB"	AF022885
Cyt2Ba5	"CytB"	AF022886
Cyt2Bb1		U82519

^aAdapted from:

http://epunix.biols.susx.ac.uk/Home/Neil_Crickmore/Bt/index.html

1.2.3 BIOINSECTICIDE POLYPEPTIDE COMPOSITIONS

The utility of bacterial crystal proteins as insecticides was extended beyond lepidopterans and dipteran larvae when the first isolation of a coleopteran-toxic *B. thuringiensis* strain was reported (Krieg *et al.*, 1983; 1984). This strain
5 (described in U. S. Patent 4,766,203, specifically incorporated herein by reference), designated *B. thuringiensis* var. *tenebrionis*, is reported to be toxic to larvae of the coleopteran insects *Agelastica alni* (blue alder leaf beetle) and *Leptinotarsa decemlineata* (Colorado potato beetle).

U. S. Patent 5,024, 837 also describes hybrid *B. thuringiensis* var. *kurstaki*
10 strains which showed activity against lepidopteran insects. U. S. Patent 4,797,279 (corresponding to EP 0221024) discloses a hybrid *B. thuringiensis* containing a plasmid from *B. thuringiensis* var. *kurstaki* encoding a lepidopteran-toxic crystal protein-encoding gene and a plasmid from *B. thuringiensis tenebrionis* encoding a coleopteran-toxic crystal protein-encoding gene. The hybrid *B. thuringiensis* strain
15 produces crystal proteins characteristic of those made by both *B. thuringiensis kurstaki* and *B. thuringiensis tenebrionis*. U. S. Patent 4,910,016 (corresponding to EP 0303379) discloses a *B. thuringiensis* isolate identified as *B. thuringiensis* MT 104 which has insecticidal activity against coleopterans and lepidopterans.

20 1.2.4 MOLECULAR GENETIC TECHNIQUES FACILITATE PROTEIN ENGINEERING

The revolution in molecular genetics over the past decade has facilitated a logical and orderly approach to engineering proteins with improved properties. Site specific and random mutagenesis methods, the advent of polymerase chain reaction (PCR™) methodologies, and related advances in the field have permitted
25 an extensive collection of tools for changing both amino acid sequence, and underlying genetic sequences for a variety of proteins of commercial, medical, and agricultural interest.

Following the rapid increase in the number and types of crystal proteins which have been identified in the past decade, researchers began to theorize about
30 using such techniques to improve the insecticidal activity of various crystal proteins. In theory, improvements to δ -endotoxins should be possible using the meth-

ods available to protein engineers working in the art, and it was logical to assume that it would be possible to isolate improved variants of the wild-type crystal proteins isolated to date. By strengthening one or more of the aforementioned steps in the mode of action of the toxin, improved molecules should provide enhanced activity, and therefore, represent a breakthrough in the field. If specific amino acid residues on the protein are identified to be responsible for a specific step in the mode of action, then these residues can be targeted for mutagenesis to improve performance

10 1.2.5 STRUCTURAL ANALYSES OF CRYSTAL PROTEINS

The combination of structural analyses of *B. thuringiensis* toxins followed by an investigation of the function of such structures, motifs, and the like has taught that specific regions of crystal protein endotoxins are, in a general way, responsible for particular functions.

15 Domain 1, for example, from Cry3Bb and Cry1Ac has been found to be responsible for ion channel activity, the initial step in formation of a pore (Walters *et al.*, 1993; Von Tersch *et al.*, 1994). Domains 2 and 3 have been found to be responsible for receptor binding and insecticidal specificity (Aronson *et al.*, 1995; Caramori *et al.*, 1991; Chen *et al.*, 1993; de Maagd *et al.*, 1996; Ge *et al.*, 1991; Lee
20 *et al.*, 1992; Lee *et al.*, 1995; Lu *et al.*, 1994; Smedley and Ellar, 1996; Smith and Ellar, 1994; Rajamohan *et al.*, 1995; Rajamohan *et al.*, 1996; Wu and Dean, 1996). Regions in domain 2 and 3 can also impact the ion channel activity of some toxins (Chen *et al.*, 1993, Wolfersberger *et al.*, 1996; Von Tersch *et al.*, 1994).

25 1.3 DEFICIENCIES IN THE PRIOR ART

Unfortunately, while many laboratories have attempted to make mutated crystal proteins, few have succeeded in making mutated crystal proteins with improved lepidopteran toxicity. In almost all of the examples of genetically-engineered *B. thuringiensis* toxins in the literature, the biological activity of the
30 mutated crystal protein is no better than that of the wild-type protein, and in many cases, the activity is decreased or destroyed altogether (Almond and Dean, 1993;

Aronson *et al.*, 1995; Chen *et al.*, 1993, Chen *et al.*, 1995; Ge *et al.*, 1991; Kwak *et al.*, 1995; Lu *et al.*, 1994; Rajamohan *et al.*, 1995; Rajamohan *et al.*, 1996; Smedley and Ellar, 1996; Smith and Ellar, 1994; Wolfersberger *et al.*, 1996; Wu and Aronson, 1992).

5 For a crystal protein having approximately 650 amino acids in the sequence of its active toxin, and the possibility of 20 different amino acids at each position in this sequence, the likelihood of arbitrarily creating a successful new structure is remote, even if a general function to a stretch of 250-300 amino acids can be assigned. Indeed, the above prior art with respect to crystal protein gene mutagenesis
10 has been concerned primarily with studying the structure and function of the crystal proteins, using mutagenesis to perturb some step in the mode of action, rather than with engineering improved toxins.

Collectively, the limited successes in the art to develop synthetic toxins with improved insecticidal activity have stifled progress in this area and con-
15 founded the search for improved endotoxins or crystal proteins. Rather than following simple and predictable rules, the successful engineering of an improved crystal protein may involve different strategies, depending on the crystal protein being improved and the insect pests being targeted. Thus, the process is highly empirical.

20 Accordingly, traditional recombinant DNA technology is clearly not routine experimentation for providing improved insecticidal crystal proteins. What are lacking in the prior art are rational methods for producing genetically-engineered *B. thuringiensis* crystal proteins that have improved insecticidal activity and, in particular, improved toxicity towards a wide range of lepidopteran insect pests.

25

2.0 SUMMARY OF THE INVENTION

The present invention seeks to overcome these and other drawbacks inherent in the prior art by providing genetically-engineered modified *B. thuringiensis* δ -endotoxins (Cry*), and in particular modified Cry3 δ -endotoxins (designated
30 Cry3* endotoxins). Also provided are nucleic acid sequences comprising one or more genes which encode such modified proteins. Particularly preferred genes in-

clude *cry3** genes such as *cry3A**, *cry3B**, and *cry3C** genes, particularly *cry3B** genes, and more particularly, *cry3Bb** genes, that encode modified crystal proteins having improved insecticidal activity against target pests.

Also disclosed are novel methods for constructing synthetic Cry3* proteins, synthetically-modified nucleic acid sequences encoding such proteins, and compositions arising therefrom. Also provided are synthetic *cry3** expression vectors and various methods of using the improved genes and vectors. In a preferred embodiment, the invention discloses and claims Cry3B* proteins and *cry3B** genes which encode improved insecticidal polypeptides.

In preferred embodiments, channel-forming toxin design methods are disclosed which have been used to produce a specific set of designed Cry3Bb* toxins with improved biological activity. These improved Cry3Bb* proteins are listed in Table 2 along with their respective amino acid changes from wild-type (WT) Cry3Bb, the nucleotide changes present in the altered *cry3Bb** gene encoding the protein, the fold increase in bioactivity over WT Cry3Bb, the structural site of the alteration, and the design method(s) used to create the new toxins.

Accordingly, the present invention provides in an overall and general sense, mutagenized Cry3 protein-encoding genes and methods of making and using such genes. As used herein the term "mutagenized *cry3* gene(s)" means one or more *cry3* genes that have been mutagenized or altered to contain one or more nucleotide sequences which are not present in the wild type sequences, and which encode mutant Cry3 crystal proteins (Cry3*) showing improved insecticidal activity. Such mutagenized *cry3* genes have been referred to in the Specification as *cry3** genes. Exemplary *cry3** genes include *cry3A**, *cry3B**, and *cry3C** genes.

Exemplary mutagenized Cry3 protein-encoding genes include *cry3B* genes. As used herein the term "mutagenized *cry3B* gene(s)" means one or more genes that have been mutagenized or altered to contain one or more nucleotide sequences which are not present in the wild type sequences, and which encode mutant Cry3B crystal proteins (Cry3B*) showing improved insecticidal activity. Such genes have been designated *cry3B** genes. Exemplary *cry3B** genes include *cry3Ba** and *cry3Bb** genes, which encode Cry3Ba* and Cry3Bb* proteins, respectively.

Likewise, the present invention provides mutagenized Cry3A protein-encoding genes and methods of making and using such genes. As used herein the term "mutagenized *cry3A* gene(s)" means one or more genes that have been mutagenized or altered to contain one or more nucleotide sequences which are not
5 present in the wild type sequences, and which encode mutant Cry3A crystal proteins (Cry3A*) showing improved insecticidal activity. Such mutagenized genes have been designated as *cry3A** genes.

In similar fashion, the present invention provides mutagenized Cry3C protein-encoding genes and methods of making and using such genes. As used herein
10 the term "mutagenized *cry3C* gene(s)" means one or more genes that have been mutagenized or altered to contain one or more nucleotide sequences which are not present in the wild type sequences, and which encode mutant Cry3C crystal proteins (Cry3C*) showing improved insecticidal activity. Such mutagenized genes have been designated as *cry3C** genes.

15 Preferably the novel sequences comprise nucleic acid sequences in which at least one, and preferably, more than one, and most preferably, a significant number, of wild-type *cry3* nucleotides have been replaced with one or more nucleotides, or where one or more nucleotides have been added to or deleted from the native nucleotide sequence for the purpose of altering, adding, or deleting the corresponding amino acids encoded by the nucleic acid sequence so mutagenized. The
20 desired result, therefore, is alteration of the amino acid sequence of the encoded crystal protein to provide toxins having improved or altered activity and/or specificity compared to that of the unmodified crystal protein.

Examples of preferred Cry2Bb*-encoding genes include *cry3Bb.60*,
25 *cry3Bb.11221*, *cry3Bb.11222*, *cry3Bb.11223*, *cry3Bb.11224*, *cry3Bb.11225*,
cry3Bb.11226, *cry3Bb.11227*, *cry3Bb.11228*, *cry3Bb.11229*, *cry3Bb.11230*,
cry3Bb.11231, *cry3Bb.11232*, *cry3Bb.11233*, *cry3Bb.11234*, *cry3Bb.11235*,
cry3Bb.11236, *cry3Bb.11237*, *cry3Bb.11238*, *cry3Bb.11239*, *cry3Bb.11241*,
cry3Bb.11242, *cry3Bb.11032*, *cry3Bb.11035*, *cry3Bb.11036*, *cry3Bb.11046*,
30 *cry3Bb.11048*, *cry3Bb.11051*, *cry3Bb.11057*, *cry3Bb.11058*, *cry3Bb.11081*,
cry3Bb.11082, *cry3Bb.11083*, *cry3Bb.11084*, *cry3Bb.11095*, and *cry3Bb.11098*.

TABLE 2
CRY3Bb* PROTEINS EXHIBITING IMPROVED ACTIVITY AGAINST SCRW LARVAE

Cry3Bb*	<i>cry3Bb*</i>	<i>cry3Bb*</i> Nucleotide Sequence	Changes	Cry3Bb* Amino Acid Changes	Structural Site of Changes	Fold Increase Over WT Activity	Design Method Used
Cry3Bb.60	—	—	—	Δ 1-159	$\Delta\alpha 1-\alpha 3$	3.6×	1, 6, 8
Cry3Bb.11221	pEG1707	A460T, C461T, A462T, C464A, T465C, T466C, T467A, A468T, A469T, G470C, T472C, T473G, G474T, A477T, A478T, G479C		T154F, P155H, L156H, L158R	$\alpha 3, 4$	6.4×	1, 8
Cry3Bb.11222	pEG1708	T687C, T688C, A689T, C691A, A692G		Y230L, H231S	$\alpha 6$	4.0×	3, 7
Cry3Bb.11223	pEG1709	T667C, T687C, T688A, A689G, C691A, A692G		S223P, Y230S	$\alpha 6$	2.8×	3
Cry3Bb.11224	pEG1710	T687C, A692G		H231R	$\alpha 6$	5.0×	7, 8
Cry3Bb.11225	pEG1711	T687C, C691A		H231N, T241S	$\alpha 6$	3.6×	7
Cry3Bb.11226	pEG1712	T687C, C691A, A692C, T693C		H231T	$\alpha 6$	3.0×	7, 8
Cry3Bb.11227	pEG1713	C868A, G869A, G870T		R290N	$\alpha 7, \beta 1$	1.9×	2, 3, 4, 6

TABLE 2 (CONT'D)

Cry3Bb* Protein Designation	<i>cry3Bb</i> * Plasmid Designation	<i>cry3Bb</i> * Nucleotide Sequence Changes	Cry3Bb* Amino Acid Changes	Structural Site of Changes	Fold Increase Over WT Activity	Design Method Used
Cry3Bb.11228	pEG1714	C932T, A938C, T942G, G949A, T954C	S311L, N313T, E317K	Iβ1, α8	4.1×	2, 4
Cry3Bb.11229	pEG1715	T931A, A933C, T942A, T945A, G949A, A953G, T954C	S311T, E317K, Y318C	Iβ1, α8	2.5×	2, 4
Cry3Bb.11230	pEG1716	T931G, A933C, C934G, T945G, C946T, A947G, G951A, T954C	S311A, L312V, Q316W	Iβ1, α8	4.7×	2, 4 8
Cry3Bb.11231	pEG1717	T687C, A692G, C932T, A938C, T942G, G949A, T954C	H231R, S311L, N313T, E317K	α6; Iβ1, α8	7.9×	2, 4, 7, 8, 10
Cry3Bb.11232	pEG1718	T931A, A933G, T935C, T936A, A938C, T939C, T942C, T945A, G951T, T954C	S311T, L312P, N313T, E317N	Iβ1, α8	5.1×	4
Cry3Bb.11233	pEG1719	T931G, A933C, T936G, T942C, C943T, T945A, C946G, G948C, T954C	S311A, Q316D	Iβ1, α8	2.2×	2, 4

TABLE 2 (CONT'D)

Cry3Bb* Protein Designation	<i>cry3Bb</i> * Plasmid Designation	<i>cry3Bb</i> * Nucleotide Sequence Changes	Cry3Bb* Amino Acid Changes	Structural Site of Changes	Fold Increase Over WT Activity	Design Method Used
Cry3Bb.11234	pEG1720	T861C, T866C, C868A, T871C, T872G, A875T, T877A, C878G, A882G	I289T, L291R, Y292F, S293R	I α 7, β 1	4.1 \times	4
Cry3Bb.11235	pEG1721	T687C, A692G, C932T	H231R, S311L	α 6; I β 1, α 8	3.2 \times	2, 4, 7, 8, 10
Cry3Bb.11236	pEG1722	T931A, C932T, A933C, T936C, T942G, T945A, T954C	S311I	I β 1, α 8	3.1 \times	2, 4
Cry3Bb.11237	pEG1723	T931A, C932T, A933C, T936C, A937G, A938T, C941A, T942C, T945A, C946A, A947T, A950T, T954C	S311I, N313H	I β 1, α 8	5.4 \times	2, 4
Cry3Bb.11238	pEG1724	A933C, T936C, A937G, A938T, C941A, T942C, T945A, C946A, A947T, A950T, T954C	N313V, T314N, Q316M, E317V	I β 1, α 8	2.6 \times	2, 4

TABLE 2 (CONT'D)

Cry3Bb* Protein Designation	<i>cry3Bb</i> * Plasmid Designation	<i>cry3Bb</i> * Nucleotide Sequence Changes	Cry3Bb* Amino Acid Changes	Structural Site of Changes	Fold Increase Over WT Activity	Design Method Used
Cry3Bb.11239	pEG1725	A933T, A938G, T939G, T942A, T944C, T945A, A947T, G948T, A950C, T954C	N313R, L315P, Q316L, E317A	I β 1, α 8	2.8×	2, 4
Cry3Bb.11241	pEG1726	A860T, T861C, G862A, C868T, G869T, T871C, A873T, T877A, C878G, A879T	Y287F, D288N, R290L	I α 7, β 1	2.6×	2, 3, 4, 6
Cry3Bb.11242	pEG1727	C868G, G869T	R290V	I α 7, β 1	2.5×	2, 3, 4, 6, 8
Cry3Bb.11032	pEG1041	A494G	D165G	α 4	3.1×	2, 4, 8
Cry3Bb.11035	pEG1046	G479A, A481C, A482C, A484C, G485A, A486C, A494G	S160N, K161P, P162H, D165G	α 4	2.7×	8
Cry3Bb.11036	pEG1047	A865G, T877C	I289V, S293P	I α 7, β 1	4.3×	4
Cry3Bb.11046	pEG1052	G479A, A481C, A482C, A484C, G485A, A486C, A494G, A865G, T877C	S160N, K161P, P162H, D165G, I289V, S293P	α 4; I α 7, β 1	2.6×	2, 4, 8, 10

TABLE 2 (CONT'D)

Cry3Bb*	<i>cry3Bb</i> *	<i>cry3Bb</i> * Nucleotide Sequence	Cry3Bb* Amino Acid Changes	Structural Site of Changes	Fold Increase Over WT Activity	Design Method Used
Protein Designation	Plasmid Designation	Changes				
Cry3Bb.11048	pEG1054	T309A, Δ310, Δ311, Δ312	D103E, ΔA104	lα2a,2b	4.3×	8
Cry3Bb.11051	pEG1057	A565G, A566G	K189G	lα4,5	3.0×	2, 3, 4
Cry3Bb.11057	pEG1062	T309A, Δ310, Δ311, Δ312, G479A, A481C, A482C, A484C, G485A, A486C, A494G	D103E, ΔA104, S160N, K161P, P162H, D165G	lα2a,2b; α4	3.4×	2, 4, 8, 10
Cry3Bb.11058	pEG1063	T309A, Δ310, Δ311, Δ312, A460T, C461T, A462T, C464A, T465C, T466C, T467A, A468T, A469T, G470C, T472C, T473G, G474T, A477T, A478T, G479C	D103E, ΔA104, T154F, P155H, L156H, L158R	lα2a,2b; lα3,4	3.5×	1, 8, 10
Cry3Bb.11081	pEG1084	A494G, T931A, A933C, T942A, T945A, G949A, T954C	D165G, S311T, E317K	α4; lβ1,α8	6.1×	2, 4, 8, 10

TABLE 2 (CONT'D)

Cry3Bb*	<i>cry3Bb</i> *	<i>cry3Bb</i> * Nucleotide Sequence	Cry3Bb* Amino Acid Changes	Structural Site of Changes	Fold Increase Over WT Activity	Design Method Used
Protein	Plasmid	Changes				
Designation	Designation					
Cry3Bb.11082	pEG1085	A494G, A865G, T877C, T914C, T931G, A933C, C934G, T945G, C946T, A947G, G951A, T954C, A1043G, T1094C	D165G, I289V, S293P, F305S, S311A, L312V, Q316W, Q348R, V365A	α 4; 1α 7, β 1; β 1; β 1, α 8; β 2; β 3b	4.9×	2, 4, 5, 8, 9, 10
Cry3Bb.11083	pEG1086	A865G, T877C, A1043G	I289V, S293P, Q348R	1α 7, β 1; β 2	7.4×	4, 5, 9, 10
Cry3Bb.11084	pEG1087	A494G, C932T	D165G, S311L	α 4; β 1, α 8	7.2×	2, 4, 8, 10
Cry3Bb.11095	pEG1095	A1043G	Q348R	β 2	4.6×	5, 9
Cry3Bb.11098	pEG1098	A494G, T687C, A692G, C932T, A938C, T942G, G949A, T954C	D165G, H231R, S311L, N313T, E317K	α 4; α 6, 1β 1, α 8	7.9×	2, 4, 7, 8

In a variety of illustrative embodiments, the inventors have shown remarkable success in generating toxins with improved insecticidal activity using these methods. In particular, the inventors have identified unique methods of analyzing and designing toxins having improved or enhanced insecticidal properties both *in vitro* and *in vivo*.

In addition to modifications of Cry3Bb peptides, those having benefit of the present teaching are now also able to make mutations in a variety of channel-forming toxins, and particularly in crystal proteins which are related to Cry3Bb either functionally or structurally. In fact, the inventors contemplate that any *B. thuringiensis* crystal protein or peptide can be analyzed using the methods disclosed herein and may be altered using the methods disclosed herein to produce crystal proteins having improved insecticidal specificity or activity. Alternatively, the inventors contemplate that those of skill in the art having the benefit of the teachings disclosed herein will be able to prepare not only mutated Cry3 toxins with improved activity, but also other crystal proteins including all of those proteins identified in Table 1, herein. In particular, the inventors contemplate the creation of Cry3* variants using one or more of the methods disclosed herein to produce toxins with improved activity. For example, the inventors note Cry3A, Cry3B, and Cry3C crystal proteins (which are known in the art) may be modified using one or more of the design strategies employed herein, to prepare synthetically-modified crystal proteins with improved properties. Likewise, one of skill in the art will even be able to utilize the teachings of the present disclosure to modify other channel forming toxins, including channel forming toxins other than *B. thuringiensis* crystal proteins, and even to modify proteins and channel toxins not yet described or characterized.

Because the structures for insecticidal crystal proteins show a remarkable conservation of protein tertiary structure (Grochulski *et al.*, 1995), and because many crystal proteins show significant amino acid sequence identity to the Cry3Bb amino acid sequence within domain 1, including proteins of the Cry1, Cry2, Cry3, Cry4, Cry5, Cry7, Cry8, Cry9, Cry10, Cry11, Cry12, Cry13, Cry14, and Cry16 classes (Table 1), now in light of the inventors' surprising discovery, for the first time, those of skill in the art having benefit of the teachings disclosed herein will

be able to broadly apply the methods of the invention to modifying a host of crystal proteins with improved activity or altered specificity. Such methods will not only be limited to the insecticidal crystal proteins disclosed in Table 1, but may also been applied to any other related crystal protein, including those yet to be identified.

In particular, the high degree of homology between Cry3A, Cry3B, and Cry3C proteins is evident in the alignment of the primary amino acid sequence of the three proteins (FIG. 17A, FIG. 17B, and FIG. 17C).

As such, the disclosed methods may be now applied to preparation of modified crystal proteins having one or more alterations introduced using one or more of the mutational design methods as disclosed herein. The inventors further contemplate that regions may be identified in one or more domains of a crystal protein, or other channel forming toxin which may be similarly modified through site-specific or random mutagenesis to generate toxins having improved activity, or alternatively, altered specificity.

In certain applications, the creation of altered toxins having increased activity against one or more insects is desired. Alternatively, it may be desirable to utilize the methods described herein for creating and identifying altered insecticidal crystal proteins which are active against a wider spectrum of susceptible insects. The inventors further contemplate that the creation of chimeric insecticidal crystal proteins comprising one or more of these mutations may be desirable for preparing "super" toxins which have the combined advantages of increased insecticidal activity and concomitant broad spectrum activity.

In light of the present disclosure, the mutagenesis of one or more codons within the sequence of a toxin may result in the generation of a host of related insecticidal proteins having improved activity. While exemplary mutations have been described for each of the design strategies employed in the present invention, the inventors contemplate that mutations may also be made in insecticidal crystal proteins, including the loop regions, helices regions, active sites of the toxins, regions involved in protein oligomerization, and the like, which will give rise to

functional bioinsecticidal crystal proteins. All such mutations are considered to fall within the scope of this disclosure.

In one illustrative embodiment, mutagenized *cry3Bb** genes are obtained which encode Cry3Bb* variants that are generally based upon the wild-type
5 Cry3Bb sequence, but that have one or more changes incorporated into the amino acid sequence of the protein using one or more of the design strategies described and claimed herein.

In these and other embodiments, the mutated genes encoding the crystal proteins may be modified so as to change about one, two, three, four, or five or so
10 amino acids in the primary sequence of the encoded polypeptide. Alternatively even more changes from the native sequence may be introduced, such that the encoded protein may have at least about 1% or 2%, or alternatively about 3% or about 4%, or even about 5% to about 10%, or about 10% to about 15%, or even about 15% to about 20% or more of the codons either altered, deleted, or otherwise
15 modified. In certain situations, it may even be desirable to alter substantially more of the primary amino acid sequence to obtain the desired modified protein. In such cases the inventors contemplate that from about 25%, to about 50%, or even from about 50% to about 75%, or more of the native (or wild-type) codons either altered, deleted, or otherwise modified. Alternatively, mutations in the amino acid se-
20 quences or underlying DNA gene sequences which result in the insertion or deletion of one or more amino acids within one or more regions of the crystal protein or peptide.

To effect such changes in the primary sequence of the encoded polypeptides, it may be desirable to mutate or delete one or more nucleotides from the nu-
25 cleic acid sequences of the genes encoding such polypeptides, or alternatively, under certain circumstances to add one or more nucleotides into the primary nucleic acid sequence at one or more sites in the sequence. Frequently, several nucleotide residues may be altered to produce the desired polypeptide. As such, the inventors contemplate that in certain embodiments it may be desirable to alter only one, two,
30 three, four, or five or so nucleotides in the primary sequence. In other embodiments, which more changes are desired, the mutagenesis may involve changing,

deleting, or inserting 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or even 20 or so nucleotide residues in the gene sequence. In still other embodiments, one may desire to mutate, delete, or insert 21, 22, 23, 24, 25, 26, 27, 28, 29, 30-40, 40-50, 50-60, 60-70, 70-80, 80-90, or even 90-100, 150, 200, 250, 300, 350, 400, 450, or
5 more nucleotides in the sequence of the gene in order to prepare a *cry3** gene which produces a Cry3* polypeptide having the desired characteristics. In fact, any number of mutations, deletions, and/or insertions may be made in the primary sequence of the gene, so long as the encoded protein has the improved insecticidal activity or specificity characteristics described herein.

10 Changing a large number of the codons in the nucleotide sequence of an endotoxin-encoding gene may be particularly desirable and often necessary to achieve the desired results, particularly in the situation of "plantizing" a DNA sequence in order to express a DNA of non-plant origin in a transformed plant cell. Such methods are routine to those of skill in the plant genetics arts, and frequently
15 many residues of a primary gene sequence will be altered to facilitate expression of the gene in the plant cell. Preferably, the changes in the gene sequence introduce no changes in the amino acid sequence, or introduce only conservative replacements in the amino acid sequence such that the polypeptide produced in the plant cell from the "plantized" nucleotide sequence is still fully functional, and has the
20 desired qualities when expressed in the plant cell.

Genes and encoded proteins mutated in the manner of the invention may also be operatively linked to other protein-encoding nucleic acid sequences, or expressed as fusion proteins. Both N-terminal and C-terminal fusion proteins are contemplated. Virtually any protein- or peptide-encoding DNA sequence, or
25 combinations thereof, may be fused to a mutated *cry3** sequence in order to encode a fusion protein. This includes DNA sequences that encode targeting peptides, proteins for recombinant expression, proteins to which one or more targeting peptides is attached, protein subunits, domains from one or more crystal proteins, and the like. Such modifications to primary nucleotide sequences to enhance, target, or
30 optimize expression of the gene sequence in a particular host cell, tissue, or cellular localization, are well-known to those of skill in the art of protein engineering and

molecular biology, and it will be readily apparent to such artisans, having benefit of the teachings of this specification, how to facilitate such changes in the nucleotide sequence to produce the polypeptides and polynucleotides disclosed herein.

In one aspect, the invention discloses and claims host cells comprising one or more of the modified crystal proteins disclosed herein, and in particular, cells of *B. thuringiensis* strains EG11221, EG11222, EG11223, EG11224, EG11225, EG11226, EG11227, EG11228, EG11229, EG11230, EG11231, EG11232, EG11233, EG11234, EG11235, EG11236, EG11237, EG11238, EG11239, EG11241, EG11242, EG11032, EG11035, EG11036, EG11046, EG11048, EG11051, EG11057, EG11058, EG11081, EG11082, EG11083, EG11084, EG11095, and EG11098 which comprise recombinant DNA segments encoding synthetically-modified Cry3Bb* crystal proteins which demonstrates improved insecticidal activity.

Likewise, the invention also discloses and claims cell cultures of *B. thuringiensis* EG11221, EG11222, EG11223, EG11224, EG11225, EG11226, EG11227, EG11228, EG11229, EG11230, EG11231, EG11232, EG11233, EG11234, EG11235, EG11236, EG11237, EG11238, EG11239, EG11241, EG11242, EG11032, EG11035, EG11036, EG11046, EG11048, EG11051, EG11057, EG11058, EG11081, EG11082, EG11083, EG11084, and EG11095, and 11098.

Such cell cultures may be biologically-pure cultures consisting of a single strain, or alternatively may be cell co-cultures consisting of one or more strains. Such cell cultures may be cultivated under conditions in which one or more additional *B. thuringiensis* or other bacterial strains are simultaneously co-cultured with one or more of the disclosed cultures, or alternatively, one or more of the cell cultures of the present invention may be combined with one or more additional *B. thuringiensis* or other bacterial strains following the independent culture of each. Such procedures may be useful when suspensions of cells containing two or more different crystal proteins are desired.

The subject cultures have been deposited under conditions that assure that access to the cultures will be available during the pendency of this patent applica-

tion to one determined by the Commissioner of Patents and Trademarks to be entitled thereto under 37 C.F.R. §1.14 and 35 U.S.C. §122. The deposits are available as required by foreign patent laws in countries wherein counterparts of the subject application, or its progeny, are filed. However, it should be understood that the availability of a deposit does not constitute a license to practice the subject invention in derogation of patent rights granted by governmental action.

Further, the subject culture deposits will be stored and made available to the public in accord with the provisions of the Budapest Treaty for the Deposit of Microorganisms, *i.e.*, they will be stored with all the care necessary to keep them viable and uncontaminated for a period of at least five years after the most recent request for the finishing of a sample of the deposit, and in any case, for a period of at least 30 (thirty) years after the date of deposit or for the enforceable life of any patent which may issue disclosing the cultures. The depositor acknowledges the duty to replace the deposits should the depository be unable to furnish a sample when requested, due to the condition of the deposits. All restrictions on the availability to the public of the subject culture deposits will be irrevocably removed upon the granting of a patent disclosing them.

Cultures shown in Table 3 were deposited in the permanent collection of the Agricultural Research Service Culture Collection, Northern Regional Research Laboratory (NRRL) under the terms of the Budapest Treaty.

TABLE 3
STRAINS OF THE PRESENT INVENTION DEPOSITED UNDER THE TERMS
OF THE BUDAPEST TREATY

Strain	Deposit Date	Protein	Accession Number (NRRL Number)
EG11032	5/27/97	Cry3Bb.11032	B-21744
EG11035	5/27/97	Cry3Bb.11035	B-21745
EG11036	5/27/97	Cry3Bb.11036	B-21746
EG11037	5/27/97	Cry3Bb.11037	B-21747
EG11046	5/27/97	Cry3Bb.11046	B-21748

TABLE 3 (CONT'D)

Strain	Deposit Date	Protein	Accession Number (NRRL Number)
EG11048	5/27/97	Cry3Bb.11048	B-21749
EG11051	5/27/97	Cry3Bb.11051	B-21750
EG11057	5/27/97	Cry3Bb.11057	B-21751
EG11058	5/27/97	Cry3Bb.11058	B-21752
EG11081	5/27/97	Cry3Bb.11081	B-21753
EG11082	5/27/97	Cry3Bb.11082	B-21754
EG11083	5/27/97	Cry3Bb.11083	B-21755
EG11084	5/27/97	Cry3Bb.11084	B-21756
EG11095	5/27/97	Cry3Bb.11095	B-21757
EG11204	5/27/97	Cry3Bb.11204	B-21758
EG11221	5/27/97	Cry3Bb.11221	B-21759
EG11222	5/27/97	Cry3Bb.11222	B-21760
EG11223	5/27/97	Cry3Bb.11223	B-21761
EG11224	5/27/97	Cry3Bb.11224	B-21762
EG11225	5/27/97	Cry3Bb.11225	B-21763
EG11226	5/27/97	Cry3Bb.11226	B-21764
EG11227	5/27/97	Cry3Bb.11227	B-12765
EG11228	5/27/97	Cry3Bb.11228	B-12766
EG11229	5/27/97	Cry3Bb.11229	B-21767
EG11230	5/27/97	Cry3Bb.11230	B-21768
EG11231	5/27/97	Cry3Bb.11231	B-21769
EG11232	5/27/97	Cry3Bb.11232	B-12770
EG11233	5/27/97	Cry3Bb.11233	B-21771
EG11234	5/27/97	Cry3Bb.11234	B-21772
EG11235	5/27/97	Cry3Bb.11235	B-21773
EG11236	5/27/97	Cry3Bb.11236	B-21774
EG11237	5/27/97	Cry3Bb.11237	B-21775
EG11238	5/27/97	Cry3Bb.11238	B-21776

TABLE 3 (CONT'D)

Strain	Deposit Date	Protein	Accession Number (NRRL Number)
EG11239	5/27/97	Cry3Bb.11239	B-21777
EG11241	5/27/97	Cry3Bb.11241	B-21778
EG11242	5/27/97	Cry3Bb.11242	B-21779

Also disclosed are methods of controlling or eradicating an insect population from an environment. Such methods generally comprise contacting the insect population to be controlled or eradicated with an insecticidally-effective amount of a Cry3* crystal protein composition. Preferred Cry3* compositions include Cry3A*, Cry3B*, and Cry3C* polypeptide compositions, with Cry3B* compositions being particularly preferred. Examples of such polypeptides include proteins selected from the group consisting of Cry3Bb-60, Cry3Bb.11221, Cry3Bb.11222, Cry3Bb.11223, Cry3Bb.11224, Cry3Bb.11225, Cry3Bb.11226, Cry3Bb.11227, Cry3Bb.11228, Cry3Bb.11229, Cry3Bb.11230, Cry3Bb.11231, Cry3Bb.11232, Cry3Bb.11233, Cry3Bb.11234, Cry3Bb.11235, Cry3Bb.11236, Cry3Bb.11237, Cry3Bb.11238, Cry3Bb.11239, Cry3Bb.11241, Cry3Bb.11242, Cry3Bb.11032, Cry3Bb.11035, Cry3Bb.11036, Cry3Bb.11046, Cry3Bb.11048, Cry3Bb.11051, Cry3Bb.11057, Cry3Bb.11058, Cry3Bb.11081, Cry3Bb.11082, Cry3Bb.11083, Cry3Bb.11084, Cry3Bb.11095, and Cry3Bb.11098.

In preferred embodiments, these Cry3Bb* crystal protein compositions comprise the amino acid sequence of any of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, SEQ ID NO:102 or SEQ ID NO:108.

2.1 METHODS FOR PRODUCING MODIFIED CRY* PROTEINS

The modified Cry* polypeptides of the present invention are preparable by a process which generally involves the steps of obtaining a nucleic acid sequence
5 encoding a Cry* polypeptide; analyzing the structure of the polypeptide to identify particular "target" sites for mutagenesis of the underlying gene sequence; introducing one or more mutations into the nucleic acid sequence to produce a change in one or more amino acid residues in the encoded polypeptide sequence; and expressing in a transformed host cell the mutagenized nucleic acid sequence under
10 conditions effective to obtain the modified Cry* protein encoded by the *cry** gene.

Means for obtaining the crystal structures of the polypeptides of the invention are well-known. Exemplary high resolution crystal structure solution sets are given in Section 9.0 of the disclosure, and include the crystal structure of both the Cry3A and Cry3B polypeptides disclosed herein. The information provided in
15 Section 9.0 permits the analyses disclosed in each of the methods herein which rely on the 3D crystal structure information for targeting mutagenesis of the polypeptides to particular regions of the primary amino acid sequences of the δ -endotoxins to obtain mutants with increased insecticidal activity or enhanced insecticidal specificity.

20 A first method for producing a modified *B. thuringiensis* Cry3Bb δ -endotoxin having improved insecticidal activity or specificity disclosed herein generally involves obtaining a high-resolution 3D crystal structure of the endotoxin, locating in the crystal structure one or more regions of bound water wherein the bound water forms a contiguous hydrated surfaces separated by no more than
25 about 16 Å; increasing the number of water molecules in this surface by increasing the hydrophobicity of one or more amino acids of the protein in the region; and obtaining the modified δ -endotoxin so produced. Exemplary δ -endotoxins include Cry3Bb.11032, Cry3Bb.11227, Cry3Bb.11241, Cry3Bb.11051, Cry3Bb.11242, and Cry3Bb.11098.

30 A second method for producing a modified *B. thuringiensis* Cry3Bb δ -endotoxin having improved insecticidal activity comprises identifying a loop re-

gion in a δ -endotoxin; modifying one or more amino acids in the loop to increase the hydrophobicity of the amino acids; and obtaining the modified δ -endotoxin so produced. Preferred δ -endotoxin produced by this method include Cry3Bb.11241, Cry3Bb.11242, Cry3Bb.11228, Cry3Bb.11229, Cry3Bb.11230, Cry3Bb.11231, 5 Cry3Bb.11233, Cry3Bb.11236, Cry3Bb.11237, Cry3Bb.11238, and Cry3Bb.11239.

A method for increasing the mobility of channel forming helices of a *B. thuringiensis* Cry3B δ -endotoxin is also provided by the present invention. The method generally comprises disrupting one or more hydrogen bonds formed between a first amino acid of one or more of the channel forming helices and a second amino acid of the δ -endotoxin. The hydrogen bonds may be formed inter- or intramolecularly, and the disrupting may consist of replacing a first or second amino acid with a third amino acid whose spatial distance is greater than about 3 Å, or whose spatial orientation bond angle is not equal to 180 ± 60 degrees relative to the hydrogen bonding site of the first or second amino acid. δ -endotoxins produced by this method and disclosed herein include Cry3Bb.11222, Cry3Bb.11223, Cry3Bb.11224, Cry3Bb.11225, Cry3Bb.11226, Cry3Bb.11227, Cry3Bb.11231, Cry3Bb.11241, and Cry3Bb.11242, and Cry3Bb.11098. 15

Also disclosed is a method of increasing the flexibility of a loop region in a channel forming domain of a *B. thuringiensis* Cry3Bb δ -endotoxin. This method comprises obtaining a crystal structure of a Cry3Bb δ -endotoxin having one or more loop regions; identifying the amino acids comprising the loop region; and altering one or more of the amino acids to reduce steric hindrance in the loop region, wherein the altering increases flexibility of the loop region in the δ -endotoxin. Examples of δ -endotoxins produced using this method include Cry3Bb.11032, Cry3Bb.11051, Cry3Bb.11228, Cry3Bb.11229, Cry3Bb.11230, Cry3Bb.11231, Cry3Bb.11232, Cry3Bb.11233, Cry3Bb.11236, Cry3Bb.11237, Cry3Bb.11238, Cry3Bb.11239, Cry3Bb.11227, Cry3Bb.11234, Cry3Bb.11241, Cry3Bb.11242, Cry3Bb.11036, and Cry3Bb.11098. 25

Another aspect of the invention is a method for increasing the activity of a δ -endotoxin, comprising reducing or eliminating binding of the δ -endotoxin to a 30

carbohydrate in a target insect gut. The eliminating or reducing may be accomplished by removal of one or more α helices of domain 1 of the δ -endotoxin, for example, by removal of α helices $\alpha 1$, $\alpha 2a/b$, and $\alpha 3$. An exemplary δ -endotoxin produced using the method is Cry3Bb.60.

5 Alternatively, the reducing or eliminating may be accomplished by replacing one or more amino acids within loop $\beta 1, \alpha 8$, with one or more amino acids having increased hydrophobicity. Such a method gives rise to δ -endotoxins such as Cry3Bb.11228, Cry3Bb.11230, Cry3B.11231, Cry3Bb.11237, and Cry3Bb.11098, which are described in detail, herein.

10 Alternatively, the reducing or eliminating is accomplished by replacing one or more specific amino acids, with any other amino acid. Such replacements are described in Table 2, and in the examples herein. One example is the δ -endotoxin designated herein as Cry3Bb.11221.

 A method of identifying a region of a Cry3Bb δ -endotoxin for targeted
15 mutagenesis comprising: obtaining a crystal structure of the δ -endotoxin; identifying from the crystal structure one or more surface-exposed amino acids in the protein; randomly substituting one or more of the surface-exposed amino acids to obtain a plurality of mutated polypeptides, wherein at least 50% of the mutated polypeptides have diminished insecticidal activity; and identifying from the plural-
20 ity of mutated polypeptides one or more regions of the Cry3Bb δ -endotoxin for targeted mutagenesis. The method may further comprise determining the amino acid sequences of a plurality of mutated polypeptides having diminished activity, and identifying one or more amino acid residues required for insecticidal activity.

 In another embodiment, the invention provides a process for producing a
25 Cry3Bb δ -endotoxin having improved insecticidal activity. The process generally involves the steps of obtaining a high-resolution crystal structure of the protein; determining the electrostatic surface distribution of the protein; identifying one or more regions of high electrostatic diversity; modifying the electrostatic diversity of the region by altering one or more amino acids in the region; and obtaining a
30 Cry3Bb δ -endotoxin which has improved insecticidal activity. In one embodiment, the electrostatic diversity may be decreased relative to the electrostatic diversity of

a native Cry3Bb δ -endotoxin. Exemplary δ -endotoxins with decreased electrostatic diversity include Cry3Bb.11227, Cry3Bb.11241, and Cry3Bb.11242. Alternatively, the electrostatic diversity may be increased relative to the electrostatic diversity of a native Cry3Bb δ -endotoxin. An exemplary δ -endotoxin with increased electrostatic diversity is Cry3Bb.11234.

Furthermore, the invention also provides a method of producing a Cry3Bb δ -endotoxin having improved insecticidal activity which involves obtaining a high-resolution crystal structure; identifying the presence of one or more metal binding sites in the protein; altering one or more amino acids in the binding site; and obtaining an altered protein, wherein the protein has improved insecticidal activity. The altering may involve the elimination of one or more metal binding sites. Exemplary δ -endotoxins include Cry3Bb.11222, Cry3Bb.11224, Cry3Bb.11225, and Cry3Bb.11226.

A further aspect of the invention involves a method of identifying a *B. thuringiensis* Cry3Bb δ -endotoxin having improved channel activity. This method in an overall sense involves obtaining a Cry3Bb δ -endotoxin suspected of having improved channel activity; and determining one or more of the following characteristics in the δ -endotoxin, and comparing such characteristics to those obtained for the wild-type unmodified δ -endotoxin: (1) the rate of channel formation, (2) the rate of growth of channel conductance or (3) the duration of open channel state. From this comparison, one may then select a δ -endotoxin which has an increased rate of channel formation compared to the wildtype δ -endotoxin. Examples of Cry3Bb δ -endotoxins prepared by this method include Cry3Bb.60, Cry3Bb.11035, Cry3Bb.11048, Cry3Bb.11032, Cry3Bb.11223, Cry3Bb.11224, Cry3Bb.11226, Cry3Bb.11221, Cry3Bb.11242, Cry3Bb.11230, and Cry3Bb.11098.

Also provided is a method for producing a modified Cry3Bb δ -endotoxin, having improved insecticidal activity which involves altering one or more non-surface amino acids located at or near the point of greatest convergence of two or more loop regions of the Cry3Bb δ -endotoxin, such that the altering decreases the mobility of one or more of the loop regions. The mobility may conveniently be

determined by comparing the thermal denaturation of the modified protein to a wild-type Cry3Bb δ -endotoxin. An exemplary crystal protein produced by this method is Cry3Bb.11095.

A further aspect of the invention involves a method for preparing a modified Cry3Bb δ -endotoxin, having improved insecticidal activity comprising modifying one or more amino acids in the loop to increase the hydrophobicity of said amino acids; and altering one or more of said amino acids to reduce steric hindrance in the loop region, wherein the altering increases flexibility of the loop region in the endotoxin. Exemplary Cry3Bb δ -endotoxins produced is selected from the group consisting of Cry3Bb.11057, Cry3Bb.11058, Cry3Bb.11081, Cry3Bb.11082, Cry3Bb.11083, Cry3Bb.11084, Cry3Bb.11231, Cry3Bb.11235, and Cry3Bb.11098.

The invention also provides a method of improving the insecticidal activity of a *B. thuringiensis* Cry3Bb δ -endotoxin, which generally comprises inserting one or more protease sensitive sites into one or more loop regions of domain 1 of the δ -endotoxin. Preferably, the loop region is α 3,4, and an exemplary δ -endotoxin so produced is Cry3Bb.11221.

2.2 POLYPEPTIDE COMPOSITIONS

The crystal proteins so produced by each of the methods described herein also represent important aspects of the invention. Such crystal proteins preferably include a protein or peptide selected from the group consisting of Cry3Bb-60, Cry3Bb.11221, Cry3Bb.11222, Cry3Bb.11223, Cry3Bb.11224, Cry3Bb.11225, Cry3Bb.11226, Cry3Bb.11227, Cry3Bb.11228, Cry3Bb.11229, Cry3Bb.11230, Cry3Bb.11231, Cry3Bb.11232, Cry3Bb.11233, Cry3Bb.11234, Cry3Bb.11235, Cry3Bb.11236, Cry3Bb.11237, Cry3Bb.11238, Cry3Bb.11239, Cry3Bb.11241, Cry3Bb.11242, Cry3Bb.11032, Cry3Bb.11035, Cry3Bb.11036, Cry3Bb.11046, Cry3Bb.11048, Cry3Bb.11051, Cry3Bb.11057, Cry3Bb.11058, Cry3Bb.11081, Cry3Bb.11082, Cry3Bb.11083, Cry3Bb.11084, Cry3Bb.11095, and Cry3Bb.11098.

In preferred embodiments, the protein comprises a contiguous amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, SEQ ID NO:102, and SEQ ID NO:108.

Highly preferred are those crystal proteins which are encoded by the nucleic acid sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:99, SEQ ID NO:101; or SEQ ID NO:107, or a nucleic acid sequence which hybridizes to the nucleic acid sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:99, SEQ ID NO:101, or SEQ ID NO:107 under conditions of moderate stringency.

Amino acid, peptide and protein sequences within the scope of the present invention include, and are not limited to the sequences set forth in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12,

SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22
SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32,
SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42,
SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52,
5 SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62,
SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100,
SEQ ID NO:102, and SEQ ID NO:108, and alterations in the amino acid sequences
including alterations, deletions, mutations, and homologs.

Compositions which comprise from about 0.5% to about 99% by weight of
10 the crystal protein, or more preferably from about 5% to about 75%, or from about
25% to about 50% by weight of the crystal protein are provided herein. Such com-
positions may readily be prepared using techniques of protein production and puri-
fication well-known to those of skill, and the methods disclosed herein. Such a
process for preparing a Cry3Bb* crystal protein generally involves the steps of
15 culturing a host cell which expresses the Cry3Bb* protein (such as a
B. thuringiensis EG11221, EG11222, EG11223, EG11224, EG11225, EG11226,
EG11227, EG11228, EG11229, EG11230, EG11231, EG11232, EG11233,
EG11234, EG11235, EG11236, EG11237, EG11238, EG11239, EG11241,
EG11242, EG11032, EG11035, EG11036, EG11046, EG11048, EG11051,
20 EG11057, EG11058, EG11081, EG11082, EG11083, EG11084, EG11095, or
EG11098 cell) under conditions effective to produce the crystal protein, and then
obtaining the crystal protein so produced.

The protein may be present within intact cells, and as such, no subsequent
protein isolation or purification steps may be required. Alternatively, the cells may
25 be broken, sonicated, lysed, disrupted, or plasmolyzed to free the crystal protein(s)
from the remaining cell debris. In such cases, one may desire to isolate, concen-
trate, or further purify the resulting crystals containing the proteins prior to use,
such as, for example, in the formulation of insecticidal compositions. The com-
position may ultimately be purified to consist almost entirely of the pure protein, or
30 alternatively, be purified or isolated to a degree such that the composition com-
prises the crystal protein(s) in an amount of from between about 0.5% and about

99% by weight, or in an amount of from between about 5% and about 95% by weight, or in an amount of from between about 15% and about 85% by weight, or in an amount of from between about 25% and about 75% by weight, or in an amount of from between about 40% and about 60% by weight *etc.*

5

2.3 RECOMBINANT VECTORS EXPRESSING *CRY3** GENES

One important embodiment of the invention is a recombinant vector which comprises a nucleic acid segment encoding one or more of the novel *B. thuringiensis* crystal proteins disclosed herein. Such a vector may be transferred
10 to and replicated in a prokaryotic or eukaryotic host, with bacterial cells being particularly preferred as prokaryotic hosts, and plant cells being particularly preferred as eukaryotic hosts.

In preferred embodiments, the recombinant vector comprises a nucleic acid segment encoding the amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, SEQ
15 ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID
20 NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:108. Highly preferred nucleic acid segments are those which have the sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID
25 NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID
30 NO:69, SEQ ID NO:99, SEQ ID NO:101, or SEQ ID NO:107.

Another important embodiment of the invention is a transformed host cell which expresses one or more of these recombinant vectors. The host cell may be either prokaryotic or eukaryotic, and particularly preferred host cells are those which express the nucleic acid segment(s) comprising the recombinant vector
5 which encode one or more *B. thuringiensis* crystal protein comprising modified amino acid sequences in one or more loop regions of domain 1, or between α helix 7 of domain 1 and β strand 1 of domain 2. Bacterial cells are particularly preferred as prokaryotic hosts, and plant cells are particularly preferred as eukaryotic hosts

In an important embodiment, the invention discloses and claims a host cell
10 wherein the modified amino acid sequences comprise one or more loop regions between α helices 1 and 2, α helices 2 and 3, α helices 3 and 4, α helices 4 and 5, α helices 5 and 6 or α helices 6 and 7 of domain 1, or between α helix 7 of domain 1 and β strand 1 of domain 2. A particularly preferred host cell is one that comprises the amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6,
15 SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56,
20 SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:108, and more preferably, one that comprises the nucleic acid sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID
25 NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID
30 NO:99, SEQ ID NO:101, or SEQ ID NO:107.

Bacterial host cells transformed with a nucleic acid segment encoding a modified Cry3Bb crystal protein according to the present invention are disclosed and claimed herein, and in particular, a *B. thuringiensis* cell having designation EG11221, EG11222, EG11223, EG11224, EG11225, EG11226, EG11227, EG11228, EG11229, EG11230, EG11231, EG11232, EG11233, EG11234, EG11235, EG11236, EG11237, EG11238, EG11239, EG11241, EG11242, EG11032, EG11035, EG11036, EG11046, EG11048, EG11051, EG11057, EG11058, EG11081, EG11082, EG11083, EG11084, EG11095, or EG11098.

In another embodiment, the invention encompasses a method of using a nucleic acid segment of the present invention that encodes a *cry3Bb** gene. The method generally comprises the steps of: (a) preparing a recombinant vector in which the *cry3Bb** gene is positioned under the control of a promoter; (b) introducing the recombinant vector into a host cell; (c) culturing the host cell under conditions effective to allow expression of the Cry3Bb* crystal protein encoded by said *cry3Bb** gene; and (d) obtaining the expressed Cry3Bb* crystal protein or peptide.

A wide variety of ways are available for introducing a *B. thuringiensis* gene expressing a toxin into the microorganism host under conditions which allow for stable maintenance and expression of the gene. One can provide for DNA constructs which include the transcriptional and translational regulatory signals for expression of the toxin gene, the toxin gene under their regulatory control and a DNA sequence homologous with a sequence in the host organism, whereby integration will occur, and/or a replication system which is functional in the host, whereby integration or stable maintenance will occur.

The transcriptional initiation signals will include a promoter and a transcriptional initiation start site. In some instances, it may be desirable to provide for regulative expression of the toxin, where expression of the toxin will only occur after release into the environment. This can be achieved with operators or a region binding to an activator or enhancers, which are capable of induction upon a change in the physical or chemical environment of the microorganisms. For example, a temperature sensitive regulatory region may be employed, where the organisms may be grown up in the laboratory without expression of a toxin, but upon release

into the environment, expression would begin. Other techniques may employ a specific nutrient medium in the laboratory, which inhibits the expression of the toxin, where the nutrient medium in the environment would allow for expression of the toxin. For translational initiation, a ribosomal binding site and an initiation
5 codon will be present.

Various manipulations may be employed for enhancing the expression of the messenger RNA, particularly by using an active promoter, as well as by employing sequences, which enhance the stability of the messenger RNA. The transcriptional and translational termination region will involve stop codon(s), a terminator region, and optionally, a polyadenylation signal. A hydrophobic "leader"
10 sequence may be employed at the amino terminus of the translated polypeptide sequence in order to promote secretion of the protein across the inner membrane.

In the direction of transcription, namely in the 5' to 3' direction of the coding or sense sequence, the construct will involve the transcriptional regulatory region, if any, and the promoter, where the regulatory region may be either 5' or 3' of
15 the promoter, the ribosomal binding site, the initiation codon, the structural gene having an open reading frame in phase with the initiation codon, the stop codon(s), the polyadenylation signal sequence, if any, and the terminator region. This sequence as a double strand may be used by itself for transformation of a microorganism host, but will usually be included with a DNA sequence involving a
20 marker, where the second DNA sequence may be joined to the toxin expression construct during introduction of the DNA into the host.

By a marker is intended a structural gene which provides for selection of those hosts which have been modified or transformed. The marker will normally
25 provide for selective advantage, for example, providing for biocide resistance, *e.g.*, resistance to antibiotics or heavy metals; complementation, so as to provide prototrophy to an auxotrophic host, or the like. Preferably, complementation is employed, so that the modified host may not only be selected, but may also be competitive in the field. One or more markers may be employed in the development of the constructs, as well as for modifying the host. The organisms may be further modified
30 by providing for a competitive advantage against other wild-type microorganisms

in the field. For example, genes expressing metal chelating agents, *e.g.*, siderophores, may be introduced into the host along with the structural gene expressing the toxin. In this manner, the enhanced expression of a siderophore may provide for a competitive advantage for the toxin-producing host, so that it may effectively
5 compete with the wild-type microorganisms and stably occupy a niche in the environment.

Where no functional replication system is present, the construct will also include a sequence of at least 50 basepairs (bp), preferably at least about 100 bp, more preferably at least about 1000 bp, and usually not more than about 2000 bp of
10 a sequence homologous with a sequence in the host. In this way, the probability of legitimate recombination is enhanced, so that the gene will be integrated into the host and stably maintained by the host. Desirably, the toxin gene will be in close proximity to the gene providing for complementation as well as the gene providing for the competitive advantage. Therefore, in the event that a toxin gene is lost, the
15 resulting organism will be likely to also lost the complementing gene and/or the gene providing for the competitive advantage, so that it will be unable to compete in the environment with the gene retaining the intact construct.

A large number of transcriptional regulatory regions are available from a wide variety of microorganism hosts, such as bacteria, bacteriophage, cyanobacteria, algae, fungi, and the like. Various transcriptional regulatory regions include
20 the regions associated with the *trp* gene, *lac* gene, *gal* gene, the λ_L and λ_R promoters, the *tac* promoter, the naturally-occurring promoters associated with the δ -endotoxin gene, where functional in the host. See for example, U. S. Patents 4,332,898; 4,342,832; and 4,356,270 (each of which is specifically incorporated
25 herein by reference). The termination region may be the termination region normally associated with the transcriptional initiation region or a different transcriptional initiation region, so long as the two regions are compatible and functional in the host.

Where stable episomal maintenance or integration is desired, a plasmid will
30 be employed which has a replication system which is functional in the host. The replication system may be derived from the chromosome, an episomal element

normally present in the host or a different host, or a replication system from a virus which is stable in the host. A large number of plasmids are available, such as pBR322, pACYC184, RSF1010, pR01614, and the like. See for example, Olson *et al.* (1982); Bagdasarian *et al.* (1981), Baum *et al.*, 1990, and U. S. Patents
5 4,356,270; 4,362,817; 4,371,625, and 5,441,884, each incorporated specifically herein by reference.

The *B. thuringiensis* gene can be introduced between the transcriptional and translational initiation region and the transcriptional and translational termination region, so as to be under the regulatory control of the initiation region. This construct will be included in a plasmid, which will include at least one replication
10 system, but may include more than one, where one replication system is employed for cloning during the development of the plasmid and the second replication system is necessary for functioning in the ultimate host. In addition, one or more markers may be present, which have been described previously. Where integration
15 is desired, the plasmid will desirably include a sequence homologous with the host genome.

The transformants can be isolated in accordance with conventional ways, usually employing a selection technique, which allows for selection of the desired organism as against unmodified organisms or transferring organisms, when present.
20 The transformants then can be tested for pesticidal activity. If desired, unwanted or ancillary DNA sequences may be selectively removed from the recombinant bacterium by employing site-specific recombination systems, such as those described in U. S. Patent 5,441,884 (specifically incorporated herein by reference).

25 2.4 CRY3 DNA SEGMENTS

A *B. thuringiensis* *cry3** gene encoding a crystal protein having one or more mutations in one or more regions of the peptide represents an important aspect of the invention. Preferably, the *cry3** gene encodes an amino acid sequence in which one or more amino acid residues have been changed based on the methods disclosed herein, and particularly those changes which have been made for the
30 purpose of altering the insecticidal activity or specificity of the crystal protein.

In accordance with the present invention, nucleic acid sequences include and are not limited to DNA, including and not limited to cDNA and genomic DNA, genes; RNA, including and not limited to mRNA and tRNA; antisense sequences, nucleosides, and suitable nucleic acid sequences such as those set forth in
5 SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID
10 NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:99, SEQ ID NO:101, or SEQ ID NO:107, and alterations in the nucleic acid sequences including alterations, deletions, mutations, and homologs capable of expressing the *B. thuringiensis* modified toxins of the present invention.

15 As such the present invention also concerns DNA segments, that are free from total genomic DNA and that encode the novel synthetically-modified crystal proteins disclosed herein. DNA segments encoding these peptide species may prove to encode proteins, polypeptides, subunits, functional domains, and the like of crystal protein-related or other non-related gene products. In addition these
20 DNA segments may be synthesized entirely *in vitro* using methods that are well-known to those of skill in the art.

As used herein, the term "DNA segment" refers to a DNA molecule that has been isolated free of total genomic DNA of a particular species. Therefore, a DNA segment encoding a crystal protein or peptide refers to a DNA segment that con-
25 tains crystal protein coding sequences yet is isolated away from, or purified free from, total genomic DNA of the species from which the DNA segment is obtained, which in the instant case is the genome of the Gram-positive bacterial genus, *Bacillus*, and in particular, the species of *Bacillus* known as *B. thuringiensis*. Included within the term "DNA segment", are DNA segments and smaller fragments of such
30 segments, and also recombinant vectors, including, for example, plasmids, cosmids, phagemids, phage, viruses, and the like.

Similarly, a DNA segment comprising an isolated or purified crystal protein-encoding gene refers to a DNA segment which may include in addition to peptide encoding sequences, certain other elements such as, regulatory sequences, isolated- substantially away from other naturally occurring genes or protein-
 5 encoding sequences. In this respect, the term "gene" is used for simplicity to refer to a functional protein-, polypeptide- or peptide-encoding unit. As will be understood by those in the art, this functional term includes both genomic sequences, operon sequences and smaller engineered gene segments that express, or may be adapted to express, proteins, polypeptides or peptides.

10 "Isolated substantially away from other coding sequences" means that the gene of interest, in this case, a gene encoding a bacterial crystal protein, forms the significant part of the coding region of the DNA segment, and that the DNA segment does not contain large portions of naturally-occurring coding DNA, such as large chromosomal fragments or other functional genes or operon coding regions.
 15 Of course, this refers to the DNA segment as originally isolated, and does not exclude genes, recombinant genes, synthetic linkers, or coding regions later added to the segment by the hand of man.

Particularly preferred DNA sequences are those encoding Cry3Bb.60, Cry3Bb.11221, Cry3Bb.11222, Cry3Bb.11223, Cry3Bb.11224, Cry3Bb.11225,
 20 Cry3Bb.11226, Cry3Bb.11227, Cry3Bb.11228, Cry3Bb.11229, Cry3Bb.11230, Cry3Bb.11231, Cry3Bb.11232, Cry3Bb.11233, Cry3Bb.11234, Cry3Bb.11235, Cry3Bb.11236, Cry3Bb.11237, Cry3Bb.11238, Cry3Bb.11239, Cry3Bb.11241, Cry3Bb.11242, Cry3Bb.11032, Cry3Bb.11035, Cry3Bb.11036, Cry3Bb.11046, Cry3Bb.11048, Cry3Bb.11051, Cry3Bb.11057, Cry3Bb.11058, Cry3Bb.11081,
 25 Cry3Bb.11082, Cry3Bb.11083, Cry3Bb.11084, Cry3Bb.11095 and Cry3Bb.11098 crystal proteins, and in particular *cry3Bb** genes such as *cry3Bb.60*, *cry3Bb.11221*, *cry3Bb.11222*, *cry3Bb.11223*, *cry3Bb.11224*, *cry3Bb.11225*, *cry3Bb.11226*, *cry3Bb.11227*, *cry3Bb.11228*, *cry3Bb.11229*, *cry3Bb.11230*, *cry3Bb.11231*, *cry3Bb.11232*, *cry3Bb.11233*, *cry3Bb.11234*, *cry3Bb.11235*, *cry3Bb.11236*,
 30 *cry3Bb.11237*, *cry3Bb.11238*, *cry3Bb.11239*, *cry3Bb.11241*, *cry3Bb.11242*, *cry3Bb.11032*, *cry3Bb.11035*, *cry3Bb.11036*, *cry3Bb.11046*, *cry3Bb.11048*,

cry3Bb.11051, cry3Bb.11057, cry3Bb.11058, cry3Bb.11081, cry3Bb.11082, cry3Bb.11083, cry3Bb.11084, cry3Bb.11095 and cry3Bb.11098. In particular embodiments, the invention concerns isolated DNA segments and recombinant vectors incorporating DNA sequences that encode a Cry peptide species that includes
5 within its amino acid sequence an amino acid sequence essentially as set forth in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ
10 ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:108.

The term "a sequence essentially as set forth in SEQ ID NO:2, SEQ ID
15 NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID
20 NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:108" means that the sequence substantially corresponds to a portion of the sequence of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID
25 NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID
30 NO:68, SEQ ID NO:70, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:108, and has relatively few amino acids that are not identical to, or a biologically functional

equivalent of, the amino acids of any of these sequences. The term "biologically functional equivalent" is well understood in the art and is further defined in detail herein (*e.g.*, see Illustrative Embodiments).

Accordingly, sequences that have between about 70% and about 75% or
5 between about 75% and about 80%, or more preferably between about 81% and
about 90%, or even more preferably between about 91% or 92% or 93% and about
97% or 98% or 99% amino acid sequence identity or functional equivalence to the
amino acids of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ
ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ
10 ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ
ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ
ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ
ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ
ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ
15 ID NO:70, SEQ ID NO:100, SEQ ID NO:102 or SEQ ID NO:108 will be sequences
that are "essentially as set forth in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6,
SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16,
SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26,
SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36,
20 SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46,
SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56,
SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66,
SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID
NO:108."

25 It will also be understood that amino acid and nucleic acid sequences may
include additional residues, such as additional N- or C-terminal amino acids or 5'
or 3' sequences, and yet still be essentially as set forth in one of the sequences dis-
closed herein, so long as the sequence meets the criteria set forth above, including
the maintenance of biological protein activity where protein expression is con-
30 cerned. The addition of terminal sequences particularly applies to nucleic acid se-
quences that may, for example, include various non-coding sequences flanking ei-

ther of the 5' or 3' portions of the coding region or may include various internal sequences, *i.e.*, introns, which are known to occur within genes.

The nucleic acid segments of the present invention, regardless of the length of the coding sequence itself, may be combined with other DNA sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their overall length may vary considerably. It is therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant DNA protocol.

For example, nucleic acid fragments may be prepared that include a short contiguous stretch encoding the peptide sequence disclosed in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:108, or that are identical to or complementary to DNA sequences which encode the peptide disclosed in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:108, and particularly the DNA segments disclosed in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID

NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:99, SEQ ID NO:101, or SEQ ID NO:107.

Highly preferred nucleic acid segments of the present invention comprise one or more *cry* genes of the invention, or a portion of one or more *cry* genes of the invention. For certain application, relatively small contiguous nucleic acid sequences are preferable, such as those which are about 14 or 15 or 16 or 17 or 18 or 19, or 20, or 30-50, 51-80, 81-100 or so nucleotides in length. Alternatively, in some embodiments, and particularly those involving preparation of recombinant vectors, transformation of suitable host cells, and preparation of transgenic plant cell, longer nucleic acid segments are preferred, particularly those that include the entire coding region of one or more *cry* genes. As such, the preferred segments may include those that are up to about 20,000 or so nucleotides in length, or alternatively, shorter sequences such as those about 19,000, about 18,000, about 17,000, about 16,000, about 15,000, about 14,000, about 13,000, about 12,000, 11,000, about 10,000, about 9,000, about 8,000, about 7,000, about 6,000, about 5,000, about 4,500, about 4,000, about 3,500, about 3,000, about 2,500, about 2,000, about 1,500, about 1,000, about 500, or about 200 or so base pairs in length. Of course, these numbers are not intended to be exclusionary of all possible intermediate lengths in the range of from about 20,000 to about 15 nucleotides, as all of these intermediate lengths are also contemplated to be useful, and fall within the scope of the present invention. It will be readily understood that "intermediate lengths", in these contexts, means any length between the quoted ranges, such as 14, 15, 16, 17, 18, 19, 20, *etc.*; 21, 22, 23, 24, 25, 26, 27, 28, 29, *etc.*; 30, 31, 32, 33, 34, 35, 36..... *etc.*; 40, 41, 42, 43, 44..... *etc.*, 50, 51, 52, 53..... *etc.*; 60, 61, 62, 63.... *etc.*, 70, 80, 90, 100, 110, 120, 130..... *etc.*; 200, 210, 220, 230, 240, 250..... *etc.*; including all integers in the entire range from about 14 to about 10,000, including those integers in the ranges 200-500; 500-1,000; 1,000-2,000; 2,000-3,000; 3,000-5,000 and the like.

In a preferred embodiment, the nucleic acid segments comprise a sequence of from about 1800 to about 18,000 base pair in length, and comprise one or more genes which encode a modified Cry3* polypeptide disclosed herein which has increased activity against Coleopteran insect pests.

5 It will also be understood that this invention is not limited to the particular nucleic acid sequences which encode peptides of the present invention, or which encode the amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26,
10 SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID
15 NO:108, including the DNA sequences which are particularly disclosed in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41,
20 SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:99, SEQ ID NO:101, or SEQ ID NO:107. Recombinant vectors and isolated DNA segments may therefore variously include the peptide-coding regions themselves,
25 coding regions bearing selected alterations or modifications in the basic coding region, or they may encode larger polypeptides that nevertheless include these peptide-coding regions or may encode biologically functional equivalent proteins or peptides that have variant amino acids sequences.

The DNA segments of the present invention encompass biologically-
30 functional, equivalent peptides. Such sequences may arise as a consequence of codon redundancy and functional equivalency that are known to occur naturally

within nucleic acid sequences and the proteins thus encoded. Alternatively, functionally-equivalent proteins or peptides may be created *via* the application of recombinant DNA technology, in which changes in the protein structure may be engineered, based on considerations of the properties of the amino acids being ex-
5 changed. Changes designed by man may be introduced through the application of site-directed mutagenesis techniques, *e.g.*, to introduce improvements to the antigenicity of the protein or to test mutants in order to examine activity at the molecular level

If desired, one may also prepare fusion proteins and peptides, *e.g.*, where
10 the peptide-coding regions are aligned within the same expression unit with other proteins or peptides having desired functions, such as for purification or immunodetection purposes (*e.g.*, proteins that may be purified by affinity chromatography and enzyme label coding regions, respectively).

Recombinant vectors form further aspects of the present invention. Particularly
15 useful vectors are contemplated to be those vectors in which the coding portion of the DNA segment, whether encoding a full length protein or smaller peptide, is positioned under the control of a promoter. The promoter may be in the form of the promoter that is naturally associated with a gene encoding peptides of the present invention, as may be obtained by isolating the 5' non-coding sequences
20 located upstream of the coding segment or exon, for example, using recombinant cloning and/or PCR™ technology, in connection with the compositions disclosed herein.

2.5 VECTORS, HOST CELLS, AND PROTEIN EXPRESSION

25 In other embodiments, it is contemplated that certain advantages will be gained by positioning the coding DNA segment under the control of a recombinant, or heterologous, promoter. As used herein, a recombinant or heterologous promoter is intended to refer to a promoter that is not normally associated with a DNA segment encoding a crystal protein or peptide in its natural environment.
30 Such promoters may include promoters normally associated with other genes, and/or promoters isolated from any bacterial, viral, eukaryotic, or plant cell. Natu-

rally, it will be important to employ a promoter that effectively directs the expression of the DNA segment in the cell type, organism, or even animal, chosen for expression. The use of promoter and cell type combinations for protein expression is generally known to those of skill in the art of molecular biology, for example, see Sambrook *et al.*, 1989. The promoters employed may be constitutive, or inducible, and can be used under the appropriate conditions to direct high level expression of the introduced DNA segment, such as is advantageous in the large-scale production of recombinant proteins or peptides. Appropriate promoter systems contemplated for use in high-level expression include, but are not limited to, the *Pichia* expression vector system (Pharmacia LKB Biotechnology).

In connection with expression embodiments to prepare recombinant proteins and peptides, it is contemplated that longer DNA segments will most often be used, with DNA segments encoding the entire peptide sequence being most preferred. However, it will be appreciated that the use of shorter DNA segments to direct the expression of crystal peptides or epitopic core regions, such as may be used to generate anti-crystal protein antibodies, also falls within the scope of the invention. DNA segments that encode peptide antigens from about 8, 9, 10, or 11 or so amino acids, and up to and including those of about 30, 40, or 50 or so amino acids in length, or more preferably, from about 8 to about 30 amino acids in length, or even more preferably, from about 8 to about 20 amino acids in length are contemplated to be particularly useful. Such peptide epitopes may be amino acid sequences which comprise contiguous amino acid sequence from SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:108.

2.6 TRANSFORMED HOST CELLS AND TRANSGENIC PLANTS

In one embodiment, the invention provides a transgenic plant having incorporated into its genome a transgene that encodes a contiguous amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, SEQ ID NO:102, and SEQ ID NO:108.

A further aspect of the invention is a transgenic plant having incorporated into its genome a *cry3Bb** transgene, provided the transgene comprises a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:99, SEQ ID NO:101, and SEQ ID NO:107. Also disclosed and claimed are progeny of such a transgenic plant, as well as its seed, progeny from such seeds, and seeds arising from the second and subsequent generation plants derived from such a transgenic plant.

The invention also discloses and claims host cells, both native, and genetically engineered, which express the novel *cry3Bb** genes to produce Cry3Bb* polypeptides. Preferred examples of bacterial host cells include *B. thuringiensis* EG11221, EG11222, EG11223, EG11224, EG11225, EG11226, EG11227, EG11228, EG11229, EG11230, EG11231, EG11232, EG11233, EG11234,

EG11235, EG11236, EG11237, EG11238, EG11239, EG11241, EG11242, EG11032, EG11035, EG11036, EG11046, EG11048, EG11051, EG11057, EG11058, EG11081, EG11082, EG11083, EG11084, EG11095, and EG11098.

Methods of using such cells to produce Cry3* crystal proteins are also disclosed. Such methods generally involve culturing the host cell (such as *B. thuringiensis* EG11221, EG11222, EG11223, EG11224, EG11225, EG11226, EG11227, EG11228, EG11229, EG11230, EG11231, EG11232, EG11233, EG11234, EG11235, EG11236, EG11237, EG11238, EG11239, EG11241, EG11242, EG11032, EG11035, EG11036, EG11046, EG11048, EG11051, EG11057, EG11058, EG11081, EG11082, EG11083, EG11084, or EG11095, or EG11098) under conditions effective to produce a Cry3* crystal protein, and obtaining the Cry3* crystal protein from said cell.

In yet another aspect, the present invention provides methods for producing a transgenic plant which expresses a nucleic acid segment encoding the novel recombinant crystal proteins of the present invention. The process of producing transgenic plants is well-known in the art. In general, the method comprises transforming a suitable host cell with one or more DNA segments which contain one or more promoters operatively linked to a coding region that encodes one or more of the disclosed *B. thuringiensis* crystal proteins. Such a coding region is generally operatively linked to a transcription-terminating region, whereby the promoter is capable of driving the transcription of the coding region in the cell, and hence providing the cell the ability to produce the recombinant protein *in vivo*. Alternatively, in instances where it is desirable to control, regulate, or decrease the amount of a particular recombinant crystal protein expressed in a particular transgenic cell, the invention also provides for the expression of crystal protein antisense mRNA. The use of antisense mRNA as a means of controlling or decreasing the amount of a given protein of interest in a cell is well-known in the art.

Another aspect of the invention comprises a transgenic plant which express a gene or gene segment encoding one or more of the novel polypeptide compositions disclosed herein. As used herein, the term "transgenic plant" is intended to refer to a plant that has incorporated DNA sequences, including but not limited to

genes which are perhaps not normally present, DNA sequences not normally transcribed into RNA or translated into a protein ("expressed"), or any other genes or DNA sequences which one desires to introduce into the non-transformed plant, such as genes which may normally be present in the non-transformed plant but
5 which one desires to either genetically engineer or to have altered expression.

It is contemplated that in some instances the genome of a transgenic plant of the present invention will have been augmented through the stable introduction of one or more Cry3Bb*-encoding transgenes, either native, synthetically modified, or mutated. In some instances, more than one transgene will be incorporated
10 into the genome of the transformed host plant cell. Such is the case when more than one crystal protein-encoding DNA segment is incorporated into the genome of such a plant. In certain situations, it may be desirable to have one, two, three, four, or even more *B. thuringiensis* crystal proteins (either native or recombinantly-engineered) incorporated and stably expressed in the transformed transgenic plant.

15 A preferred gene which may be introduced includes, for example, a crystal protein-encoding a DNA sequence from bacterial origin, and particularly one or more of those described herein which are obtained from *Bacillus* spp. Highly preferred nucleic acid sequences are those obtained from *B. thuringiensis*, or any of those sequences which have been genetically engineered to decrease or increase the
20 insecticidal activity of the crystal protein in such a transformed host cell.

Means for transforming a plant cell and the preparation of a transgenic cell line are well-known in the art, and are discussed herein. Vectors, plasmids, cosmids, YACs (yeast artificial chromosomes) and DNA segments for use in transforming such cells will, of course, generally comprise either the operons, genes, or
25 gene-derived sequences of the present invention, either native, or synthetically-derived, and particularly those encoding the disclosed crystal proteins. These DNA constructs can further include structures such as promoters, enhancers, polylinkers, or even gene sequences which have positively- or negatively-regulating activity upon the particular genes of interest as desired. The DNA segment or gene may
30 encode either a native or modified crystal protein, which will be expressed in the

resultant recombinant cells, and/or which will impart an improved phenotype to the regenerated plant

Such transgenic plants may be desirable for increasing the insecticidal resistance of a monocotyledonous or dicotyledonous plant, by incorporating into such a plant, a transgenic DNA segment encoding a Cry3Bb* crystal protein which is toxic to coleopteran insects. Particularly preferred plants include grains such as corn, wheat, rye, rice, barley, and oats; legumes such as soybeans; tubers such as potatoes; fiber crops such as flax and cotton; turf and pasture grasses; ornamental plants; shrubs; trees; vegetables, berries, citrus, fruits, cacti, succulents, and other commercially-important crops including garden and houseplants.

In a related aspect, the present invention also encompasses a seed produced by the transformed plant, a progeny from such seed, and a seed produced by the progeny of the original transgenic plant, produced in accordance with the above process. Such progeny and seeds will have one or more crystal protein transgene(s) stably incorporated into its genome, and such progeny plants will inherit the traits afforded by the introduction of a stable transgene in Mendelian fashion. All such transgenic plants having incorporated into their genome transgenic DNA segments encoding one or more Cry3Bb* crystal proteins or polypeptides are aspects of this invention. Particularly preferred transgenes for the practice of the invention include nucleic acid segments comprising one or more *cry3Bb** gene(s).

2.7 BIOLOGICAL FUNCTIONAL EQUIVALENTS

Modification and changes may be made in the structure of the peptides of the present invention and DNA segments which encode them and still obtain a functional molecule that encodes a protein or peptide with desirable characteristics. The following is a discussion based upon changing the amino acids of a protein to create an equivalent, or even an improved, second-generation molecule. In particular embodiments of the invention, mutated crystal proteins are contemplated to be useful for increasing the insecticidal activity of the protein, and consequently increasing the insecticidal activity and/or expression of the recombinant transgene in

a plant cell. The amino acid changes may be achieved by changing the codons of the DNA sequence, according to the codons given in Table 4.

TABLE 4

Amino Acids			Codons						
Alanine	Ala	A	GCA	GCC	GCG	GCU			
Cysteine	Cys	C	UGC	UGU					
Aspartic Acid	Asp	D	GAC	GAU					
Glutamic Acid	Glu	E	GAA	GAG					
Phenylalanine	Phe	F	UUC	UUU					
Glycine	Gly	G	GGA	GGC	GGG	GGU			
Histidine	His	H	CAC	CAU					
Isoleucine	Ile	I	AUA	AUC	AUU				
Lysine	Lys	K	AAA	AAG					
Leucine	Leu	L	UUA	UUG	CUA	CUC	CUG	CUU	
Methionine	Met	M	AUG						
Asparagine	Asn	N	AAC	AAU					
Proline	Pro	P	CCA	CCC	CCG	CCU			
Glutamine	Gln	Q	CAA	CAG					
Arginine	Arg	R	AGA	AGG	CGA	CGC	CGG	CGU	
Serine	Ser	S	AGC	AGU	UCA	UCC	UCG	UCU	
Threonine	Thr	T	ACA	ACC	ACG	ACU			
Valine	Val	V	GUA	GUC	GUG	GUU			
Tryptophan	Trp	W	UGG						
Tyrosine	Tyr	Y	UAC	UAU					

5

For example, certain amino acids may be substituted for other amino acids in a protein structure without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites on substrate molecules. Since it is the interactive capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid

10

sequence substitutions can be made in a protein sequence, and, of course, its underlying DNA coding sequence, and nevertheless obtain a protein with like properties. It is thus contemplated by the inventors that various changes may be made in the peptide sequences of the disclosed compositions, or corresponding DNA sequences
5 which encode said peptides without appreciable loss of their biological utility or activity.

In making such changes, the hydropathic index of amino acids may be considered. The importance of the hydropathic amino acid index in conferring interactive biologic function on a protein is generally understood in the art (Kyte and
10 Doolittle, 1982, incorporate herein by reference). It is accepted that the relative hydropathic character of the amino acid contributes to the secondary structure of the resultant protein, which in turn defines the interaction of the protein with other molecules, for example, enzymes, substrates, receptors, DNA, antibodies, antigens, and the like.

Each amino acid has been assigned a hydropathic index on the basis of their hydrophobicity and charge characteristics (Kyte and Doolittle, 1982), these are:
15 isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (−0.4); threonine (−0.7); serine (−0.8); tryptophan (−0.9); tyrosine (−1.3); proline (−1.6); histidine (−3.2); glutamate (−3.5); glutamine (−3.5); aspartate (−3.5); asparagine (−3.5); lysine (−3.9); and arginine (−4.5).
20

It is known in the art that certain amino acids may be substituted by other amino acids having a similar hydropathic index or score and still result in a protein with similar biological activity, *i.e.*, still obtain a biological functionally equivalent
25 protein. In making such changes, the substitution of amino acids whose hydropathic indices are within ± 2 is preferred, those which are within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

It is also understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity. U. S. Patent 4,554,101, specifically
30 incorporated herein by reference, states that the greatest local average hydro-

philicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein.

As detailed in U. S. Patent 4,554,101, the following hydrophilicity values have been assigned to amino acid residues: arginine (+3.0); lysine (+3.0); aspartate
5 (+3.0 \pm 1); glutamate (+3.0 \pm 1); serine (+0.3); asparagine (+0.2); glutamine (+0.2); glycine (0); threonine (-0.4); proline (-0.5 \pm 1); alanine (-0.5); histidine (-0.5); cysteine (-1.0); methionine (-1.3); valine (-1.5); leucine (-1.8); isoleucine (-1.8); tyrosine (-2.3); phenylalanine (-2.5); tryptophan (-3.4).

It is understood that an amino acid can be substituted for another having a
10 similar hydrophilicity value and still obtain a biologically equivalent, and in particular, an immunologically equivalent protein. In such changes, the substitution of amino acids whose hydrophilicity values are within ± 2 is preferred, those which are within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

15 As outlined above, amino acid substitutions are generally therefore based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, and the like. Exemplary substitutions which take various of the foregoing characteristics into consideration are well known to those of skill in the art and include: arginine and lysine; glutamate
20 and aspartate; serine and threonine; glutamine and asparagine; and valine, leucine and isoleucine.

3.0 BRIEF DESCRIPTION OF THE DRAWINGS

The drawings form part of the present specification and are included to
25 further demonstrate certain aspects of the present invention. The invention may be better understood by reference to one or more of these drawings in combination with the detailed description of specific embodiments presented herein.

FIG. 1. Schematic representation of the monomeric structure of Cry3Bb.

30 **FIG. 2.** Stereoscopic view of the monomeric structure of Cry3Bb with associated water molecules (represented by dots).

FIG. 3A. Schematic representation of domain 1 of Cry3Bb

FIG. 3B. Diagram of the positions of the 7 helices that comprise domain 1.

FIG. 4. Domain 1 of Cry3Bb is organized into seven α helices illustrated in FIG. 3A (schematic representation) and FIG. 3B (schematic diagram). The α helices and amino acids residues are shown.

FIG. 5A. Schematic representation of domain 2 of Cry3Bb.

FIG. 5B. Diagram of the positions of the 11 β strands that compose the 3 β sheets of domain 2.

FIG. 6. Domain 2 of Cry3Bb is a collection of three anti-parallel β sheets illustrated in FIG. 5. The amino acids that define these sheets is listed below ($\alpha 8$, amino acids 322-328, also is included in domain 2):

FIG. 7A. Schematic representation of domain 3 of Cry3Bb.

FIG. 7B. Diagram of the positions of the β strands that comprise domain 3.

FIG. 8. Domain 3 (FIG. 7) is a loosely organized collection of β strands and loops; no β sheets are present. The β strands contain the amino acids limited below:

FIG. 9A. A "side" view of the dimeric structure of Cry3Bb. The helical bundles of domains 1 can be seen in the middle of the molecule.

FIG. 9B. A "top" view of the dimeric structure of Cry3Bb. The helical bundles of domains 1 can be seen in the middle of the molecule.

FIG. 10. A graphic representation of the growth in conductance with time of channels formed by Cry3A and Cry3Bb in planar lipid bilayers. Cry3A forms channels with higher conductances much more rapidly than Cry3Bb.

FIG. 11. A map of pEG1701 which contains the *Cry3Bb* gene with the *cryIF* terminator.

FIG. 12. The results of replicated 1-dose assays against SCRW larvae of Cry3Bb proteins altered in the 1B2,3 region.

FIG. 13. The results of replicated, 1-dose assays against SCRW larvae of Cry3Bb proteins altered in the 1B6, 7 region.

FIG. 14. The results of replicated, 1-dose screens against SCRW larvae of Cry3Bb proteins altered in the 1B10,11 region.

FIG. 15. Single channel recordings of channels formed by Cry3Bb.11230 and WT Cry3Bb in planar lipid bilayers. Cry3Bb.11230 forms channels with well resolved open and closed states while Cry3Bb rarely does.

FIG. 16. Single channel recordings of channels formed by Cry3Bb and Cry3Bb.60, a truncated form of Cry3Bb. Cry3Bb.60 forms channels more quickly than Cry3Bb and, unlike Cry3Bb, produces channels with well resolved open and closed states.

FIG. 17A. Sequence alignment of the amino acid sequence of Cry3A, Cry3B, and Cry3C.

FIG. 17B. Shown is a continuation of alignment of the amino acid sequence of Cry3A, Cry3B, and Cry3C shown in FIG. 17A.

FIG. 17C. Shown is a continuation of alignment of the amino acid sequence of Cry3A, Cry3B, and Cry3C shown in FIG. 17A.

4.0 DESCRIPTION OF ILLUSTRATIVE EMBODIMENTS

The invention defines new *B. thuringiensis* (Bt) insecticidal δ -endotoxin proteins and the biochemical and biophysical strategies used to design the new proteins. Delta-endotoxins are a class of insecticidal proteins produced by *B. thuringiensis* that form cation-selective channels in planar lipid bilayers (English and Slatin, 1992). The new δ -endotoxins are based on the parent structure of the coleopteran-active, δ -endotoxin Cry3Bb. Like other members of the coleopteran-active class of δ -endotoxins, including Cry3A and Cry3B, Cry3Bb exhibits excellent insecticidal activity against the Colorado Potato Beetle (*Leptinotarsa decemlineata*). However, unlike Cry3A and Cry3B, Cry3Bb is also active against the southern corn rootworm or SCRW (*Diabrotica undecimpunctata howardi* Barber) and the western corn rootworm or WCRW (*Diabrotica virgifera virgifera* LeConte). The new insecticidal proteins described herein were specifically designed to improve the biological activity of the parent Cry3Bb protein. In addition, the design strategies themselves are novel inventions capable of being applied to and

improving *B. thuringiensis* δ -endotoxins in general. *B. thuringiensis* δ -endotoxins are also members of a larger class of bacterial toxins that form ion channels (see English and Slatin 1992, for a review). The inventors, therefore, believe that these design strategies can also be applied to any biologically active, channel-forming protein to improve its biological properties.

The designed Cry3Bb proteins were engineered using one or more of the following strategies including (1) identification and alteration of protease-sensitive sites and proteolytic processing; (2) analysis and manipulation of bound water; (3) manipulation of hydrogen bonds around mobile regions; (4) loop analysis and loop redesign around flexible helices; (5) loop design around β strands and β sheets; (6) identification and redesign of complex electrostatic surfaces; (7) identification and removal of metal binding sites; (8) alteration of quaternary structure; (9) identification and design of structural residues; and (10) combinations of any and all sites defined by strategies 1-9. These design strategies permit the identification and redesign of specific sites on Cry3Bb, ultimately creating new proteins with improved insecticidal activities. These new proteins are designated Cry3Bb designed proteins and are named Cry3Bb followed by a period and a suffix (*e.g.*, Cry3Bb.60, Cry3Bb.11231). The new proteins are listed in Table 2 along with the specific sites on the molecule that were modified, the amino-acid sequence changes at those sites that improve biological activity, the improved insecticidal activities and the design method used to identify that specific site.

4.1 SOME ADVANTAGES OF THE INVENTION

Mutagenesis studies with *cry* genes have failed to identify a significant number of mutant crystal proteins which have improved broad-spectrum insecticidal activity, that is, with improved toxicity towards a range of insect pest species. Since agricultural crops are typically threatened by more than one insect pest species at any given time, desirable mutant crystal proteins are preferably those that exhibit improvements in toxicity towards multiple insect pest species. Previous failures to identify such mutants may be attributed to the choice of sites targeted for mutagenesis. For example, with respect to the related protein, Cry1C, sites

within domain 2 and domain 3 have been the principal targets of mutagenesis efforts, primarily because these domains are believed to be important for receptor binding and in determining insecticidal specificity (Aronson *et al.*, 1995; Chen *et al.* 1993; de Maagd *et al.*, 1996; Lee *et al.*, 1992; Lee *et al.*, 1995; Lu *et al.*, 1994; 5 Smedley and Ellar, 1996; Smith and Ellar, 1994; Rajamohan *et al.*, 1995; Rajamohan *et al.*, 1996)

In contrast, the present inventors reasoned that the toxicity of Cry3 proteins, and specifically the toxicity of the Cry3Bb protein, may be improved against a broader array of target pests by targeting regions involved in ion channel function 10 rather than regions of the molecule directly involved in receptor interactions, namely domains 2 and 3. Accordingly, the inventors opted to target regions within domain 1 of Cry3Bb for mutagenesis for the purpose of isolating Cry3Bb mutants with improved broad spectrum toxicity. Indeed, in the present invention, Cry3Bb mutants are described that show improved toxicity towards several coleopteran 15 pests.

At least one, and probably more than one, α helix of domain 1 is involved in the formation of ion channels and pores within the insect midgut epithelium (Gazit and Shai, 1993; Gazit and Shai, 1995). Rather than target for mutagenesis the sequences encoding the α helices of domain 1 as others have (Wu and Aronson, 20 1992; Aronson *et al.*, 1995; Chen *et al.*, 1995), the present inventors opted to target exclusively sequences encoding amino acid residues adjacent to or lying within the predicted loop regions of Cry3Bb that separate these α helices. Amino acid residues within these loop regions or amino acid residues capping the end of an α helix and lying adjacent to these loop regions may affect the spatial relationships among 25 these α helices. Consequently, the substitution of these amino acid residues may result in subtle changes in tertiary structure, or even quaternary structure, that positively impact the function of the ion channel. Amino acid residues in the loop regions of domain 1 are exposed to the solvent and thus are available for various molecular interactions. Altering these amino acids could result in greater stability 30 of the protein by eliminating or occluding protease-sensitive sites. Amino acid substitutions that change the surface charge of domain 1 could alter ion channel

efficiency or alter interactions with the brush border membrane or with other portions of the toxin molecule, allowing binding or insertion to be more effective.

According to this invention, base substitutions are made in the underlying *cry3Bb* nucleic acid residues in order to change particular codons of the corresponding polypeptides, and particularly, in those loop regions between α -helices. The insecticidal activity of a crystal protein ultimately dictates the level of crystal protein required for effective insect control. The potency of an insecticidal protein should be maximized as much as possible in order to provide for its economic and efficient utilization in the field. The increased potency of an insecticidal protein in a bioinsecticide formulation would be expected to improve the field performance of the bioinsecticide product. Alternatively, increased potency of an insecticidal protein in a bioinsecticide formulation may promote use of reduced amounts of bioinsecticide per unit area of treated crop, thereby allowing for more cost-effective use of the bioinsecticide product. When expressed *in planta*, the production of crystal proteins with improved insecticidal activity can be expected to improve plant resistance to susceptible insect pests.

4.2 METHODS FOR CULTURING *B. THURINGIENSIS* TO PRODUCE CRYSTAL PROTEINS

The *B. thuringiensis* strains described herein may be cultured using standard known media and fermentation techniques. Upon completion of the fermentation cycle, the bacteria may be harvested by first separating the *B. thuringiensis* spores and crystals from the fermentation broth by means well known in the art. The recovered *B. thuringiensis* spores and crystals can be formulated into a wettable powder, a liquid concentrate, granules or other formulations by the addition of surfactants, dispersants, inert carriers and other components to facilitate handling and application for particular target pests. The formulation and application procedures are all well known in the art.

4.3 RECOMBINANT HOST CELLS FOR EXPRESSION OF *CRY** GENES

The nucleotide sequences of the subject invention can be introduced into a wide variety of microbial hosts. Expression of the toxin gene results, directly or indirectly, in the intracellular production and maintenance of the pesticide. With suitable hosts, *e.g.*, *Pseudomonas*, the microbes can be applied to the sites of coleopteran insects where they will proliferate and be ingested by the insects. The result is a control of the unwanted insects. Alternatively, the microbe hosting the toxin gene can be treated under conditions that prolong the activity of the toxin produced in the cell. The treated cell then can be applied to the environment of target pest(s). The resulting product retains the toxicity of the *B. thuringiensis* toxin.

Suitable host cells, where the pesticide-containing cells will be treated to prolong the activity of the toxin in the cell when the then treated cell is applied to the environment of target pest(s), may include either prokaryotes or eukaryotes, normally being limited to those cells which do not produce substances toxic to higher organisms, such as mammals. However, organisms which produce substances toxic to higher organisms could be used, where the toxin is unstable or the level of application sufficiently low as to avoid any possibility of toxicity to a mammalian host. As hosts, of particular interest will be the prokaryotes and the lower eukaryotes, such as fungi. Illustrative prokaryotes, both Gram-negative and Gram-positive, include *Enterobacteriaceae*, such as *Escherichia*, *Erwinia*, *Shigella*, *Salmonella*, and *Proteus*; *Bacillaceae*; *Rhizobiceae*, such as *Rhizobium*; *Spirillaceae*, such as *Photobacterium*, *Zymomonas*, *Serratia*, *Aeromonas*, *Vibrio*, *Desulfovibrio*, *Spirillum*; *Lactobacillaceae*; *Pseudomonadaceae*, such as *Pseudomonas* and *Acetobacter*; *Azotobacteraceae*, *Actinomycetales*, and *Nitrobacteraceae*. Among eukaryotes are fungi, such as *Phycomycetes* and *Ascomycetes*, which includes yeast, such as *Saccharomyces* and *Schizosaccharomyces*; and *Basidiomycetes* yeast, such as *Rhodotorula*, *Aureobasidium*, *Sporobolomyces*, and the like.

Characteristics of particular interest in selecting a host cell for purposes of production include ease of introducing the *B. thuringiensis* gene into the host,

availability of expression systems, efficiency of expression, stability of the pesticide in the host, and the presence of auxiliary genetic capabilities. Characteristics of interest for use as a pesticide microcapsule include protective qualities for the pesticide, such as thick cell walls, pigmentation, and intracellular packaging or
5 formation of inclusion bodies; leaf affinity; lack of mammalian toxicity; attractiveness to pests for ingestion; ease of killing and fixing without damage to the toxin; and the like. Other considerations include ease of formulation and handling, economics, storage stability, and the like.

Host organisms of particular interest include yeast, such as *Rhodotorula*
10 *sp.*, *Aureobasidium sp.*, *Saccharomyces sp.*, and *Sporobolomyces sp.*; phylloplane organisms such as *Pseudomonas sp.*, *Erwinia sp.* and *Flavobacterium sp.*; or such other organisms as *Escherichia*, *Lactobacillus sp.*, *Bacillus sp.*, *Streptomyces sp.*, and the like. Specific organisms include *Pseudomonas aeruginosa*, *Pseudomonas fluorescens*, *Saccharomyces cerevisiae*, *B. thuringiensis*, *Escherichia coli*, *B. subtilis*,
15 *B. megaterium*, *B. cereus*, *Streptomyces lividans* and the like.

Treatment of the microbial cell, *e.g.*, a microbe containing the *B. thuringiensis* toxin gene, can be by chemical or physical means, or by a combination of chemical and/or physical means, so long as the technique does not deleteriously affect the properties of the toxin, nor diminish the cellular capability in
20 protecting the toxin. Examples of chemical reagents are halogenating agents, particularly halogens of atomic no. 17-80. More particularly, iodine can be used under mild conditions and for sufficient time to achieve the desired results. Other suitable techniques include treatment with aldehydes, such as formaldehyde and glutaraldehyde; anti-infectives, such as zephiran chloride and cetylpyridinium chloride;
25 alcohols, such as isopropyl and ethanol; various histologic fixatives, such as Lugol's iodine, Bouin's fixative, and Helly's fixatives, (see *e.g.*, Humason, 1967); or a combination of physical (heat) and chemical agents that preserve and prolong the activity of the toxin produced in the cell when the cell is administered to the host animal. Examples of physical means are short wavelength radiation such as γ -
30 radiation and X-radiation, freezing, UV irradiation, lyophilization, and the like. The cells employed will usually be intact and be substantially in the proliferative

form when treated, rather than in a spore form, although in some instances spores may be employed.

Where the *B. thuringiensis* toxin gene is introduced *via* a suitable vector into a microbial host, and said host is applied to the environment in a living state, it is essential that certain host microbes be used. Microorganism hosts are selected which are known to occupy the "phytosphere" (phylloplane, phyllosphere, rhizosphere, and/or rhizoplane) of one or more crops of interest. These microorganisms are selected so as to be capable of successfully competing in the particular environment (crop and other insect habitats) with the wild-type microorganisms, provide for stable maintenance and expression of the gene expressing the polypeptide pesticide, and, desirably, provide for improved protection of the pesticide from environmental degradation and inactivation.

A large number of microorganisms are known to inhabit the phylloplane (the surface of the plant leaves) and/or the rhizosphere (the soil surrounding plant roots) of a wide variety of important crops. These microorganisms include bacteria, algae, and fungi. Of particular interest are microorganisms, such as bacteria, *e.g.*, genera *Bacillus* (including the species and subspecies *B. thuringiensis kurstaki* HD-1, *B. thuringiensis kurstaki* HD-73, *B. thuringiensis sotto*, *B. thuringiensis berliner*, *B. thuringiensis thuringiensis*, *B. thuringiensis tolworthi*, *B. thuringiensis dendrolimus*, *B. thuringiensis alesti*, *B. thuringiensis galleriae*, *B. thuringiensis aizawai*, *B. thuringiensis subtoxicus*, *B. thuringiensis entomocidus*, *B. thuringiensis tenebrionis* and *B. thuringiensis san diego*); *Pseudomonas*, *Erwinia*, *Serratia*, *Klebsiella*, *Zanthomonas*, *Streptomyces*, *Rhizobium*, *Rhodopseudomonas*, *Methylophilus*, *Agrobacterium*, *Acetobacter*, *Lactobacillus*, *Arthrobacter*, *Azotobacter*, *Leuconostoc*, and *Alcaligenes*; fungi, particularly yeast, *e.g.*, genera *Saccharomyces*, *Cryptococcus*, *Kluyveromyces*, *Sporobolomyces*, *Rhodotorula*, and *Aureobasidium*. Of particular interest are such phytosphere bacterial species as *Pseudomonas syringae*, *Pseudomonas fluorescens*, *Serratia marcescens*, *Acetobacter xylinum*, *Agrobacterium tumefaciens*, *Rhodobacter sphaeroides*, *Xanthomonas campestris*, *Rhizobium melioli*, *Alcaligenes eutrophus*, and *Azotobacter vinlandii*; and phytosphere yeast species such as *Rhodotorula rubra*, *R. glutinis*, *R. marina*,

R. aurantiaca, *Cryptococcus albidus*, *C. diffluens*, *C. laurentii*, *Saccharomyces rosei*, *S. pretoriensis*, *S. cerevisiae*, *Sporobolomyces roseus*, *S. odoratus*, *Kluyveromyces veronae*, and *Aureobasidium pollulans*.

5 4.4 DEFINITIONS

In accordance with the present invention, nucleic acid sequences include and are not limited to DNA (including and not limited to genomic or extragenomic DNA), genes, RNA (including and not limited to mRNA and tRNA), nucleosides, and suitable nucleic acid segments either obtained from native sources, chemically
10 synthesized, modified, or otherwise prepared by the hand of man. The following words and phrases have the meanings set forth below.

A, an: In accordance with long standing patent law convention, the words "a" and "an" when used in this application, including the claims, denotes "one or more".

15 **Broad-spectrum:** Refers to a wide range of insect species.

Broad-spectrum activity: The toxicity towards a wide range of insect species.

Expression: The combination of intracellular processes, including transcription and translation undergone by a coding DNA molecule such as a structural
20 gene to produce a polypeptide.

Insecticidal activity: The toxicity towards insects.

Insecticidal specificity: The toxicity exhibited by a crystal protein or proteins, microbe or plant, towards multiple insect species.

Intraorder specificity: The toxicity of a particular crystal protein towards
25 insect species within an Order of insects (*e.g.*, Order Coleoptera).

Interorder specificity: The toxicity of a particular crystal protein towards insect species of different Orders (*e.g.*, Orders Coleoptera and Diptera).

LC₅₀: The lethal concentration of crystal protein that causes 50% mortality of the insects treated.

30 **LC₉₅:** The lethal concentration of crystal protein that causes 95% mortality of the insects treated.

Promoter: A recognition site on a DNA sequence or group of DNA sequences that provide an expression control element for a structural gene and to which RNA polymerase specifically binds and initiates RNA synthesis (transcription) of that gene.

5 **Regeneration:** The process of growing a plant from a plant cell (*e.g.*, plant protoplast or explant).

Structural gene: A gene that is expressed to produce a polypeptide.

Transformation: A process of introducing an exogenous DNA sequence (*e.g.*, a vector, a recombinant DNA molecule) into a cell or protoplast in which that
10 exogenous DNA is incorporated into a chromosome or is capable of autonomous replication.

Transformed cell: A cell whose DNA has been altered by the introduction of an exogenous DNA molecule into that cell.

Transgenic cell: Any cell derived or regenerated from a transformed cell
15 or derived from a transgenic cell. Exemplary transgenic cells include plant calli derived from a transformed plant cell and particular cells such as leaf, root, stem, *e.g.*, somatic cells, or reproductive (germ) cells obtained from a transgenic plant.

Transgenic plant: A plant or progeny thereof derived from a transformed plant cell or protoplast, wherein the plant DNA contains an introduced exogenous
20 DNA molecule not originally present in a native, non-transgenic plant of the same strain. The terms "transgenic plant" and "transformed plant" have sometimes been used in the art as synonymous terms to define a plant whose DNA contains an exogenous DNA molecule. However, it is thought more scientifically correct to refer to a regenerated plant or callus obtained from a transformed plant cell or protoplast
25 as being a transgenic plant, and that usage will be followed herein.

Vector: A DNA molecule capable of replication in a host cell and/or to which another DNA segment can be operatively linked so as to bring about replication of the attached segment. A plasmid is an exemplary vector.

As used herein, the designations "CryIII" and "Cry3" are synonymous, as
30 are the designations "CryIIIB2" and "Cry3Bb." Likewise, the inventors have utilized the generic term Cry3Bb* to denote any and all Cry3Bb variants which com-

prise amino acid sequences modified in the protein. Similarly, *cry3Bb** is meant to denote any and all nucleic acid segments and/or genes which encode a Cry3Bb* protein, *etc.*

5 4.5 PREPARATION OF *CRY3** POLYNUCLEOTIDES

Once the structure of the desired peptide to be mutagenized has been analyzed using one or more of the design strategies disclosed herein, it will be desirable to introduce one or more mutations into either the protein or, alternatively, into the DNA sequence encoding the protein for the purpose of producing a mutated protein with altered bioinsecticidal properties.

To that end, the present invention encompasses both site-specific mutagenesis methods and random mutagenesis of a nucleic acid segment encoding a crystal protein in the manner described herein. In particular, methods are disclosed for the mutagenesis of nucleic acid segments encoding the amino acid sequences using one or more of the design strategies described herein. Using the assay methods described herein, one may then identify mutants arising from these procedures which have improved insecticidal properties or altered specificity, either intraorder or interorder.

The means for mutagenizing a DNA segment encoding a crystal protein are well-known to those of skill in the art. Modifications may be made by random, or site-specific mutagenesis procedures. The nucleic acid may be modified by altering its structure through the addition or deletion of one or more nucleotides from the sequence.

Mutagenesis may be performed in accordance with any of the techniques known in the art such as and not limited to synthesizing an oligonucleotide having one or more mutations within the sequence of a particular crystal protein. A "suitable host" is any host which will express Cry3Bb, such as and not limited to *B. thuringiensis* and *E. coli*. Screening for insecticidal activity, in the case of Cry3Bb includes and is not limited to coleopteran-toxic activity which may be screened for by techniques known in the art.

In particular, site-specific mutagenesis is a technique useful in the preparation of individual peptides, or biologically functional equivalent proteins or peptides, through specific mutagenesis of the underlying DNA. The technique further provides a ready ability to prepare and test sequence variants, for example, incorporating one or more of the foregoing considerations, by introducing one or more nucleotide sequence changes into the DNA. Site-specific mutagenesis allows the production of mutants through the use of specific oligonucleotide sequences which encode the DNA sequence of the desired mutation, as well as a sufficient number of adjacent nucleotides, to provide a primer sequence of sufficient size and sequence complexity to form a stable duplex on both sides of the deletion junction being traversed. Typically, a primer of about 17 to about 75 nucleotides or more in length is preferred, with about 10 to about 25 or more residues on both sides of the junction of the sequence being altered.

In general, the technique of site-specific mutagenesis is well known in the art, as exemplified by various publications. As will be appreciated, the technique typically employs a phage vector which exists in both a single stranded and double stranded form. Typical vectors useful in site-directed mutagenesis include vectors such as the M13 phage. These phage are readily commercially available and their use is generally well known to those skilled in the art. Double stranded plasmids are also routinely employed in site directed mutagenesis which eliminates the step of transferring the gene of interest from a plasmid to a phage.

In general, site-directed mutagenesis in accordance herewith is performed by first obtaining a single-stranded vector or melting apart of two strands of a double stranded vector which includes within its sequence a DNA sequence which encodes the desired peptide. An oligonucleotide primer bearing the desired mutated sequence is prepared, generally synthetically. This primer is then annealed with the single-stranded vector, and subjected to DNA polymerizing enzymes such as *E. coli* polymerase I Klenow fragment, in order to complete the synthesis of the mutation-bearing strand. Thus, a heteroduplex is formed wherein one strand encodes the original non-mutated sequence and the second strand bears the desired mutation. This heteroduplex vector is then used to transform or transfect appro-

appropriate cells, such as *E. coli* cells, and clones are selected which include recombinant vectors bearing the mutated sequence arrangement. A genetic selection scheme was devised by Kunkel *et al.* (1987) to enrich for clones incorporating the mutagenic oligonucleotide. Alternatively, the use of PCRTM with commercially available thermostable enzymes such as *Taq* polymerase may be used to incorporate a mutagenic oligonucleotide primer into an amplified DNA fragment that can then be cloned into an appropriate cloning or expression vector. The PCRTM-mediated mutagenesis procedures of Tomic *et al.* (1990) and Upender *et al.* (1995) provide two examples of such protocols. A PCRTM employing a thermostable ligase in addition to a thermostable polymerase may also be used to incorporate a phosphorylated mutagenic oligonucleotide into an amplified DNA fragment that may then be cloned into an appropriate cloning or expression vector. The mutagenesis procedure described by Michael (1994) provides an example of one such protocol.

The preparation of sequence variants of the selected peptide-encoding DNA segments using site-directed mutagenesis is provided as a means of producing potentially useful species and is not meant to be limiting as there are other ways in which sequence variants of peptides and the DNA sequences encoding them may be obtained. For example, recombinant vectors encoding the desired peptide sequence may be treated with mutagenic agents, such as hydroxylamine, to obtain sequence variants.

As used herein, the term "oligonucleotide directed mutagenesis procedure" refers to template-dependent processes and vector-mediated propagation which result in an increase in the concentration of a specific nucleic acid molecule relative to its initial concentration, or in an increase in the concentration of a detectable signal, such as amplification. As used herein, the term "oligonucleotide directed mutagenesis procedure" is intended to refer to a process that involves the template-dependent extension of a primer molecule. The term template dependent process refers to nucleic acid synthesis of an RNA or a DNA molecule wherein the sequence of the newly synthesized strand of nucleic acid is dictated by the well-known rules of complementary base pairing (see, for example, Watson, 1987).

Typically, vector mediated methodologies involve the introduction of the nucleic acid fragment into a DNA or RNA vector, the clonal amplification of the vector, and the recovery of the amplified nucleic acid fragment. Examples of such methodologies are provided by U. S. Patent 4,237,224, specifically incorporated herein
5 by reference in its entirety

A number of template dependent processes are available to amplify the target sequences of interest present in a sample. One of the best known amplification methods is the polymerase chain reaction (PCR™) which is described in detail in U. S. Patents 4,683,195, 4,683,202 and 4,800,159 (each of which is specifically
10 incorporated herein by reference in its entirety). Briefly, in PCR™, two primer sequences are prepared which are complementary to regions on opposite complementary strands of the target sequence. An excess of deoxynucleoside triphosphates are added to a reaction mixture along with a DNA polymerase (*e.g.*, *Taq* polymerase). If the target sequence is present in a sample, the primers will bind to
15 the target and the polymerase will cause the primers to be extended along the target sequence by adding on nucleotides. By raising and lowering the temperature of the reaction mixture, the extended primers will dissociate from the target to form reaction products, excess primers will bind to the target and to the reaction products and the process is repeated. Preferably a reverse transcriptase PCR™ amplification
20 procedure may be performed in order to quantify the amount of mRNA amplified. Polymerase chain reaction methodologies are well known in the art.

Another method for amplification is the ligase chain reaction (referred to as LCR), disclosed in Eur. Pat. Appl. Publ. No. 320,308, incorporated herein by reference in its entirety. In LCR, two complementary probe pairs are prepared, and in
25 the presence of the target sequence, each pair will bind to opposite complementary strands of the target such that they abut. In the presence of a ligase, the two probe pairs will link to form a single unit. By temperature cycling, as in PCR™, bound ligated units dissociate from the target and then serve as "target sequences" for ligation of excess probe pairs. U. S. Patent 4,883,750, specifically incorporated
30 herein by reference in its entirety, describes an alternative method of amplification similar to LCR for binding probe pairs to a target sequence.

Qbeta ReplicaseTM, described in Intl. Pat. Appl. Publ. No. PCT/US87/00880, incorporated herein by reference in its entirety, may also be used as still another amplification method in the present invention. In this method, a replicative sequence of RNA which has a region complementary to that of a target is added to a sample in the presence of an RNA polymerase. The polymerase will copy the replicative sequence which can then be detected.

An isothermal amplification method, in which restriction endonucleases and ligases are used to achieve the amplification of target molecules that contain nucleotide 5'-[α -thio]triphosphates in one strand of a restriction site (Walker *et al.*, 1992, incorporated herein by reference in its entirety), may also be useful in the amplification of nucleic acids in the present invention.

Strand Displacement Amplification (SDA) is another method of carrying out isothermal amplification of nucleic acids which involves multiple rounds of strand displacement and synthesis, *i.e.*, nick translation. A similar method, called Repair Chain Reaction (RCR) is another method of amplification which may be useful in the present invention and is involves annealing several probes throughout a region targeted for amplification, followed by a repair reaction in which only two of the four bases are present. The other two bases can be added as biotinylated derivatives for easy detection. A similar approach is used in SDA

Sequences can also be detected using a cyclic probe reaction (CPR). In CPR, a probe having 3' and 5' end sequences of non-Cry-specific DNA and an internal sequence of a Cry-specific RNA is hybridized to DNA which is present in a sample. Upon hybridization, the reaction is treated with RNaseH, and the products of the probe identified as distinctive products generating a signal which are released after digestion. The original template is annealed to another cycling probe and the reaction is repeated. Thus, CPR involves amplifying a signal generated by hybridization of a probe to a *cry*-specific expressed nucleic acid

Still other amplification methods described in Great Britain Pat. Appl. No. 2 202 328, and in Intl. Pat. Appl. Publ. No. PCT/US89/01025, each of which is incorporated herein by reference in its entirety, may be used in accordance with the present invention. In the former application, "modified" primers are used in a

PCRTM like, template and enzyme dependent synthesis. The primers may be modified by labeling with a capture moiety (*e.g.*, biotin) and/or a detector moiety (*e.g.*, enzyme). In the latter application, an excess of labeled probes are added to a sample. In the presence of the target sequence, the probe binds and is cleaved
5 catalytically. After cleavage, the target sequence is released intact to be bound by excess probe. Cleavage of the labeled probe signals the presence of the target sequence

Other nucleic acid amplification procedures include transcription-based amplification systems (TAS) (Kwoh *et al.*, 1989; Intl. Pat. Appl. Publ. No. WO
10 88/10315, incorporated herein by reference in its entirety), including nucleic acid sequence based amplification (NASBA) and 3SR. In NASBA, the nucleic acids can be prepared for amplification by standard phenol/chloroform extraction, heat denaturation of a sample, treatment with lysis buffer and minispin columns for isolation of DNA and RNA or guanidinium chloride extraction of RNA. These
15 amplification techniques involve annealing a primer which has crystal protein-specific sequences. Following polymerization, DNA/RNA hybrids are digested with RNase H while double stranded DNA molecules are heat denatured again. In either case the single stranded DNA is made fully double stranded by addition of second crystal protein-specific primer, followed by polymerization. The double
20 stranded DNA molecules are then multiply transcribed by a polymerase such as T7 or SP6. In an isothermal cyclic reaction, the RNAs are reverse transcribed into double stranded DNA, and transcribed once against with a polymerase such as T7 or SP6. The resulting products, whether truncated or complete, indicate crystal protein-specific sequences.

25 Eur. Pat. Appl. Publ. No. 329,822, incorporated herein by reference in its entirety, disclose a nucleic acid amplification process involving cyclically synthesizing single-stranded RNA ("ssRNA"), ssDNA, and double-stranded DNA (dsDNA), which may be used in accordance with the present invention. The ssRNA is a first template for a first primer oligonucleotide, which is elongated by
30 reverse transcriptase (RNA-dependent DNA polymerase). The RNA is then removed from resulting DNA:RNA duplex by the action of ribonuclease H (RNase

H, an RNase specific for RNA in a duplex with either DNA or RNA). The resultant ssDNA is a second template for a second primer, which also includes the sequences of an RNA polymerase promoter (exemplified by T7 RNA polymerase) 5' to its homology to its template. This primer is then extended by DNA polymerase (exemplified by the large "Klenow" fragment of *E. coli* DNA polymerase I), resulting as a double-stranded DNA ("dsDNA") molecule, having a sequence identical to that of the original RNA between the primers and having additionally, at one end, a promoter sequence. This promoter sequence can be used by the appropriate RNA polymerase to make many RNA copies of the DNA. These copies can then re-enter the cycle leading to very swift amplification. With proper choice of enzymes, this amplification can be done isothermally without addition of enzymes at each cycle. Because of the cyclical nature of this process, the starting sequence can be chosen to be in the form of either DNA or RNA

Intl. Pat. Appl. Publ. No. WO 89/06700, incorporated herein by reference in its entirety, disclose a nucleic acid sequence amplification scheme based on the hybridization of a promoter/primer sequence to a target single-stranded DNA ("ssDNA") followed by transcription of many RNA copies of the sequence. This scheme is not cyclic; *i.e.*, new templates are not produced from the resultant RNA transcripts. Other amplification methods include "RACE" (Frohman, 1990), and "one-sided PCRTM" (Ohara, 1989) which are well-known to those of skill in the art.

Methods based on ligation of two (or more) oligonucleotides in the presence of nucleic acid having the sequence of the resulting "di-oligonucleotide", thereby amplifying the di-oligonucleotide (Wu and Dean, 1996, incorporated herein by reference in its entirety), may also be used in the amplification of DNA sequences of the present invention.

4.6 PHAGE-RESISTANT VARIANTS

In certain embodiments, one may desired to prepare one or more phage resistant variants of the *B. thuringiensis* mutants prepared by the methods described herein. To do so, an aliquot of a phage lysate is spread onto nutrient agar and allowed to dry. An aliquot of the phage sensitive bacterial strain is then plated di-

rectly over the dried lysate and allowed to dry. The plates are incubated at 30°C. The plates are incubated for 2 days and, at that time, numerous colonies could be seen growing on the agar. Some of these colonies are picked and subcultured onto nutrient agar plates. These apparent resistant cultures are tested for resistance by cross streaking with the phage lysate. A line of the phage lysate is streaked on the plate and allowed to dry. The presumptive resistant cultures are then streaked across the phage line. Resistant bacterial cultures show no lysis anywhere in the streak across the phage line after overnight incubation at 30°C. The resistance to phage is then reconfirmed by plating a lawn of the resistant culture onto a nutrient agar plate. The sensitive strain is also plated in the same manner to serve as the positive control. After drying, a drop of the phage lysate is plated in the center of the plate and allowed to dry. Resistant cultures showed no lysis in the area where the phage lysate has been placed after incubation at 30°C for 24 hours.

4.7 CRYSTAL PROTEIN COMPOSITIONS AS INSECTICIDES AND METHODS OF USE

Order Coleoptera comprises numerous beetle species including ground beetles, reticulated beetles, skin and larder beetles, long-horned beetles, leaf beetles, weevils, bark beetles, ladybird beetles, soldier beetles, stag beetles, water scavenger beetles, and a host of other beetles. A brief taxonomy of the Order is given at the website <http://www.ncbi.nlm.nih.gov/Taxonomy/tax.html>.

Particularly important among the Coleoptera are the agricultural pests included within the infraorders *Chrysomeliformia* and *Cucujiformia*. Members of the infraorder *Chrysomeliformia*, including the leaf beetles (*Chrysomelidae*) and the weevils (*Curculionidae*), are particularly problematic to agriculture, and are responsible for a variety of insect damage to crops and plants. The infraorder *Cucujiformia* includes the families *Coccinellidae*, *Cucujidae*, *Lagridae*, *Meloidae*, *Rhipiphoridae*, and *Tenebrionidae*. Within this infraorder, members of the family *Chrysomelidae* (which includes the genera *Exema*, *Chrysomela*, *Oreina*, *Chrysolina*, *Leptinotarsa*, *Goniocena*, *Oulema*, *Monozia*, *Ophraella*, *Cerotoma*,

Diabrotica, and *Lachnaia*), are well-known for their potential to destroy agricultural crops.

As the toxins of the present invention have been shown to be effective in combatting a variety of members of the order Coleoptera, the inventors contemplate that the insects of many Coleopteran genera may be controlled or eradicated using the polypeptide compositions described herein. Likewise, the methods described herein for generating modified polypeptides having enhanced insect specificity may also be useful in extending the range of the insecticidal activity of the modified polypeptides to other insect species within, and outside of, the Order Coleoptera.

As such, the inventors contemplate that the crystal protein compositions disclosed herein will find particular utility as insecticides for topical and/or systemic application to field crops, including but not limited to rice, wheat, alfalfa, corn (maize), soybeans, tobacco, potato, barley, canola (rapeseed), sugarbeet, sugarcane, flax, rye, oats, cotton, sunflower; grasses, such as pasture and turf grasses; fruits, citrus, nuts, trees, shrubs and vegetables; as well as ornamental plants, cacti, succulents, and the like.

Disclosed and claimed is a composition comprising an insecticidally-effective amount of a Cry3Bb* crystal protein composition. The composition preferably comprises the amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, or SEQ ID NO:108 or biologically-functional equivalents thereof.

The insecticide composition may also comprise a Cry3Bb* crystal protein that is encoded by a nucleic acid sequence having the sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ

ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:99, or SEQ ID NO:108, or, alternatively, a nucleic acid sequence which hybridizes to the nucleic acid sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:99, or SEQ ID NO:107 under conditions of moderate stringency.

The insecticidal compositions may comprise one or more *B. thuringiensis* cell types, or one or more cultures of such cells, or, alternatively, a mixture of one or more *B. thuringiensis* cells which express one or more of the novel crystal proteins of the invention in combination with another insecticidal composition. In certain aspects it may be desirable to prepare compositions which contain a plurality of crystal proteins, either native or modified, for treatment of one or more types of susceptible insects. The *B. thuringiensis* cells of the invention can be treated prior to formulation to prolong the insecticidal activity when the cells are applied to the environment of the target insect(s). Such treatment can be by chemical or physical means, or by a combination of chemical and/or physical means, so long as the technique does not deleteriously affect the properties of the insecticide, nor diminish the cellular capability in protecting the insecticide. Examples of chemical reagents are halogenerating agents, particularly halogens of atomic no. 17-80. More particularly, iodine can be used under mild conditions and for sufficient time to achieve the desired results. Other suitable techniques include treatment with al-

dehydres, such as formaldehyde and glutaraldehyde; anti-infectives, such as zephiran chloride; alcohols, such as isopropyl and ethanol; various histologic fixatives, such as Bouin's fixative and Helly's fixative (*see* Humason, 1967); or a combination of physical (heat) and chemical agents that prolong the activity of the
5 δ -endotoxin produced in the cell when the cell is applied to the environment of the target pest(s). Examples of physical means are short wavelength radiation such as gamma-radiation and X-radiation, freezing, UV irradiation, lyophilization, and the like.

The inventors contemplate that any formulation methods known to those of
10 skill in the art may be employed using the proteins disclosed herein to prepare such bioinsecticide compositions. It may be desirable to formulate whole cell preparations, cell extracts, cell suspensions, cell homogenates, cell lysates, cell supernatants, cell filtrates, or cell pellets of a cell culture (preferably a bacterial cell culture such as a *B. thuringiensis* cell culture described in Table 3) that expresses one
15 or more *cry3Bb** DNA segments to produce the encoded Cry3Bb* protein(s) or peptide(s). The methods for preparing such formulations are known to those of skill in the art, and may include, *e.g.*, desiccation, lyophilization, homogenization, extraction, filtration, centrifugation, sedimentation, or concentration of one or more cultures of bacterial cells, such as *B. thuringiensis* cells described in Table 3, which
20 express the Cry3Bb* peptide(s) of interest.

In one preferred embodiment, the bioinsecticide composition comprises an oil flowable suspension comprising lysed or unlysed bacterial cells, spores, or crystals which contain one or more of the novel crystal proteins disclosed herein. Preferably the cells are *B. thuringiensis* cells, however, any such bacterial host cell
25 expressing the novel nucleic acid segments disclosed herein and producing a crystal protein is contemplated to be useful, such as *Bacillus* spp., including *B. megaterium*, *B. subtilis*; *B. cereus*, *Escherichia* spp., including *E. coli*, and/or *Pseudomonas* spp., including *P. cepacia*, *P. aeruginosa*, and *P. fluorescens*. Alternatively, the oil flowable suspension may consist of a combination of one or
30 more of the following compositions: lysed or unlysed bacterial cells, spores, crystals, and/or purified crystal proteins.

In a second preferred embodiment, the bioinsecticide composition comprises a water dispersible granule or powder. This granule or powder may comprise lysed or unlysed bacterial cells, spores, or crystals which contain one or more of the novel crystal proteins disclosed herein. Preferred sources for these compositions include bacterial cells such as *B. thuringiensis* cells, however, bacteria of the genera *Bacillus*, *Escherichia*, and *Pseudomonas* which have been transformed with a DNA segment disclosed herein and expressing the crystal protein are also contemplated to be useful. Alternatively, the granule or powder may consist of a combination of one or more of the following compositions: lysed or unlysed bacterial cells, spores, crystals, and/or purified crystal proteins.

In a third important embodiment, the bioinsecticide composition comprises a wettable powder, spray, emulsion, colloid, aqueous or organic solution, dust, pellet, or colloidal concentrate. Such a composition may contain either unlysed or lysed bacterial cells, spores, crystals, or cell extracts as described above, which contain one or more of the novel crystal proteins disclosed herein. Preferred bacterial cells are *B. thuringiensis* cells, however, bacteria such as *B. megaterium*, *B. subtilis*, *B. cereus*, *E. coli*, or *Pseudomonas* spp. cells transformed with a DNA segment disclosed herein and expressing the crystal protein are also contemplated to be useful. Such dry forms of the insecticidal compositions may be formulated to dissolve immediately upon wetting, or alternatively, dissolve in a controlled-release, sustained-release, or other time-dependent manner. Alternatively, such a composition may consist of a combination of one or more of the following compositions: lysed or unlysed bacterial cells, spores, crystals, and/or purified crystal proteins.

In a fourth important embodiment, the bioinsecticide composition comprises an aqueous solution or suspension or cell culture of lysed or unlysed bacterial cells, spores, crystals, or a mixture of lysed or unlysed bacterial cells, spores, and/or crystals, such as those described above which contain one or more of the novel crystal proteins disclosed herein. Such aqueous solutions or suspensions may be provided as a concentrated stock solution which is diluted prior to application, or alternatively, as a diluted solution ready-to-apply.

For these methods involving application of bacterial cells, the cellular host containing the Crystal protein gene(s) may be grown in any convenient nutrient medium, where the DNA construct provides a selective advantage, providing for a selective medium so that substantially all or all of the cells retain the
5 *B. thuringiensis* gene. These cells may then be harvested in accordance with conventional ways. Alternatively, the cells can be treated prior to harvesting.

When the insecticidal compositions comprise *B. thuringiensis* cells, spores, and/or crystals containing the modified crystal protein(s) of interest, such compositions may be formulated in a variety of ways. They may be employed as wettable
10 powders, granules or dusts, by mixing with various inert materials, such as inorganic minerals (phyllosilicates, carbonates, sulfates, phosphates, and the like) or botanical materials (powdered corncobs, rice hulls, walnut shells, and the like). The formulations may include spreader-sticker adjuvants, stabilizing agents, other pesticidal additives, or surfactants. Liquid formulations may be aqueous-based or
15 non-aqueous and employed as foams, suspensions, emulsifiable concentrates, or the like. The ingredients may include rheological agents, surfactants, emulsifiers, dispersants, or polymers.

Alternatively, the novel Cry3Bb-derived mutated crystal proteins may be prepared by native or recombinant bacterial expression systems *in vitro* and iso-
20 lated for subsequent field application. Such protein may be either in crude cell lysates, suspensions, colloids, *etc.*, or alternatively may be purified, refined, buffered, and/or further processed, before formulating in an active biocidal formulation. Likewise, under certain circumstances, it may be desirable to isolate crystals and/or spores from bacterial cultures expressing the crystal protein and apply solu-
25 tions, suspensions, or colloidal preparations of such crystals and/or spores as the active bioinsecticidal composition.

Another important aspect of the invention is a method of controlling coleopterian insects which are susceptible to the novel compositions disclosed herein. Such a method generally comprises contacting the insect or insect population, colony, *etc.*, with an insecticidally-effective amount of a Cry3Bb* crystal protein
30 composition. The method may utilize Cry3Bb* crystal proteins such as those

disclosed in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, or SEQ ID NO:108, or biologically functional equivalents thereof.

10 Alternatively, the method may utilize one or more Cry3Bb* crystal proteins which are encoded by the nucleic acid sequences of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:99, SEQ ID NO:101, or SEQ ID NO:107, or by one or more nucleic acid sequences which hybridize to the sequences of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:99, SEQ ID NO:101, or SEQ ID NO:107, under conditions of moderate, or higher, stringency. The methods for identifying sequences which hybridize to those disclosed under conditions of moderate or higher stringency are well-known to those of skill in the art, and are discussed herein.

Regardless of the method of application, the amount of the active component(s) are applied at an insecticidally-effective amount, which will vary depending on such factors as, for example, the specific coleopteran insects to be controlled, the specific plant or crop to be treated, the environmental conditions, and the method, rate, and quantity of application of the insecticidally-active composition.

The insecticide compositions described may be made by formulating either the bacterial cell, crystal and/or spore suspension, or isolated protein component with the desired agriculturally-acceptable carrier. The compositions may be formulated prior to administration in an appropriate means such as lyophilized, freeze-dried, dessicated, or in an aqueous carrier, medium or suitable diluent, such as saline or other buffer. The formulated compositions may be in the form of a dust or granular material, or a suspension in oil (vegetable or mineral), or water or oil/water emulsions, or as a wettable powder, or in combination with any other carrier material suitable for agricultural application. Suitable agricultural carriers can be solid or liquid and are well known in the art. The term "agriculturally-acceptable carrier" covers all adjuvants, *e.g.*, inert components, dispersants, surfactants, tackifiers, binders, *etc.* that are ordinarily used in insecticide formulation technology; these are well known to those skilled in insecticide formulation. The formulations may be mixed with one or more solid or liquid adjuvants and prepared by various means, *e.g.*, by homogeneously mixing, blending and/or grinding the insecticidal composition with suitable adjuvants using conventional formulation techniques.

The insecticidal compositions of this invention are applied to the environment of the target coleopteran insect, typically onto the foliage of the plant or crop to be protected, by conventional methods, preferably by spraying. The strength and duration of insecticidal application will be set with regard to conditions specific to the particular pest(s), crop(s) to be treated and particular environmental conditions. The proportional ratio of active ingredient to carrier will naturally depend on the chemical nature, solubility, and stability of the insecticidal composition, as well as the particular formulation contemplated.

Other application techniques, *e.g.*, dusting, sprinkling, soaking, soil injection, soil tilling, seed coating, seedling coating, spraying, aerating, misting, atomizing, and the like, are also feasible and may be required under certain circumstances such as *e.g.*, insects that cause root or stalk infestation, or for application to delicate
5 vegetation or ornamental plants. These application procedures are also well-known to those of skill in the art.

The insecticidal composition of the invention may be employed in the method of the invention singly or in combination with other compounds, including and not limited to other pesticides. The method of the invention may also be used
10 in conjunction with other treatments such as surfactants, detergents, polymers or time-release formulations. The insecticidal compositions of the present invention may be formulated for either systemic or topical use.

The concentration of insecticidal composition which is used for environmental, systemic, or foliar application will vary widely depending upon the nature
15 of the particular formulation, means of application, environmental conditions, and degree of biocidal activity. Typically, the bioinsecticidal composition will be present in the applied formulation at a concentration of at least about 1% by weight and may be up to and including about 99% by weight. Dry formulations of the compositions may be from about 1% to about 99% or more by weight of the composition,
20 while liquid formulations may generally comprise from about 1% to about 99% or more of the active ingredient by weight. Formulations which comprise intact bacterial cells will generally contain from about 10^4 to about 10^{12} cells/mg

The insecticidal formulation may be administered to a particular plant or target area in one or more applications as needed, with a typical field application
25 rate per hectare ranging on the order of from about 1 g to about 1 kg, 2 kg, 5, kg, or more of active ingredient.

4.8 NUCLEIC ACID SEGMENTS AS HYBRIDIZATION PROBES AND PRIMERS

In addition to their use in directing the expression of crystal proteins or
30 peptides of the present invention, the nucleic acid sequences contemplated herein also have a variety of other uses. For example, they also have utility as probes or

primers in nucleic acid hybridization embodiments. As such, it is contemplated that nucleic acid segments that comprise a sequence region that consists of at least a 14 nucleotide long contiguous sequence that has the same sequence as, or is complementary to, a 14 nucleotide long contiguous DNA segment of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:99, SEQ ID NO:101, or SEQ ID NO:107 will find particular utility. Longer contiguous identical or complementary sequences, *e.g.*, those of about 20, 30, 40, 50, 100, 200, 500, 1000, 2000, 5000, 10000 *etc.* (including all intermediate lengths and up to and including full-length sequences will also be of use in certain embodiments.

The ability of such nucleic acid probes to specifically hybridize to crystal protein-encoding sequences will enable them to be of use in detecting the presence of complementary sequences in a given sample. However, other uses are envisioned, including the use of the sequence information for the preparation of mutant species primers, or primers for use in preparing other genetic constructions.

Nucleic acid molecules having sequence regions consisting of contiguous nucleotide stretches of 10-14, 15-20, 30, 50, or even of 100-200 nucleotides or so, identical or complementary to DNA sequences of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:99, SEQ ID NO:101, or SEQ ID NO:107 are particularly contemplated as hybridization probes for use in,

e.g., Southern and Northern blotting. Smaller fragments will generally find use in hybridization embodiments, wherein the length of the contiguous complementary region may be varied, such as between about 10-14 and about 100 or 200 nucleotides, but larger contiguous complementary stretches may be used, according to the
5 length complementary sequences one wishes to detect.

The use of a hybridization probe of about 14 nucleotides in length allows the formation of a duplex molecule that is both stable and selective. Molecules having contiguous complementary sequences over stretches greater than 14 bases in length are generally preferred, though, in order to increase stability and selectiv-
10 ity of the hybrid, and thereby improve the quality and degree of specific hybrid molecules obtained. One will generally prefer to design nucleic acid molecules having gene-complementary stretches of 15 to 20 contiguous nucleotides, or even longer where desired.

Of course, fragments may also be obtained by other techniques such as,
15 *e.g.*, by mechanical shearing or by restriction enzyme digestion. Small nucleic acid segments or fragments may be readily prepared by, for example, directly synthesizing the fragment by chemical means, as is commonly practiced using an automated oligonucleotide synthesizer. Also, fragments may be obtained by application of nucleic acid reproduction technology, such as the PCR™ technology of U. S. Pat-
20 ents 4,683,195 and 4,683,202 (each incorporated herein by reference), by introducing selected sequences into recombinant vectors for recombinant production, and by other recombinant DNA techniques generally known to those of skill in the art of molecular biology.

Accordingly, the nucleotide sequences of the invention may be used for
25 their ability to selectively form duplex molecules with complementary stretches of DNA fragments. Depending on the application envisioned, one will desire to employ varying conditions of hybridization to achieve varying degrees of selectivity of probe towards target sequence. For applications requiring high selectivity, one will typically desire to employ relatively stringent conditions to form the hybrids,
30 *e.g.*, one will select relatively low salt and/or high temperature conditions, such as provided by about 0.02 M to about 0.15 M NaCl at temperatures of about 50°C to

about 70°C. Such selective conditions tolerate little, if any, mismatch between the probe and the template or target strand, and would be particularly suitable for isolating crystal protein-encoding DNA segments. Detection of DNA segments *via* hybridization is well-known to those of skill in the art, and the teachings of U. S. Patents 4,965,188 and 5,176,995 (each incorporated herein by reference) are exemplary of the methods of hybridization analyses. Teachings such as those found in the texts of Maloy *et al.*, 1994; Segal 1976; Prokop, 1991; and Kuby, 1994, are particularly relevant.

Of course, for some applications, for example, where one desires to prepare mutants employing a mutant primer strand hybridized to an underlying template or where one seeks to isolate crystal protein-encoding sequences from related species, functional equivalents, or the like, less stringent hybridization conditions will typically be needed in order to allow formation of the heteroduplex. In these circumstances, one may desire to employ conditions such as about 0.15 M to about 0.9 M salt, at temperatures ranging from about 20°C to about 55°C. Cross-hybridizing species can thereby be readily identified as positively hybridizing signals with respect to control hybridizations. In any case, it is generally appreciated that conditions can be rendered more stringent by the addition of increasing amounts of formamide, which serves to destabilize the hybrid duplex in the same manner as increased temperature. Thus, hybridization conditions can be readily manipulated, and thus will generally be a method of choice depending on the desired results.

In certain embodiments, it will be advantageous to employ nucleic acid sequences of the present invention in combination with an appropriate means, such as a label, for determining hybridization. A wide variety of appropriate indicator means are known in the art, including fluorescent, radioactive, enzymatic or other ligands, such as avidin/biotin, which are capable of giving a detectable signal. In preferred embodiments, one will likely desire to employ a fluorescent label or an enzyme tag, such as urease, alkaline phosphatase or peroxidase, instead of radioactive or other environmental undesirable reagents. In the case of enzyme tags, colorimetric indicator substrates are known that can be employed to provide a means

visible to the human eye or spectrophotometrically, to identify specific hybridization with complementary nucleic acid-containing samples.

In general, it is envisioned that the hybridization probes described herein will be useful both as reagents in solution hybridization as well as in embodiments employing a solid phase. In embodiments involving a solid phase, the test DNA (or RNA) is adsorbed or otherwise affixed to a selected matrix or surface. This fixed, single-stranded nucleic acid is then subjected to specific hybridization with selected probes under desired conditions. The selected conditions will depend on the particular circumstances based on the particular criteria required (depending, for example, on the G+C content, type of target nucleic acid, source of nucleic acid, size of hybridization probe, *etc.*). Following washing of the hybridized surface so as to remove nonspecifically bound probe molecules, specific hybridization is detected, or even quantitated, by means of the label.

4.9 CHARACTERISTICS OF MODIFIED CRY3 δ -ENDOTOXINS

The present invention provides novel polypeptides that define a whole or a portion of a *B. thuringiensis* cry3Bb.60, cry3Bb.11221, cry3Bb.11222, cry3Bb.11223, cry3Bb.11224, cry3Bb.11225, cry3Bb.11226, cry3Bb.11227, cry3Bb.11228, cry3Bb.11229, cry3Bb.11230, cry3Bb.11231, cry3Bb.11232, cry3Bb.11233, cry3Bb.11234, cry3Bb.11235, cry3Bb.11236, cry3Bb.11237, cry3Bb.11238, cry3Bb.11239, cry3Bb.11241, cry3Bb.11242, cry3Bb.11032, cry3Bb.11035, cry3Bb.11036, cry3Bb.11046, cry3Bb.11048, cry3Bb.11051, cry3Bb.11057, cry3Bb.11058, cry3Bb.11081, cry3Bb.11082, cry3Bb.11083, cry3Bb.11084, cry3Bb.11095 and cry3Bb.11098-encoded crystal protein.

4.10 CRYSTAL PROTEIN NOMENCLATURE

The inventors have arbitrarily assigned the designations Cry3Bb.60, Cry3Bb.11221, Cry3Bb.11222, Cry3Bb.11223, Cry3Bb.11224, Cry3Bb.11225, Cry3Bb.11226, Cry3Bb.11227, Cry3Bb.11228, Cry3Bb.11229, Cry3Bb.11230, Cry3Bb.11231, Cry3Bb.11232, Cry3Bb.11233, Cry3Bb.11234, Cry3Bb.11235, Cry3Bb.11236, Cry3Bb.11237, Cry3Bb.11238, Cry3Bb.11239, Cry3Bb.11241,

Cry3Bb.11242, Cry3Bb.11032, Cry3Bb.11035, Cry3Bb.11036, Cry3Bb.11046, Cry3Bb.11048, Cry3Bb.11051, Cry3Bb.11057, Cry3Bb.11058, Cry3Bb.11081, Cry3Bb.11082, Cry3Bb.11083, Cry3Bb.11084, Cry3Bb.11095 and Cry3Bb.11098 to the novel proteins of the invention.

5 Likewise, the arbitrary designations of *cry3Bb.60*, *cry3Bb.11221*, *cry3Bb.11222*, *cry3Bb.11223*, *cry3Bb.11224*, *cry3Bb.11225*, *cry3Bb.11226*, *cry3Bb.11227*, *cry3Bb.11228*, *cry3Bb.11229*, *cry3Bb.11230*, *cry3Bb.11231*, *cry3Bb.11232*, *cry3Bb.11233*, *cry3Bb.11234*, *cry3Bb.11235*, *cry3Bb.11236*, *cry3Bb.11237*, *cry3Bb.11238*, *cry3Bb.11239*, *cry3Bb.11241*, *cry3Bb.11242*,
10 *cry3Bb.11032*, *cry3Bb.11035*, *cry3Bb.11036*, *cry3Bb.11046*, *cry3Bb.11048*, *cry3Bb.11051*, *cry3Bb.11057*, *cry3Bb.11058*, *cry3Bb.11081*, *cry3Bb.11082*, *cry3Bb.11083*, *cry3Bb.11084*, *cry3Bb.11095* and *Cry3Bb.11098* have been assigned to the novel nucleic acid sequences which encode these polypeptides, respectively. While formal assignment of gene and protein designations based on the
15 revised nomenclature of crystal protein endotoxins (Table 1) may be made by the committee on the nomenclature of *B. thuringiensis*, any re-designations of the compositions of the present invention are also contemplated to be fully within the scope of the present disclosure.

20 4.11 TRANSFORMED HOST CELLS AND TRANSGENIC PLANTS

A bacterium, a yeast cell, or a plant cell or a plant transformed with an expression vector of the present invention is also contemplated. A transgenic bacterium, yeast cell, plant cell or plant derived from such a transformed or transgenic cell is also one aspect of the invention.

25 Such transformed host cells are often desirable for use in the production of endotoxins and for expression of the various DNA gene constructs disclosed herein. In some aspects of the invention, it is often desirable to modulate, regulate, or otherwise control the expression of the gene segments disclosed herein. Such methods are routine to those of skill in the molecular genetic arts. Typically, when
30 increased or over-expression of a particular gene is desired, various manipulations may be employed for enhancing the expression of the messenger RNA, particularly

by using an active promoter, as well as by employing sequences, which enhance the stability of the messenger RNA in the particular transformed host cell.

Typically, the initiation and translational termination region will involve stop codon(s), a terminator region, and optionally, a polyadenylation signal. In the
5 direction of transcription, namely in the 5' to 3' direction of the coding or sense sequence, the construct will involve the transcriptional regulatory region, if any, and the promoter, where the regulatory region may be either 5' or 3' of the promoter, the ribosomal binding site, the initiation codon, the structural gene having an open reading frame in phase with the initiation codon, the stop codon(s), the
10 polyadenylation signal sequence, if any, and the terminator region. This sequence as a double strand may be used by itself for transformation of a microorganism host, but will usually be included with a DNA sequence involving a marker, where the second DNA sequence may be joined to the δ -endotoxin expression construct during introduction of the DNA into the host.

15 By a marker is intended a structural gene which provides for selection of those hosts which have been modified or transformed. The marker will normally provide for selective advantage, for example, providing for biocide resistance, *e.g.*, resistance to antibiotics or heavy metals; complementation, so as to provide prototrophy to an auxotrophic host, or the like. Preferably, complementation is employed,
20 so that the modified host may not only be selected, but may also be competitive in the field. One or more markers may be employed in the development of the constructs, as well as for modifying the host. The organisms may be further modified by providing for a competitive advantage against other wild-type microorganisms in the field. For example, genes expressing metal chelating agents, *e.g.*, sidero-
25 phores, may be introduced into the host along with the structural gene expressing the δ -endotoxin. In this manner, the enhanced expression of a siderophore may provide for a competitive advantage for the δ -endotoxin-producing host, so that it may effectively compete with the wild-type microorganisms and stably occupy a niche in the environment.

30 Where no functional replication system is present, the construct will also include a sequence of at least 50 basepairs (bp), preferably at least about 100 bp,

and usually not more than about 1000 bp of a sequence homologous with a sequence in the host. In this way, the probability of legitimate recombination is enhanced, so that the gene will be integrated into the host and stably maintained by the host. Desirably, the δ -endotoxin gene will be in close proximity to the gene
5 providing for complementation as well as the gene providing for the competitive advantage. Therefore, in the event that a δ -endotoxin gene is lost, the resulting organism will be likely to also lose the complementing gene and/or the gene providing for the competitive advantage, so that it will be unable to compete in the environment with the gene retaining the intact construct.

10 The crystal protein-encoding gene can be introduced between the transcriptional and translational initiation region and the transcriptional and translational termination region, so as to be under the regulatory control of the initiation region. This construct will be included in a plasmid, which will include at least one replication system, but may include more than one, where one replication system is
15 employed for cloning during the development of the plasmid and the second replication system is necessary for functioning in the ultimate host. In addition, one or more markers may be present, which have been described previously. Where integration is desired, the plasmid will desirably include a sequence homologous with the host genome.

20 The transformants can be isolated in accordance with conventional ways, usually employing a selection technique, which allows for selection of the desired organism as against unmodified organisms or transferring organisms, when present. The transformants then can be tested for pesticidal activity.

Suitable host cells, where the pesticide-containing cells will be treated to
25 prolong the activity of the δ -endotoxin in the cell when the then treated cell is applied to the environment of target pest(s), may include either prokaryotes or eukaryotes, normally being limited to those cells which do not produce substances toxic to higher organisms, such as mammals. However, organisms which produce substances toxic to higher organisms could be used, where the δ -endotoxin is unstable or the level of application sufficiently low as to avoid any possibility of toxicity
30 to a mammalian host. As hosts, of particular interest will be the prokaryotes

and the lower eukaryotes, such as fungi. Illustrative prokaryotes, both Gram-negative and -positive, include *Enterobacteriaceae*, such as *Escherichia*, *Erwinia*, *Shigella*, *Salmonella*, and *Proteus*; *Bacillaceae*; *Rhizobiceae*, such as *Rhizobium*; *Spirillaceae*, such as photobacterium, *Zymomonas*, *Serratia*, *Aeromonas*, *Vibrio*,
5 *Desulfovibrio*, *Spirillum*; *Lactobacillaceae*; phylloplane organisms such as members of the *Pseudomonadaceae* (including *Pseudomonas* spp. and *Acetobacter* spp.); *Azotobacteraceae* and *Nitrobacteraceae*; *Flavobacterium* spp.; members of the *Bacillaceae* such as *Lactobacillus* spp., *Bifidobacterium*, and *Bacillus* spp., and the like. Particularly preferred host cells include *Pseudomonas aeruginosa*, *Pseu-*
10 *domonas fluorescens*, *Bacillus thuringiensis*, *Escherichia coli*, *Bacillus subtilis*, and the like.

Among eukaryotes are fungi, such as *Phycomycetes* and *Ascomycetes*, which includes yeast, such as *Schizosaccharomyces*; and *Basidiomycetes*, *Rhodotorula*, *Aureobasidium*, *Sporobolomyces*, *Saccharomyces* spp., and *Sporobolomy-*
15 *ces* spp.

Characteristics of particular interest in selecting a host cell for purposes of production include ease of introducing the δ -endotoxin gene into the host, availability of expression systems, efficiency of expression, stability of the pesticide in the host, and the presence of auxiliary genetic capabilities. Characteristics of inter-
20 est for use as a pesticide microcapsule include protective qualities for the pesticide, such as thick cell walls, pigmentation, and intracellular packaging or formation of inclusion bodies; leaf affinity; lack of mammalian toxicity; attractiveness to pests for ingestion; ease of killing and fixing without damage to the δ -endotoxin; and the like. Other considerations include ease of formulation and handling, economics,
25 storage stability, and the like.

The cell will usually be intact and be substantially in the proliferative form when treated, rather than in a spore form, although in some instances spores may be employed. Treatment of the recombinant microbial cell can be done as disclosed *infra*. The treated cells generally will have enhanced structural stability
30 which will enhance resistance to environmental conditions.

Genes or other nucleic acid segments, as disclosed herein, can be inserted into host cells using a variety of techniques which are well known in the art. For example, a large number of cloning vectors comprising a replication system in *E. coli* and a marker that permits selection of the transformed cells are available for
5 preparation for the insertion of foreign genes into higher organisms, including plants. The vectors comprise, for example, pBR322, pUC series, M13mp series, pACYC184, *etc.* Accordingly, the sequence coding for the δ -endotoxin can be inserted into the vector at a suitable restriction site. The resulting plasmid is used for transformation into *E. coli*. The *E. coli* cells are cultivated in a suitable nutrient
10 medium, then harvested and lysed. The plasmid is recovered. Sequence analysis, restriction analysis, electrophoresis, and other biochemical-molecular biological methods are generally carried out as methods of analysis. After each manipulation, the DNA sequence used can be cleaved and joined to the next DNA sequence. Each plasmid sequence can be cloned in the same or other plasmids. Depending on
15 the method of inserting desired genes into the plant, other DNA sequences may be necessary.

Methods for DNA transformation of plant cells include *Agrobacterium*-mediated plant transformation, protoplast transformation, gene transfer into pollen, injection into reproductive organs, injection into immature embryos and particle
20 bombardment. Each of these methods has distinct advantages and disadvantages. Thus, one particular method of introducing genes into a particular plant strain may not necessarily be the most effective for another plant strain, but it is well known which methods are useful for a particular plant strain.

Suitable methods are believed to include virtually any method by which
25 DNA can be introduced into a cell, such as by *Agrobacterium* infection, direct delivery of DNA such as, for example, by PEG-mediated transformation of protoplasts (Omirulleh *et al.*, 1993), by desiccation/inhibition-mediated DNA uptake, by electroporation, by agitation with silicon carbide fibers, by acceleration of DNA coated particles, *etc.* In certain embodiments, acceleration methods are preferred
30 and include, for example, microprojectile bombardment and the like.

Technology for introduction of DNA into cells is well-known to those of skill in the art. Four general methods for delivering a gene into cells have been described: (1) chemical methods (Graham and van der Eb, 1973; Zatloukal *et al.*, 1992); (2) physical methods such as microinjection (Capecchi, 1980), electroporation (Wong and Neumann, 1982; Fromm *et al.*, 1985) and the gene gun (Johnston and Tang, 1994; Fynan *et al.*, 1993); (3) viral vectors (Clapp, 1993; Lu *et al.*, 1993; Eglitis and Anderson, 1988; Eglitis *et al.*, 1988); and (4) receptor-mediated mechanisms (Curiel *et al.*, 1991; 1992; Wagner *et al.*, 1992).

A large number of techniques are available for inserting DNA into a plant host cell. Those techniques include transformation with T-DNA using *Agrobacterium tumefaciens* or *Agrobacterium rhizogenes* as transformation agent, fusion, injection, or electroporation as well as other possible methods. If agrobacteria are used for the transformation, the DNA to be inserted has to be cloned into special plasmids, namely either into an intermediate vector or into a binary vector. The intermediate vectors can be integrated into the Ti or Ri plasmid by homologous recombination owing to sequences that are homologous to sequences in the T-DNA. The Ti or Ri plasmid also comprises the *vir* region necessary for the transfer of the T-DNA.

Intermediate vectors cannot replicate themselves in agrobacteria. The intermediate vector can be transferred into *Agrobacterium tumefaciens* by means of a helper plasmid (conjugation). Binary vectors can replicate themselves both in *E. coli* and in agrobacteria. They comprise a selection marker gene and a linker or polylinker which are framed by the right and left T-DNA border regions. They can be transformed directly into agrobacteria (Holsters *et al.*, 1978). The agrobacterium used as host cell is to comprise a plasmid carrying a *vir* region. The *vir* region is necessary for the transfer of the T-DNA into the plant cell. Additional T-DNA may be contained. The bacterium so transformed is used for the transformation of plant cells. Plant explants can advantageously be cultivated with *Agrobacterium tumefaciens* or *Agrobacterium rhizogenes* for the transfer of the DNA into the plant cell. Whole plants can then be regenerated from the infected plant material (for example, pieces of leaf, segments of stalk, roots, but also protoplasts or

suspension-cultivated cells) in a suitable medium, which may contain antibiotics or biocides for selection. The plants so obtained can then be tested for the presence of the inserted DNA. No special demands are made of the plasmids in the case of injection-and electroporation. It is possible to use ordinary plasmids, such as, for example, pUC derivatives. If, for example, the Ti or Ri plasmid is used for the transformation of the plant cell, then at least the right border, but often the right and the left border of the Ti or Ri plasmid T-DNA, has to be joined as the flanking region of the genes to be inserted. The use of T-DNA for the transformation of plant cells has been intensively researched and sufficiently described in Eur. Pat. Appl. No. EP 120 516; Hockema (1985); An *et al.*, 1985, Herrera-Estrella *et al.*, (1983), Bevan *et al.*, (1983), and Klee *et al.*, (1985).

A particularly useful Ti plasmid cassette vector for transformation of dicotyledonous plants consists of the enhanced CaMV35S promoter (EN35S) and the 3' end including polyadenylation signals from a soybean gene encoding the α' -subunit of β -conglycinin. Between these two elements is a multilinker containing multiple restriction sites for the insertion of genes of interest.

The vector preferably contains a segment of pBR322 which provides an origin of replication in *E. coli* and a region for homologous recombination with the disarmed T-DNA in *Agrobacterium* strain ACO; the *oriV* region from the broad host range plasmid RK1; the streptomycin/spectinomycin resistance gene from Tn7; and a chimeric NPTII gene, containing the CaMV35S promoter and the nopaline synthase (NOS) 3' end, which provides kanamycin resistance in transformed plant cells.

Optionally, the enhanced CaMV35S promoter may be replaced with the 1.5 kb mannopine synthase (MAS) promoter (Velten *et al.*, 1984). After incorporation of a DNA construct into the vector, it is introduced into *A. tumefaciens* strain ACO which contains a disarmed Ti plasmid. Cointegrate Ti plasmid vectors are selected and subsequently may be used to transform a dicotyledonous plant.

A. tumefaciens ACO is a disarmed strain similar to pTiB6SE described by Fraley *et al.* (1985). For construction of ACO the starting *Agrobacterium* strain was the strain A208 which contains a nopaline-type Ti plasmid. The Ti plasmid

was disarmed in a manner similar to that described by Fraley *et al.* (1985) so that essentially all of the native T-DNA was removed except for the left border and a few hundred base pairs of T-DNA inside the left border. The remainder of the T-DNA extending to a point just beyond the right border was replaced with a novel
5 piece of DNA including (from left to right) a segment of pBR322, the *oriV* region from plasmid RK2, and the kanamycin resistance gene from Tn601. The pBR322 and *oriV* segments are similar to these segments and provide a region of homology for cointegrate formation.

Once the inserted DNA has been integrated in the genome, it is relatively
10 stable there and, as a rule, does not come out again. It normally contains a selection marker that confers on the transformed plant cells resistance to a biocide or an antibiotic, such as kanamycin, G 418, bleomycin, hygromycin, or chloramphenicol, *inter alia*. The individually employed marker should accordingly permit the selection of transformed cells rather than cells that do not contain the inserted DNA.

15

4.11.1 ELECTROPORATION

The application of brief, high-voltage electric pulses to a variety of animal and plant cells leads to the formation of nanometer-sized pores in the plasma membrane. DNA is taken directly into the cell cytoplasm either through these pores or
20 as a consequence of the redistribution of membrane components that accompanies closure of the pores. Electroporation can be extremely efficient and can be used both for transient expression of clones genes and for establishment of cell lines that carry integrated copies of the gene of interest. Electroporation, in contrast to calcium phosphate-mediated transfection and protoplast fusion, frequently gives rise
25 to cell lines that carry one, or at most a few, integrated copies of the foreign DNA.

The introduction of DNA by means of electroporation, is well-known to those of skill in the art. In this method, certain cell wall-degrading enzymes, such as pectin-degrading enzymes, are employed to render the target recipient cells more susceptible to transformation by electroporation than untreated cells. Alternatively,
30 recipient cells are made more susceptible to transformation, by mechanical wounding. To effect transformation by electroporation one may employ either fri-

able tissues such as a suspension culture of cells, or embryogenic callus, or alternatively, one may transform immature embryos or other organized tissues directly. One would partially degrade the cell walls of the chosen cells by exposing them to pectin-degrading enzymes (pectolyases) or mechanically wounding in a controlled manner. Such cells would then be recipient to DNA transfer by electroporation, which may be carried out at this stage, and transformed cells then identified by a suitable selection or screening protocol dependent on the nature of the newly incorporated DNA.

4.11.2 MICROPROJECTILE BOMBARDMENT

A further advantageous method for delivering transforming DNA segments to plant cells is microprojectile bombardment. In this method, particles may be coated with nucleic acids and delivered into cells by a propelling force. Exemplary particles include those comprised of tungsten, gold, platinum, and the like.

An advantage of microprojectile bombardment, in addition to it being an effective means of reproducibly stably transforming monocots, is that neither the isolation of protoplasts (Cristou *et al.*, 1988) nor the susceptibility to *Agrobacterium* infection is required. An illustrative embodiment of a method for delivering DNA into maize cells by acceleration is a Biolistics Particle Delivery System, which can be used to propel particles coated with DNA or cells through a screen, such as a stainless steel or Nytex screen, onto a filter surface covered with corn cells cultured in suspension. The screen disperses the particles so that they are not delivered to the recipient cells in large aggregates. It is believed that a screen intervening between the projectile apparatus and the cells to be bombarded reduces the size of projectiles aggregate and may contribute to a higher frequency of transformation by reducing damage inflicted on the recipient cells by projectiles that are too large.

For the bombardment, cells in suspension are preferably concentrated on filters or solid culture medium. Alternatively, immature embryos or other target cells may be arranged on solid culture medium. The cells to be bombarded are positioned at an appropriate distance below the macroprojectile stopping plate. If

desired, one or more screens are also positioned between the acceleration device and the cells to be bombarded. Through the use of techniques set forth herein one may obtain up to 1000 or more foci of cells transiently expressing a marker gene. The number of cells in a focus which express the exogenous gene product 48 hours post-bombardment often range from 1 to 10 and average 1 to 3.

In bombardment transformation, one may optimize the prebombardment culturing conditions and the bombardment parameters to yield the maximum numbers of stable transformants. Both the physical and biological parameters for bombardment are important in this technology. Physical factors are those that involve manipulating the DNA/microprojectile precipitate or those that affect the flight and velocity of either the macro- or microprojectiles. Biological factors include all steps involved in manipulation of cells before and immediately after bombardment, the osmotic adjustment of target cells to help alleviate the trauma associated with bombardment, and also the nature of the transforming DNA, such as linearized DNA or intact supercoiled plasmids. It is believed that pre-bombardment manipulations are especially important for successful transformation of immature embryos.

Accordingly, it is contemplated that one may wish to adjust various of the bombardment parameters in small scale studies to fully optimize the conditions. One may particularly wish to adjust physical parameters such as gap distance, flight distance, tissue distance, and helium pressure. One may also minimize the trauma reduction factors (TRFs) by modifying conditions which influence the physiological state of the recipient cells and which may therefore influence transformation and integration efficiencies. For example, the osmotic state, tissue hydration and the subculture stage or cell cycle of the recipient cells may be adjusted for optimum transformation. The execution of other routine adjustments will be known to those of skill in the art in light of the present disclosure.

4.11.3 AGROBACTERIUM-MEDIATED TRANSFER

Agrobacterium-mediated transfer is a widely applicable system for introducing genes into plant cells because the DNA can be introduced into whole plant

tissues, thereby bypassing the need for regeneration of an intact plant from a protoplast. The use of *Agrobacterium*-mediated plant integrating vectors to introduce DNA into plant cells is well known in the art. See, for example, the methods described (Fraley *et al.*, 1985; Rogers *et al.*, 1987). Further, the integration of the Ti-DNA is a relatively precise process resulting in few rearrangements. The region of DNA to be transferred is defined by the border sequences, and intervening DNA is usually inserted into the plant genome as described (Spielmann *et al.*, 1986; Jorgensen *et al.*, 1987).

Modern *Agrobacterium* transformation vectors are capable of replication in *E. coli* as well as *Agrobacterium*, allowing for convenient manipulations as described (Klee *et al.*, 1985). Moreover, recent technological advances in vectors for *Agrobacterium*-mediated gene transfer have improved the arrangement of genes and restriction sites in the vectors to facilitate construction of vectors capable of expressing various polypeptide coding genes. The vectors described (Rogers *et al.*, 1987), have convenient multi-linker regions flanked by a promoter and a polyadenylation site for direct expression of inserted polypeptide coding genes and are suitable for present purposes. In addition, *Agrobacterium* containing both armed and disarmed Ti genes can be used for the transformations. In those plant strains where *Agrobacterium*-mediated transformation is efficient, it is the method of choice because of the facile and defined nature of the gene transfer.

Agrobacterium-mediated transformation of leaf disks and other tissues such as cotyledons and hypocotyls appears to be limited to plants that *Agrobacterium* naturally infects. *Agrobacterium*-mediated transformation is most efficient in dicotyledonous plants. Few monocots appear to be natural hosts for *Agrobacterium*, although transgenic plants have been produced in asparagus using *Agrobacterium* vectors as described (Bytebier *et al.*, 1987). Therefore, commercially important cereal grains such as rice, corn, and wheat must usually be transformed using alternative methods. However, as mentioned above, the transformation of asparagus using *Agrobacterium* can also be achieved (see, for example, Bytebier *et al.*, 1987).

A transgenic plant formed using *Agrobacterium* transformation methods typically contains a single gene on one chromosome. Such transgenic plants can

be referred to as being heterozygous for the added gene. However, inasmuch as use of the word "heterozygous" usually implies the presence of a complementary gene at the same locus of the second chromosome of a pair of chromosomes, and there is no such gene in a plant containing one added gene as here, it is believed
5 that a more accurate name for such a plant is an independent segregant, because the added, exogenous gene segregates independently during mitosis and meiosis.

More preferred is a transgenic plant that is homozygous for the added structural gene; *i.e.*, a transgenic plant that contains two added genes, one gene at the same locus on each chromosome of a chromosome pair. A homozygous trans-
10 genic plant can be obtained by sexually mating (selfing) an independent segregant transgenic plant that contains a single added gene, germinating some of the seed produced and analyzing the resulting plants produced for enhanced carboxylase activity relative to a control (native, non-transgenic) or an independent segregant transgenic plant.

15 It is to be understood that two different transgenic plants can also be mated to produce offspring that contain two independently segregating added, exogenous genes. Selfing of appropriate progeny can produce plants that are homozygous for both added, exogenous genes that encode a polypeptide of interest. Back-crossing to a parental plant and out-crossing with a non-transgenic plant are also contemplated.
20

Transformation of plant protoplasts can be achieved using methods based on calcium phosphate precipitation, polyethylene glycol treatment, electroporation, and combinations of these treatments (see, *e.g.*, Potrykus *et al.*, 1985; Lorz *et al.*, 1985; Fromm *et al.*, 1985; Uchimiya *et al.*, 1986; Callis *et al.*, 1987; Marcotte *et al.*, 1988).
25

Application of these systems to different plant strains depends upon the ability to regenerate that particular plant strain from protoplasts. Illustrative methods for the regeneration of cereals from protoplasts are described (Fujimura *et al.*, 1985; Toriyama *et al.*, 1986; Yamada *et al.*, 1986; Abdullah *et al.*, 1986).

30 To transform plant strains that cannot be successfully regenerated from protoplasts, other ways to introduce DNA into intact cells or tissues can be utilized.

For example, regeneration of cereals from immature embryos or explants can be effected as described (Vasil, 1988). In addition, "particle gun" or high-velocity microprojectile technology can be utilized (Vasil, 1992).

Using that latter technology, DNA is carried through the cell wall and into
5 the cytoplasm on the surface of small metal particles as described (Klein *et al.*, 1987; Klein *et al.*, 1988; McCabe *et al.*, 1988). The metal particles penetrate through several layers of cells and thus allow the transformation of cells within tissue explants.

10 4.11.4 GENE EXPRESSION IN PLANTS

Although great progress has been made in recent years with respect to preparation of transgenic plants which express bacterial proteins such as *B. thuringiensis* crystal proteins, the results of expressing native bacterial genes in plants are often disappointing. Unlike microbial genetics, little was known by
15 early plant geneticists about the factors which affected heterologous expression of foreign genes in plants. In recent years, however, several potential factors have been implicated as responsible in varying degrees for the level of protein expression from a particular coding sequence. For example, scientists now know that maintaining a significant level of a particular mRNA in the cell is indeed a critical
20 factor. Unfortunately, the causes for low steady state levels of mRNA encoding foreign proteins are many. First, full length RNA synthesis may not occur at a high frequency. This could, for example, be caused by the premature termination of RNA during transcription or due to unexpected mRNA processing during transcription. Second, full length RNA may be produced in the plant cell, but then
25 processed (splicing, polyA addition) in the nucleus in a fashion that creates a non-functional mRNA. If the RNA is not properly synthesized, terminated and polyadenylated, it cannot move to the cytoplasm for translation. Similarly, in the cytoplasm, if mRNAs have reduced half lives (which are determined by their primary or secondary sequence) insufficient protein product will be produced. In addition,
30 there is an effect, whose magnitude is uncertain, of translational efficiency on mRNA half-life. In addition, every RNA molecule folds into a particular structure,

or perhaps family of structures, which is determined by its sequence. The particular structure of any RNA might lead to greater or lesser stability in the cytoplasm. Structure *per se* is probably also a determinant of mRNA processing in the nucleus. Unfortunately, it is impossible to predict, and nearly impossible to determine, the structure of any RNA (except for tRNA) *in vitro* or *in vivo*. However, it is likely that dramatically changing the sequence of an RNA will have a large effect on its folded structure. It is likely that structure *per se* or particular structural features also have a role in determining RNA stability.

To overcome these limitations in foreign gene expression, researchers have identified particular sequences and signals in RNAs that have the potential for having a specific effect on RNA stability. In certain embodiments of the invention, therefore, there is a desire to optimize expression of the disclosed nucleic acid segments *in planta*. One particular method of doing so, is by alteration of the bacterial gene to remove sequences or motifs which decrease expression in a transformed plant cell. The process of engineering a coding sequence for optimal expression *in planta* is often referred to as "plantizing" a DNA sequence.

Particularly problematic sequences are those which are A+T rich. Unfortunately, since *B. thuringiensis* has an A+T rich genome, native crystal protein gene sequences must often be modified for optimal expression in a plant. The sequence motif ATTTA (or AUUUA as it appears in RNA) has been implicated as a destabilizing sequence in mammalian cell mRNA (Shaw and Kamen, 1986). Many short lived mRNAs have A+T rich 3' untranslated regions, and these regions often have the ATTTA sequence, sometimes present in multiple copies or as multimers (e.g., ATTTATTTA...). Shaw and Kamen showed that the transfer of the 3' end of an unstable mRNA to a stable RNA (globin or VA1) decreased the stable RNA's half life dramatically. They further showed that a pentamer of ATTTA had a profound destabilizing effect on a stable message, and that this signal could exert its effect whether it was located at the 3' end or within the coding sequence. However, the number of ATTTA sequences and/or the sequence context in which they occur also appear to be important in determining whether they function as destabilizing sequences. Shaw and Kamen showed that a trimer of ATTTA had much less

effect than a pentamer on mRNA stability and a dimer or a monomer had no effect on stability (Shaw and Kamen, 1987). Note that multimers of ATTTA such as a pentamer automatically create an A+T rich region. This was shown to be a cytoplasmic effect, not nuclear. In other unstable mRNAs, the ATTTA sequence may
5 be present in only a single copy, but it is often contained in an A+T rich region. From the animal cell data collected to date, it appears that ATTTA at least in some contexts is important in stability, but it is not yet possible to predict which occurrences of ATTTA are destabilizing elements or whether any of these effects are likely to be seen in plants.

10 Some studies on mRNA degradation in animal cells also indicate that RNA degradation may begin in some cases with nucleolytic attack in A+T rich regions. It is not clear if these cleavages occur at ATTTA sequences. There are also examples of mRNAs that have differential stability depending on the cell type in which they are expressed or on the stage within the cell cycle at which they are expressed.
15 For example, histone mRNAs are stable during DNA synthesis but unstable if DNA synthesis is disrupted. The 3' end of some histone mRNAs seems to be responsible for this effect (Pandey and Marzluff, 1987). It does not appear to be mediated by ATTTA, nor is it clear what controls the differential stability of this mRNA. Another example is the differential stability of IgG mRNA in B lymphocytes during B cell maturation (Genovese and Milcarek, 1988). A final example is
20 the instability of a mutant β -thalassemic globin mRNA. In bone marrow cells, where this gene is normally expressed, the mutant mRNA is unstable, while the wild-type mRNA is stable. When the mutant gene is expressed in HeLa or L cells *in vitro*, the mutant mRNA shows no instability (Lim *et al.*, 1988). These examples all provide evidence that mRNA stability can be mediated by cell type or cell
25 cycle specific factors. Furthermore this type of instability is not yet associated with specific sequences. Given these uncertainties, it is not possible to predict which RNAs are likely to be unstable in a given cell. In addition, even the ATTTA motif may act differentially depending on the nature of the cell in which the RNA
30 is present. Shaw and Kamen (1987) have reported that activation of protein kinase C can block degradation mediated by ATTTA.

The addition of a polyadenylate string to the 3' end is common to most eukaryotic mRNAs, both plant and animal. The currently accepted view of polyA addition is that the nascent transcript extends beyond the mature 3' terminus. Contained within this transcript are signals for polyadenylation and proper 3' end formation. This processing at the 3' end involves cleavage of the mRNA and addition of polyA to the mature 3' end. By searching for consensus sequences near the polyA tract in both plant and animal mRNAs, it has been possible to identify consensus sequences that apparently are involved in polyA addition and 3' end cleavage. The same consensus sequences seem to be important to both of these processes. These signals are typically a variation on the sequence AATAAA. In animal cells, some variants of this sequence that are functional have been identified; in plant cells there seems to be an extended range of functional sequences (Wickens and Stephenson, 1984; Dean *et al.*, 1986). Because all of these consensus sequences are variations on AATAAA, they all are A+T rich sequences. This sequence is typically found 15 to 20 bp before the polyA tract in a mature mRNA. Studies in animal cells indicate that this sequence is involved in both polyA addition and 3' maturation. Site directed mutations in this sequence can disrupt these functions (Conway and Wickens, 1988; Wickens *et al.*, 1987). However, it has also been observed that sequences up to 50 to 100 bp 3' to the putative polyA signal are also required; *i.e.*, a gene that has a normal AATAAA but has been replaced or disrupted downstream does not get properly polyadenylated (Gil and Proudfoot, 1984; Sadofsky and Alwine, 1984; McDevitt *et al.*, 1984). That is, the polyA signal itself is not sufficient for complete and proper processing. It is not yet known what specific downstream sequences are required in addition to the polyA signal, or if there is a specific sequence that has this function. Therefore, sequence analysis can only identify potential polyA signals.

In naturally occurring mRNAs that are normally polyadenylated, it has been observed that disruption of this process, either by altering the polyA signal or other sequences in the mRNA, profound effects can be obtained in the level of functional mRNA. This has been observed in several naturally occurring mRNAs, with results that are gene-specific so far.

It has been shown that in natural mRNAs proper polyadenylation is important in mRNA accumulation, and that disruption of this process can effect mRNA levels significantly. However, insufficient knowledge exists to predict the effect of changes in a normal gene. In a heterologous gene, it is even harder to predict the consequences. However, it is possible that the putative sites identified are dysfunctional. That is, these sites may not act as proper polyA sites, but instead function as aberrant sites that give rise to unstable mRNAs.

In animal cell systems, AATAAA is by far the most common signal identified in mRNAs upstream of the polyA, but at least four variants have also been found (Wickens and Stephenson, 1984). In plants, not nearly so much analysis has been done, but it is clear that multiple sequences similar to AATAAA can be used. The plant sites in Table 5 called major or minor refer only to the study of Dean *et al.* (1986) which analyzed only three types of plant gene. The designation of polyadenylation sites as major or minor refers only to the frequency of their occurrence as functional sites in naturally occurring genes that have been analyzed. In the case of plants this is a very limited database. It is hard to predict with any certainty that a site designated major or minor is more or less likely to function partially or completely when found in a heterologous gene such as those encoding the crystal proteins of the present invention.

TABLE 5
POLYADENYLATION SITES IN PLANT GENES

PA	AATAAA	Major consensus site
P1A	AATAAT	Major plant site
P2A	AACCAA	Minor plant site
P3A	ATATAA	"
P4A	AATCAA	"
P5A	ATACTA	"
P6A	ATAAAA	"
P7A	ATGAAA	"
P8A	AAGCAT	"
P9A	ATTAAT	"
P10A	ATACAT	"
P11A	AAAATA	"
P12A	ATTAAA	Minor animal site
P13A	AATTAA	"
P14A	AATACA	"
P15A	CATAAA	"

5 The present invention provides a method for preparing synthetic plant genes which genes express their protein product at levels significantly higher than the wild-type genes which were commonly employed in plant transformation heretofore. In another aspect, the present invention also provides novel synthetic plant genes which encode non-plant proteins.

10 As described above, the expression of native *B. thuringiensis* genes in plants is often problematic. The nature of the coding sequences of *B. thuringiensis* genes distinguishes them from plant genes as well as many other heterologous genes expressed in plants. In particular, *B. thuringiensis* genes are very rich (~62%) in adenine (A) and thymine (T) while plant genes and most other bacterial genes which have been expressed in plants are on the order of 45-55% A+T.

Due to the degeneracy of the genetic code and the limited number of codon choices for any amino acid, most of the "excess" A+T of the structural coding sequences of some *Bacillus* species are found in the third position of the codons. That is, genes of some *Bacillus* species have A or T as the third nucleotide in many
5 codons. Thus A+T content in part can determine codon usage bias. In addition, it is clear that genes evolve for maximum function in the organism in which they evolve. This means that particular nucleotide sequences found in a gene from one organism, where they may play no role except to code for a particular stretch of amino acids, have the potential to be recognized as gene control elements in another organism (such as transcriptional promoters or terminators, polyA addition
10 sites, intron splice sites, or specific mRNA degradation signals). It is perhaps surprising that such misread signals are not a more common feature of heterologous gene expression, but this can be explained in part by the relatively homogeneous A+T content (~50%) of many organisms. This A+T content plus the nature of the
15 genetic code put clear constraints on the likelihood of occurrence of any particular oligonucleotide sequence. Thus, a gene from *E. coli* with a 50% A+T content is much less likely to contain any particular A+T rich segment than a gene from *B. thuringiensis*.

Typically, to obtain high-level expression of the S-endotoxin genes in
20 plants, existing structural coding sequence ("structural gene") which codes for the S-endotoxin are modified by removal of ATTTA sequences and putative polyadenylation signals by site directed mutagenesis of the DNA comprising the structural gene. It is most preferred that substantially all the polyadenylation signals and ATTTA sequences are removed although enhanced expression levels are observed with only partial removal of either of the above identified sequences. Alternately if a synthetic gene is prepared which codes for the expression of the subject protein, codons are selected to avoid the ATTTA sequence and putative polyadenylation signals. For purposes of the present invention putative polyadenylation signals include, but are not necessarily limited to, AATAAA, AATAAT,
25 AACCAA, ATATAA, AATCAA, ATACTA, ATAAAA, ATGAAA, AAGCAT, ATTAAT, ATACAT, AAAATA, ATTAAA, AATTAA, AATACA and
30

CATAAA. In replacing the ATTTA sequences and polyadenylation signals, codons are preferably utilized which avoid the codons which are rarely found in plant genomes.

The selected DNA sequence is scanned to identify regions with greater than
5 four consecutive adenine (A) or thymine (T) nucleotides. The A+T regions are scanned for potential plant polyadenylation signals. Although the absence of five or more consecutive A or T nucleotides eliminates most plant polyadenylation signals, if there are more than one of the minor polyadenylation signals identified within ten nucleotides of each other, then the nucleotide sequence of this region is
10 preferably altered to remove these signals while maintaining the original encoded amino acid sequence.

The second step is to consider the about 15 to about 30 or so nucleotide residues surrounding the A+T rich region identified in step one. If the A+T content of the surrounding region is less than 80%, the region should be examined for
15 polyadenylation signals. Alteration of the region based on polyadenylation signals is dependent upon (1) the number of polyadenylation signals present and (2) presence of a major plant polyadenylation signal.

The extended region is examined for the presence of plant polyadenylation signals. The polyadenylation signals are removed by site-directed mutagenesis of
20 the DNA sequence. The extended region is also examined for multiple copies of the ATTTA sequence which are also removed by mutagenesis.

It is also preferred that regions comprising many consecutive A+T bases or G+C bases are disrupted since these regions are predicted to have a higher likelihood to form hairpin structure due to self-complementarity. Therefore, insertion of
25 heterogeneous base pairs would reduce the likelihood of self-complementary secondary structure formation which are known to inhibit transcription and/or translation in some organisms. In most cases, the adverse effects may be minimized by using sequences which do not contain more than five consecutive A+T or G+C.

4.11.5 SYNTHETIC OLIGONUCLEOTIDES FOR MUTAGENESIS

When oligonucleotides are used in the mutagenesis, it is desirable to maintain the proper amino acid sequence and reading frame, without introducing common restriction sites such as *Bgl*III, *Hind*III, *Sac*I, *Kpn*I, *Eco*RI, *Nco*I, *Pst*I and *Sall* into the modified gene. These restriction sites are found in poly-linker insertion sites of many cloning vectors. Of course, the introduction of new polyadenylation signals, ATTTA sequences or consecutive stretches of more than five A+T or G+C, should also be avoided. The preferred size for the oligonucleotides is about 40 to about 50 bases, but fragments ranging from about 18 to about 100 bases have been utilized. In most cases, a minimum of about 5 to about 8 base pairs of homology to the template DNA on both ends of the synthesized fragment are maintained to insure proper hybridization of the primer to the template. The oligonucleotides should avoid sequences longer than five base pairs A+T or G+C. Codons used in the replacement of wild-type codons should preferably avoid the TA or CG doublet wherever possible. Codons are selected from a plant preferred codon table (such as Table 6 below) so as to avoid codons which are rarely found in plant genomes, and efforts should be made to select codons to preferably adjust the G+C content to about 50%.

TABLE 6
PREFERRED CODON USAGE IN PLANTS

Amino Acid	Codon	Percent Usage in Plants
ARG	CGA	7
	CGC	11
	CGG	5
	CGU	25
	AGA	29
	AGG	23
LEU	CUA	8
	CUC	20
	CUG	10
	CUU	28
	UUA	5
	UUG	30
SER	UCA	14
	UCC	26
	UCG	3
	UCU	21
	AGC	21
	AGU	15
THR	ACA	21
	ACC	41
	ACG	7
	ACU	31
PRO	CCA	45
	CCC	19
	CCG	9

TABLE 6 (CONT'D)

Amino Acid	Codon	Percent Usage in Plants
	CCU	26
ALA	GCA	23
	GCC	32
	GCG	3
	GCU	41
GLY	GGA	32
	GGC	20
	GGG	11
	GGU	37
ILE	AUA	12
	AUC	45
	AUU	43
VAL	GUA	9
	GUC	20
	GUG	28
	GUU	43
LYS	AAA	36
	AAG	64
ASN	AAC	72
	AAU	28
GLN	CAA	64
	CAG	36
HIS	CAC	65
	CAU	35
GLU	GAA	48
	GAG	52
ASP	GAC	48
	GAU	52
TYR	UAC	68
	UAU	32

TABLE 6 (CONT'D)

Amino Acid	Codon	Percent Usage in Plants
CYS	UGC	78
	UGU	22
PHE	UUC	56
	UUU	44
MET	AUG	100
TRP	UGG	100

Regions with many consecutive A+T bases or G+C bases are predicted to have a higher likelihood to form hairpin structures due to self-complementarity.

- 5 Disruption of these regions by the insertion of heterogeneous base pairs is preferred and should reduce the likelihood of the formation of self-complementary secondary structures such as hairpins which are known in some organisms to inhibit transcription (transcriptional terminators) and translation (attenuators).

- 10 Alternatively, a completely synthetic gene for a given amino acid sequence can be prepared, with regions of five or more consecutive A+T or G+C nucleotides being avoided. Codons are selected avoiding the TA and CG doublets in codons whenever possible. Codon usage can be normalized against a plant preferred codon usage table (such as Table 6) and the G+C content preferably adjusted to about 50%. The resulting sequence should be examined to ensure that there are
- 15 minimal putative plant polyadenylation signals and ATTTA sequences. Restriction sites found in commonly used cloning vectors are also preferably avoided. However, placement of several unique restriction sites throughout the gene is useful for analysis of gene expression or construction of gene variants.

20 4.11.6 "PLANTIZED" GENE CONSTRUCTS

The expression of a plant gene which exists in double-stranded DNA form involves transcription of messenger RNA (mRNA) from one strand of the DNA by RNA polymerase enzyme, and the subsequent processing of the mRNA primary

transcript inside the nucleus. This processing involves a 3' non-translated region which adds polyadenylate nucleotides to the 3' end of the RNA. Transcription of DNA into mRNA is regulated by a region of DNA usually referred to as the "promoter." The promoter region contains a sequence of bases that signals RNA
5 polymerase to associate with the DNA and to initiate the transcription of mRNA using one of the DNA strands as a template to make a corresponding strand of RNA.

A number of promoters which are active in plant cells have been described in the literature. These include the nopaline synthase (NOS) and octopine synthase
10 (OCS) promoters (which are carried on tumor-inducing plasmids of *Agrobacterium tumefaciens*), the Cauliflower Mosaic Virus (CaMV) 19S and 35S promoters, the light-inducible promoter from the small subunit of ribulose bis-phosphate carboxylase (ssRUBISCO, a very abundant plant polypeptide) and the mannopine synthase (MAS) promoter (Velten *et al.*, 1984 and Velten and Schell, 1985). All of these
15 promoters have been used to create various types of DNA constructs which have been expressed in plants (see *e.g.*, Int. Pat. Appl. Publ. No. WO 84/02913).

Promoters which are known or are found to cause transcription of RNA in plant cells can be used in the present invention. Such promoters may be obtained from plants or plant viruses and include, but are not limited to, the CaMV35S pro-
20 moter and promoters isolated from plant genes such as ssRUBISCO genes. As described below, it is preferred that the particular promoter selected should be capable of causing sufficient expression to result in the production of an effective amount of protein.

The promoters used in the DNA constructs (*i.e.* chimeric plant genes) of the
25 present invention may be modified, if desired, to affect their control characteristics. For example, the CaMV35S promoter may be ligated to the portion of the ssRUBISCO gene that represses the expression of ssRUBISCO in the absence of light, to create a promoter which is active in leaves but not in roots. The resulting chimeric promoter may be used as described herein. For purposes of this description,
30 the phrase "CaMV35S" promoter thus includes variations of CaMV35S promoter, *e.g.*, promoters derived by means of ligation with operator regions, random or con-

trolled mutagenesis, *etc.* Furthermore, the promoters may be altered to contain multiple "enhancer sequences" to assist in elevating gene expression.

The RNA produced by a DNA construct of the present invention also contains a 5' non-translated leader sequence. This sequence can be derived from the promoter selected to express the gene, and can be specifically modified so as to increase translation of the mRNA. The 5' non-translated regions can also be obtained from viral RNA's, from suitable eukaryotic genes, or from a synthetic gene sequence. The present invention is not limited to constructs, as presented in the following examples. Rather, the non-translated leader sequence can be part of the 5' end of the non-translated region of the coding sequence for the virus coat protein, or part of the promoter sequence, or can be derived from an unrelated promoter or coding sequence. In any case, it is preferred that the sequence flanking the initiation site conform to the translational consensus sequence rules for enhanced translation initiation reported by Kozak (1984).

The *cry* DNA constructs of the present invention may also contain one or more modified or fully-synthetic structural coding sequences which have been changed to enhance the performance of the *cry* gene in plants. The structural genes of the present invention may optionally encode a fusion protein comprising an amino-terminal chloroplast transit peptide or secretory signal sequence.

The DNA construct also contains a 3' non-translated region. The 3' non-translated region contains a polyadenylation signal which functions in plants to cause the addition of polyadenylate nucleotides to the 3' end of the viral RNA. Examples of suitable 3' regions are (1) the 3' transcribed, non-translated regions containing the polyadenylation signal of *Agrobacterium* tumor-inducing (Ti) plasmid genes, such as the nopaline synthase (NOS) gene, and (2) plant genes like the soybean storage protein (7S) genes and the small subunit of the RuBP carboxylase (E9) gene.

4.12 METHODS FOR PRODUCING INSECT-RESISTANT TRANSGENIC PLANTS

By transforming a suitable host cell, such as a plant cell, with a recombinant *cry** gene-containing segment, the expression of the encoded crystal protein

(i.e., a bacterial crystal protein or polypeptide having insecticidal activity against coleopterans) can result in the formation of insect-resistant plants.

By way of example, one may utilize an expression vector containing a coding region for a *B. thuringiensis* crystal protein and an appropriate selectable
5 marker to transform a suspension of embryonic plant cells, such as wheat or corn cells using a method such as particle bombardment (Maddock *et al.*, 1991; Vasil *et al.*, 1992) to deliver the DNA coated on microprojectiles into the recipient cells. Transgenic plants are then regenerated from transformed embryonic calli that express the insecticidal proteins.

10 The formation of transgenic plants may also be accomplished using other methods of cell transformation which are known in the art such as *Agrobacterium*-mediated DNA transfer (Fraley *et al.*, 1983). Alternatively, DNA can be introduced into plants by direct DNA transfer into pollen (Zhou *et al.*, 1983; Hess, 1987; Luo *et al.*, 1988), by injection of the DNA into reproductive organs of a
15 plant (Pena *et al.*, 1987), or by direct injection of DNA into the cells of immature embryos followed by the rehydration of desiccated embryos (Neuhaus *et al.*, 1987; Benbrook *et al.*, 1986).

The regeneration, development, and cultivation of plants from single plant protoplast transformants or from various transformed explants is well known in the
20 art (Weissbach and Weissbach, 1988). This regeneration and growth process typically includes the steps of selection of transformed cells, culturing those individualized cells through the usual stages of embryonic development through the rooted plantlet stage. Transgenic embryos and seeds are similarly regenerated. The resulting transgenic rooted shoots are thereafter planted in an appropriate plant
25 growth medium such as soil.

The development or regeneration of plants containing the foreign, exogenous gene that encodes a polypeptide of interest introduced by *Agrobacterium* from leaf explants can be achieved by methods well known in the art such as described (Horsch *et al.*, 1985). In this procedure, transformants are cultured in the presence
30 of a selection agent and in a medium that induces the regeneration of shoots in the plant strain being transformed as described (Fraley *et al.*, 1983).

This procedure typically produces shoots within two to four months and those shoots are then transferred to an appropriate root-inducing medium containing the selective agent and an antibiotic to prevent bacterial growth. Shoots that rooted in the presence of the selective agent to form plantlets are then transplanted
5 to soil or other media to allow the production of roots. These procedures vary depending upon the particular plant strain employed, such variations being well known in the art.

Preferably, the regenerated plants are self-pollinated to provide homozygous transgenic plants, as discussed before. Otherwise, pollen obtained from the
10 regenerated plants is crossed to seed-grown plants of agronomically important, preferably inbred lines. Conversely, pollen from plants of those important lines is used to pollinate regenerated plants. A transgenic plant of the present invention containing a desired polypeptide is cultivated using methods well known to one skilled in the art.

15 Such plants can form germ cells and transmit the transformed trait(s) to progeny plants. Likewise, transgenic plants can be grown in the normal manner and crossed with plants that have the same transformed hereditary factors or other hereditary factors. The resulting hybrid individuals have the corresponding phenotypic properties. A transgenic plant of this invention thus has an increased amount
20 of a coding region (*e.g.*, a mutated *cry* gene) that encodes the mutated Cry polypeptide of interest. A preferred transgenic plant is an independent segregant and can transmit that gene and its activity to its progeny. A more preferred transgenic plant is homozygous for that gene, and transmits that gene to all of its offspring on sexual mating.

25 Seed from a transgenic plant may be grown in the field or greenhouse, and resulting sexually mature transgenic plants are self-pollinated to generate true breeding plants. The progeny from these plants become true breeding lines that are evaluated for, by way of example, increased insecticidal capacity against coleopteran insects, preferably in the field, under a range of environmental conditions.
30 The inventors contemplate that the present invention will find particular utility in the creation of transgenic plants of commercial interest including various grasses,

grains, fibers, tubers, legumes, ornamental plants, cacti, succulents, fruits, berries, and vegetables, as well as a number of nut- and fruit-bearing trees and plants.

4.13 METHODS FOR PRODUCING COMBINATORIAL CRY3* VARIANTS

5 Crystal protein mutants containing substitutions in one or more domains may be constructed *via* a number of techniques. For instance, sequences of highly related genes can be readily shuffled using the PCRTM-based technique described by Stemmer (1994). Alternatively, if suitable restriction sites are available, the mutations of one *cry* gene may be combined with the mutations of a second *cry*
10 gene by routine subcloning methodologies. If a suitable restriction site is not available, one may be generated by oligonucleotide directed mutagenesis using any number of procedures known to those skilled in the art. Alternatively, splice-overlap extension PCRTM (Horton *et al.*, 1989) may be used to combine mutations in different regions of a crystal protein. In this procedure, overlapping DNA frag-
15 ments generated by the PCRTM and containing different mutations within their unique sequences may be annealed and used as a template for amplification using flanking primers to generate a hybrid gene sequence. Finally, *cry** mutants may be combined by simply using one *cry* mutant as a template for oligonucleotide-directed mutagenesis using any number of protocols such as those described
20 herein.

4.14 ISOLATING HOMOLOGOUS GENE AND GENE FRAGMENTS

The genes and δ -endotoxins according to the subject invention include not only the full length sequences disclosed herein but also fragments of these se-
25 quences, or fusion proteins, which retain the characteristic insecticidal activity of the sequences specifically exemplified herein.

It should be apparent to a person skill in this art that insecticidal δ -endotoxins can be identified and obtained through several means. The specific genes, or portions thereof, may be obtained from a culture depository, or con-
30 structed synthetically, for example, by use of a gene machine. Variations of these genes may be readily constructed using standard techniques for making point mu-

tations. Also, fragments of these genes can be made using commercially available exonucleases or endonucleases according to standard procedures. For example, enzymes such as *Bal31* or site-directed mutagenesis can be used to systematically cut off nucleotides from the ends of these genes. Also, genes which code for active
5 fragments may be obtained using a variety of other restriction enzymes. Proteases may be used to directly obtain active fragments of these δ -endotoxins.

Equivalent δ -endotoxins and/or genes encoding these equivalent δ -endotoxins can also be isolated from *Bacillus* strains and/or DNA libraries using the teachings provided herein. For example, antibodies to the δ -endotoxins dis-
10 closed and claimed herein can be used to identify and isolate other δ -endotoxins from a mixture of proteins. Specifically, antibodies may be raised to the portions of the δ -endotoxins which are most constant and most distinct from other *B. thuringiensis* δ -endotoxins. These antibodies can then be used to specifically identify equivalent δ -endotoxins with the characteristic insecticidal activity by
15 immunoprecipitation, enzyme linked immunoassay (ELISA), or Western blotting.

A further method for identifying the δ -endotoxins and genes of the subject invention is through the use of oligonucleotide probes. These probes are nucleotide sequences having a detectable label. As is well known in the art, if the probe molecule and nucleic acid sample hybridize by forming a strong bond between the
20 two molecules, it can be reasonably assumed that the probe and sample are essentially identical. The probe's detectable label provides a means for determining in a known manner whether hybridization has occurred. Such a probe analysis provides a rapid method for identifying formicidal δ -endotoxin genes of the subject invention.

25 The nucleotide segments which are used as probes according to the invention can be synthesized by use of DNA synthesizers using standard procedures. In the use of the nucleotide segments as probes, the particular probe is labeled with any suitable label known to those skilled in the art, including radioactive and non-radioactive labels. Typical radioactive labels include ^{32}P , ^{125}I , ^{35}S , or the like. A
30 probe labeled with a radioactive isotope can be constructed from a nucleotide sequence complementary to the DNA sample by a conventional nick translation re-

action, using a DNase and DNA polymerase. The probe and sample can then be combined in a hybridization buffer solution and held at an appropriate temperature until annealing occurs. Thereafter, the membrane is washed free of extraneous materials, leaving the sample and bound probe molecules typically detected and
5 quantified by autoradiography and/or liquid scintillation counting.

Non-radioactive labels include, for example, ligands such as biotin or thyroxine, as well as enzymes such as hydrolases or peroxidases, or the various chemiluminescers such as luciferin, or fluorescent compounds like fluorescein and its derivatives. The probe may also be labeled at both ends with different types of
10 labels for ease of separation, as, for example, by using an isotopic label at the end mentioned above and a biotin label at the other end.

Duplex formation and stability depend on substantial complementarity between the two strands of a hybrid, and, as noted above, a certain degree of mismatch can be tolerated. Therefore, the probes of the subject invention include mutations (both single and multiple), deletions, insertions of the described sequences,
15 and combinations thereof, wherein said mutations, insertions and deletions permit formation of stable hybrids with the target polynucleotide of interest. Mutations, insertions, and deletions can be produced in a given polynucleotide sequence in many ways, by methods currently known to an ordinarily skilled artisan, and perhaps by other methods which may become known in the future.
20

The potential variations in the probes listed is due, in part, to the redundancy of the genetic code. Because of the redundancy of the genetic code, *i.e.*, more than one coding nucleotide triplet (codon) can be used for most of the amino acids used to make proteins. Therefore different nucleotide sequences can code for
25 a particular amino acid. Thus, the amino acid sequences of the *B. thuringiensis* δ -endotoxins and peptides can be prepared by equivalent nucleotide sequences encoding the same amino acid sequence of the protein or peptide. Accordingly, the subject invention includes such equivalent nucleotide sequences. Also, inverse or complement sequences are an aspect of the subject invention and can be readily
30 used by a person skilled in this art. In addition it has been shown that proteins of identified structure and function may be constructed by changing the amino acid

sequence if such changes do not alter the protein secondary structure (Kaiser and Kezdy, 1984). Thus, the subject invention includes mutants of the amino acid sequence depicted herein which do not alter the protein secondary structure, or if the structure is altered, the biological activity is substantially retained. Further, the invention also includes mutants of organisms hosting all or part of a δ -endotoxin encoding a gene of the invention. Such mutants can be made by techniques well known to persons skilled in the art. For example, UV irradiation can be used to prepare mutants of host organisms. Likewise, such mutants may include asporogenous host cells which also can be prepared by procedures well known in the art.

10

4.15 RIBOZYMES

Ribozymes are enzymatic RNA molecules which cleave particular mRNA species. In certain embodiments, the inventors contemplate the selection and utilization of ribozymes capable of cleaving the RNA segments of the present invention, and their use to reduce activity of target mRNAs in particular cell types or tissues.

Six basic varieties of naturally-occurring enzymatic RNAs are known presently. Each can catalyze the hydrolysis of RNA phosphodiester bonds *in trans* (and thus can cleave other RNA molecules) under physiological conditions. In general, enzymatic nucleic acids act by first binding to a target RNA. Such binding occurs through the target binding portion of a enzymatic nucleic acid which is held in close proximity to an enzymatic portion of the molecule that acts to cleave the target RNA. Thus, the enzymatic nucleic acid first recognizes and then binds a target RNA through complementary base-pairing, and once bound to the correct site, acts enzymatically to cut the target RNA. Strategic cleavage of such a target RNA will destroy its ability to direct synthesis of an encoded protein. After an enzymatic nucleic acid has bound and cleaved its RNA target, it is released from that RNA to search for another target and can repeatedly bind and cleave new targets.

The enzymatic nature of a ribozyme is advantageous over many technologies, such as antisense technology (where a nucleic acid molecule simply binds to a nucleic acid target to block its translation) since the concentration of ribozyme

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necessary to affect a therapeutic treatment is lower than that of an antisense oligonucleotide. This advantage reflects the ability of the ribozyme to act enzymatically. Thus, a single ribozyme molecule is able to cleave many molecules of target RNA. In addition, the ribozyme is a highly specific inhibitor, with the specificity
5 of inhibition depending not only on the base pairing mechanism of binding to the target RNA, but also on the mechanism of target RNA cleavage. Single mismatches, or base-substitutions, near the site of cleavage can completely eliminate catalytic activity of a ribozyme. Similar mismatches in antisense molecules do not prevent their action (Woolf *et al.*, 1992). Thus, the specificity of action of a ribozyme is greater than that of an antisense oligonucleotide binding the same RNA
10 site.

The enzymatic nucleic acid molecule may be formed in a hammerhead, hairpin, a hepatitis δ virus, group I intron or RNaseP RNA (in association with an RNA guide sequence) or Neurospora VS RNA motif. Examples of hammerhead
15 motifs are described by Rossi *et al.* (1992); examples of hairpin motifs are described by Hampel *et al.* (Eur. Pat. EP 0360257), Hampel and Tritz (1989), Hampel *et al.* (1990) and Cech *et al.* (U. S. Patent 5,631,359; an example of the hepatitis δ virus motif is described by Perrotta and Been (1992); an example of the RNaseP motif is described by Guerrier-Takada *et al.* (1983); Neurospora VS RNA
20 ribozyme motif is described by Collins (Saville and Collins, 1990; Saville and Collins, 1991; Collins and Olive, 1993); and an example of the Group I intron is described by Cech *et al.* (U.S. Patent 4,987,071). All that is important in an enzymatic nucleic acid molecule of this invention is that it has a specific substrate binding site which is complementary to one or more of the target gene RNA re-
25 gions, and that it have nucleotide sequences within or surrounding that substrate binding site which impart an RNA cleaving activity to the molecule. Thus the ribozyme constructs need not be limited to specific motifs mentioned herein.

The invention provides a method for producing a class of enzymatic cleaving agents which exhibit a high degree of specificity for the RNA of a desired target.
30 The enzymatic nucleic acid molecule is preferably targeted to a highly conserved sequence region of a target mRNA such that specific treatment of a disease

or condition can be provided with either one or several enzymatic nucleic acids. Such enzymatic nucleic acid molecules can be delivered exogenously to specific cells as required. Alternatively, the ribozymes can be expressed from DNA or RNA vectors that are delivered to specific cells.

5 Small enzymatic nucleic acid motifs (*e.g.*, of the hammerhead or the hairpin structure) may be used for exogenous delivery. The simple structure of these molecules increases the ability of the enzymatic nucleic acid to invade targeted regions of the mRNA structure. Alternatively, catalytic RNA molecules can be expressed within cells from eukaryotic promoters (*e.g.*, Scanlon *et al.*, 1991; Kashani-Sabet *et al.*, 1992; Dropulic *et al.*, 1992; Weerasinghe *et al.*, 1991; Ojwang
10 *et al.*, 1992; Chen *et al.*, 1992; Sarver *et al.*, 1990). Those skilled in the art realize that any ribozyme can be expressed in eukaryotic cells from the appropriate DNA vector. The activity of such ribozymes can be augmented by their release from the primary transcript by a second ribozyme (Draper *et al.*, Int. Pat. Appl. Publ. No.
15 WO 93/23569, and Sullivan *et al.*, Int. Pat. Appl. Publ. No. WO 94/02595, both hereby incorporated in their totality by reference herein; Ohkawa *et al.*, 1992; Taira *et al.*, 1991; Ventura *et al.*, 1993).

Ribozymes may be added directly, or can be complexed with cationic lipids, lipid complexes, packaged within liposomes, or otherwise delivered to target
20 cells. The RNA or RNA complexes can be locally administered to relevant tissues *ex vivo*, or *in vivo* through injection, aerosol inhalation, infusion pump or stent, with or without their incorporation in biopolymers.

Ribozymes may be designed as described in Draper *et al.* (Int. Pat. Appl. Publ. No. WO 93/23569), or Sullivan *et al.*, (Int. Pat. Appl. Publ. No. WO
25 94/02595) and synthesized to be tested *in vitro* and *in vivo*, as described. Such ribozymes can also be optimized for delivery. While specific examples are provided, those in the art will recognize that equivalent RNA targets in other species can be utilized when necessary.

Hammerhead or hairpin ribozymes may be individually analyzed by computer folding (Jaeger *et al.*, 1989) to assess whether the ribozyme sequences fold
30 into the appropriate secondary structure. Those ribozymes with unfavorable intra-

molecular interactions between the binding arms and the catalytic core are eliminated from consideration. Varying binding arm lengths can be chosen to optimize activity. Generally, at least 5 bases on each arm are able to bind to, or otherwise interact with, the target RNA.

5 Ribozymes of the hammerhead or hairpin motif may be designed to anneal to various sites in the mRNA message, and can be chemically synthesized. The method of synthesis used follows the procedure for normal RNA synthesis as described in Usman *et al.* (1987) and in Scaringe *et al.* (1990) and makes use of common nucleic acid protecting and coupling groups, such as dimethoxytrityl at
10 the 5'-end, and phosphoramidites at the 3'-end. Average stepwise coupling yields are typically >98%. Hairpin ribozymes may be synthesized in two parts and annealed to reconstruct an active ribozyme (Chowrira and Burke, 1992). Ribozymes may be modified extensively to enhance stability by modification with nuclease resistant groups, for example, 2'-amino, 2'-C-allyl, 2'-fluoro, 2'-O-methyl, 2'-H (for
15 a review *see* Usman and Cedergren, 1992). Ribozymes may be purified by gel electrophoresis using general methods or by high pressure liquid chromatography and resuspended in water.

 Ribozyme activity can be optimized by altering the length of the ribozyme binding arms, or chemically synthesizing ribozymes with modifications that prevent their degradation by serum ribonucleases (*see e.g.*, Int. Pat. Appl. Publ. No.
20 WO 92/07065; Perrault *et al.*, 1990; Pieken *et al.*, 1991; Usman and Cedergren, 1992; Int. Pat. Appl. Publ. No. WO 93/15187; Int. Pat. Appl. Publ. No. WO 91/03162; Eur. Pat. Appl. Publ. No. 92110298.4; U.S. Patent 5,334,711; and Int. Pat. Appl. Publ. No. WO 94/13688, which describe various chemical modifications
25 that can be made to the sugar moieties of enzymatic RNA molecules), modifications which enhance their efficacy in cells, and removal of stem II bases to shorten RNA synthesis times and reduce chemical requirements.

 Sullivan *et al.* (Int. Pat. Appl. Publ. No. WO 94/02595) describes the general methods for delivery of enzymatic RNA molecules. Ribozymes may be administered to cells by a variety of methods known to those familiar to the art, including, but not restricted to, encapsulation in liposomes, by iontophoresis, or by
30

incorporation into other vehicles, such as hydrogels, cyclodextrins, biodegradable nanocapsules, and bioadhesive microspheres. For some indications, ribozymes may be directly delivered *ex vivo* to cells or tissues with or without the aforementioned vehicles. Alternatively, the RNA/vehicle combination may be locally delivered by direct inhalation, by direct injection or by use of a catheter, infusion pump or stent. Other routes of delivery include, but are not limited to, intravascular, intramuscular, subcutaneous or joint injection, aerosol inhalation, oral (tablet or pill form), topical, systemic, ocular, intraperitoneal and/or intrathecal delivery. More detailed descriptions of ribozyme delivery and administration are provided in Sullivan *et al.* (Int. Pat. Appl. Publ. No. WO 94/02595) and Draper *et al.* (Int. Pat. Appl. Publ. No. WO 93/23569) which have been incorporated by reference herein.

Another means of accumulating high concentrations of a ribozyme(s) within cells is to incorporate the ribozyme-encoding sequences into a DNA expression vector. Transcription of the ribozyme sequences are driven from a promoter for eukaryotic RNA polymerase I (pol I), RNA polymerase II (pol II), or RNA polymerase III (pol III). Transcripts from pol II or pol III promoters will be expressed at high levels in all cells; the levels of a given pol II promoter in a given cell type will depend on the nature of the gene regulatory sequences (enhancers, silencers, *etc.*) present nearby. Prokaryotic RNA polymerase promoters may also be used, providing that the prokaryotic RNA polymerase enzyme is expressed in the appropriate cells (Elroy-Stein and Moss, 1990; Gao and Huang, 1993; Lieber *et al.*, 1993; Zhou *et al.*, 1990). Ribozymes expressed from such promoters can function in mammalian cells (*e.g.* Kashani-Saber *et al.*, 1992; Ojwang *et al.*, 1992; Chen *et al.*, 1992; Yu *et al.*, 1993; L'Huillier *et al.*, 1992; Lisiewicz *et al.*, 1993). Such transcription units can be incorporated into a variety of vectors for introduction into mammalian cells, including but not restricted to, plasmid DNA vectors, viral DNA vectors (such as adenovirus or adeno-associated vectors), or viral RNA vectors (such as retroviral, semliki forest virus, sindbis virus vectors).

Ribozymes of this invention may be used as diagnostic tools to examine genetic drift and mutations within cell lines or cell types. They can also be used to assess levels of the target RNA molecule. The close relationship between ri-

bozyme activity and the structure of the target RNA allows the detection of mutations in any region of the molecule which alters the base-pairing and three-dimensional structure of the target RNA. By using multiple ribozymes described in this invention, one may map nucleotide changes which are important to RNA
5 structure and function *in vitro*, as well as in cells and tissues. Cleavage of target RNAs with ribozymes may be used to inhibit gene expression and define the role (essentially) of specified gene products in particular cells or cell types.

5.0 EXAMPLES

10 The following examples are included to demonstrate preferred embodiments of the invention. It should be appreciated by those of skill in the art that the techniques disclosed in the examples which follow represent techniques discovered by the inventor to function well in the practice of the invention, and thus can be considered to constitute preferred modes for its practice. However, those of skill in
15 the art should, in light of the present disclosure, appreciate that many changes can be made in the specific embodiments which are disclosed and still obtain a like or similar result without departing from the spirit and scope of the invention.

5.1 EXAMPLE 1 -- THREE-DIMENSIONAL STRUCTURE OF CRY3Bb

20 The three-dimensional structure of Cry3Bb was determined by X-ray crystallography. Crystallization of Cry3Bb and X-ray diffraction data collection were performed as described by Cody *et al.* (1992). The crystal structure of Cry3Bb was refined to a residual R factor of 18.0% using data collected to 2.4 Å resolution. The crystals belong to the space group C222₁ with unit cell dimensions $a = 122.44$,
25 $b = 131.81$, and $c = 105.37$ Å and contain one molecule in the asymmetric unit. Atomic coordinates for Cry3Bb are described in Example 31 and listed in Section 9.

The structure of Cry3Bb is similar to that of Cry3A (Li *et al.*, 1991). It consists of 5825 protein atoms from 588 residues (amino acids 64 - 652) forming
30 three discrete domains (FIG. 1). A total of 251 water molecules have been identified in the Cry3Bb structure (FIG. 2). Domain 1 (residues 64 - 294) is a seven

helical bundle formed by six helices twisted around the central helix, $\alpha 5$ (FIG. 3). The amino acids forming each helix are listed in FIG. 4. Domain 2 (residues 295 - 502) contains three antiparallel β -sheets (FIG. 5A and FIG. 5B). Sheets 1 and 2, each composed of 4 β strands, form the distinctive "Greek key" motif. The outer surface of sheet 3, composed of 3 β strands, makes contact with helix $\alpha 7$ of domain 1. FIG. 6 lists the amino acids comprising each β strand in domain 2. A small α helix, $\alpha 8$ which follows β strand 1, is also included in domain 2. Domain 3 (residues 503 - 652) has a "jelly roll" β -barrel topology which has a hydrophobic core and is nearly parallel to the a and perpendicular to the c axes of the lattice (FIG. 7A and FIG. 7B). The amino acids comprising each β strand of domain 3 are listed in FIG. 8.

The monomers of Cry3Bb in the crystal form a dimeric quaternary structure along a two-fold axis parallel to the a axis (FIG. 9A and FIG. 9B). Helix $\alpha 6$ lies in a cleft formed by the interface of domain 1 and domains 1 and 3 of its symmetry related molecule. There are numerous close hydrogen bonding contacts along this surface, confirming the structural stability of the dimer.

5.2 EXAMPLE 2 -- PREPARATION OF CRY3Bb.60

B. thuringiensis EG7231 was grown through sporulation in C2 medium with chloramphenicol (Cml) selection. The solids from this culture were recovered by centrifugation and washed with water. The toxin was purified by recrystallization from 4.0 M NaBr (Cody *et al.*, 1992). The purified Cry3Bb was solubilized in 10 ml of 50 mM KOH/100 mg Cry3Bb and buffered to pH 9.0 with 100 mM CAPS (pH 9.0). The soluble toxin was treated with trypsin at a weight ratio of 50 mg toxin to 1 mg trypsin. After 20 min of trypsin digestion the predominant protein visualized by SDS-polyacrylamide gel electrophoresis (SDS-PAGE) was 60 kDa. Further digestion of the 60-kDa toxin was not observed. FIG. 4 illustrates the Coomassie-stained Cry3Bb and Cry3Bb.60 following SDS-PAGE.

5.3 EXAMPLE 3 -- PURIFICATION AND SEQUENCING OF CRY3Bb.60

Cry3Bb.60 was electrophoretically purified by SDS-PAGE and electroblotted to Immobilon-P[®] (Millipore) membrane by semi-dry transfer at 15V for 30 min. The membrane was then washed twice with water and stained with 0.025% R-250, 40% methanol. To reduce the background, the blot was destained with 50% methanol until the stained protein bands were visible. The blot was then air dried, and the stained Cry3Bb.60 band was cut out of the membrane. This band was sent to the Tufts University Sequencing Laboratory (Boston, MA) for N-terminal sequencing. The experimentally-determined N-terminal amino acid sequence is shown in Table 7 beside the known amino acid sequence starting at amino acid residue 160.

TABLE 7

**AMINO ACID SEQUENCE OF THE N-TERMINUS OF CRY3Bb.60 AND
COMPARISON TO THE KNOWN SEQUENCE OF CRY3Bb**

Deduced Sequence	Known Sequence	Residue #
S	S	160
K	K	161
R	R	162
S	S	163
Q	Q	164
D	D	165
R	R	166

5.4 EXAMPLE 4 -- BIOACTIVITY OF CRY3Bb.60

Cry3Bb was prepared for bioassay by solubilization in a minimal amount of 50 mM KOH, 10 ml per 100 mg toxin, and buffered to pH 9.0 with 100 mM CAPS, pH 9.0. Cry3Bb.60 was prepared as described in Example 1. Both preparations were kept at room temperature 12 to 16 hours prior to bioassay. After seven days the mortality of the population was determined and analyzed to deter-

mine the lethal concentration of each toxin. These results are numerized in Table 8.

TABLE 8

5 **BIOACTIVITY OF CRY3Bb AND CRY3Bb.60 AGAINST THE SOUTHERN CORN
ROOTWORM (*DIABLOTICA UNDECIMPUNCTATA*)**

	LC ₅₀ mg/well	95% C. I.
Cry3Bb	24.09	15 - 39
Cry3Bb.60	6.72	5.25 - 8.4

5.5 Example 5 -- Ion-Channel Formation by Cry3Bb and CryB2.60

Cry3Bb.60 and Cry3Bb were evaluated for their ability to form ion chan-
10 nels in planar lipid bilayers. Bilayers of phosphatidylcholine were formed on Tef-
lon[®] supports over a 0.7-mm hole. A bathing solution of 3.5 ml 100 mM KOH, 10
mM CaCl₂, 100 mM CAPS (pH 9.5) was placed on either side of the Teflon[®] par-
tition. The toxin was added to one side of the partition and a voltage of 60 mV was
imposed across the phosphatidylcholine bilayer. Any leakage of ions through the
15 membrane was amplified and recorded. An analysis of the frequency of the con-
ductances created by either Cry3Bb or Cry3Bb.60 are illustrated in FIG. 5A and
FIG. 5B. Cry3Bb.60 readily formed ion channels whereas Cry3Bb rarely formed
channels.

20 **5.6 EXAMPLE 6 -- FORMATION OF HIGH MOLECULAR-WEIGHT OLIGOMERS**

Individual molecules of Cry3Bb or Cry3Bb.60 form a complex with an-
other like molecule. The ability of Cry3Bb to form an oligomer is not reproducibly
apparent. The complex cannot be repeatedly observed to form under nondenatur-
ing conditions. Cry3Bb.60 formed a significantly greater amount of a higher mo-
25 lecular-weight complex (≥120 kDa) with other Cry3Bb.60 molecules. Oligomers
of Cry3Bb are demonstrated by the intensity of the Coomassie-stained SDS poly-
acrylamide gel. Oligomerization is visualized on SDS-PAGE by not heating sam-
ples prior to loading on the gel to retain some nondenatured toxin. These data sug-

gest that Cry3Bb.60 more readily forms the higher order complex than Cry3Bb alone. Oligomerization is also observed by studying the conductance produced by these molecules and the time-dependent increase in conductance. This change in conductance can be attributed to oligomerization of the toxin.

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5.7 EXAMPLE 7 -- DESIGN METHOD 1: IDENTIFICATION AND ALTERATION OF PROTEASE-SENSITIVE SITES AND PROTEOLYTIC PROCESSING

It has been reported in the literature that treatment of Cry3A toxin protein with trypsin, an enzyme that cleaves proteins on the carboxyl side of available lysine and arginine residues, yields a stable cleavage product of 55 kDa from the 67 kDa native protein (Carroll *et al.*, 1989). N-terminal sequencing of the 55 kDa product showed cleavage occurs at amino acid residue R158. The truncated Cry3A protein was found to retain the same level of insecticidal activity as the native protein. Cry3Bb toxin protein was also treated with trypsin. After digestion, the protein size decreased from 68 kDa, the molecular weight of the native Cry3Bb toxin, to 60 kDa. No further digestion was observed. N-terminal sequencing revealed the trypsin cleavage site of the truncated toxin (Cry3Bb.60) to be amino acid R159 in $\alpha 3,4$ of Cry3Bb. Unexpectedly, the bioactivity of the truncated Cry3Bb toxin was found to increase.

Using this method, protease digestion of a *B. thuringiensis* toxin protein, a proteolytically sensitive site was identified on Cry3Bb, and a more highly active form of the protein (Cry3Bb.60) was identified. Modifications to this proteolytically-sensitive site by introducing an additional protease recognition site also resulted in the isolation of a biologically more active protein. It is also possible that removal of other protease-sensitive site(s) may improve activity. Proteolytically sensitive regions, once identified, may be modified or utilized to produce biologically more active toxins.

5.7.1 CRY3Bb.60

Treatment of solubilized Cry3Bb toxin protein with trypsin results in the isolation of a stable, truncated Cry3Bb toxin protein with a molecular weight of 60 kDa (Cry3Bb.60). N-terminal sequencing of Cry3Bb.60 shows the trypsin-sensitive site to be R159 in $\alpha 3,4$ of the native toxin. Trypsin digestion results in the removal of helices 1-3 from the native Cry3Bb but also increases the activity of the toxin against SCRW larvae approximately four-fold.

Cry3Bb.60 is a unique toxin with enhanced insecticidal use over the parent Cry3Bb. Improved biological activity, is only one parameter that distinguishes it as a new toxin. Aside from the reduced size, Cry3Bb.60 is also a more soluble protein. Cry3Bb precipitates from solution at pH 6.5 while Cry3Bb.60 remains in solution from pH 4.5 to pH 12. Cry3Bb.60 also forms ion channels with greater frequency than Cry3Bb.

Cry3Bb.60 is produced by either the proteolytic removal of the first 159 amino acid residues, or the *in vivo* production of this toxin, by bacteria or plants expressing the gene for Cry3Bb.60, that is, the Cry3Bb gene without the first 483 nucleotides.

In conclusion, Cry3Bb.60 is distinct from Cry3Bb in several important ways: enhanced insecticidal activity; enhanced range of solubility; enhanced ability to form channels; and reduced size.

5.7.2 EG11221

Semi-random mutagenesis of the trypsin-sensitive $\alpha 3,4$ region of Cry3Bb resulted in the isolation of Cry3Bb.11221, a designed Cry3Bb protein that exhibits over a 6-fold increase in activity against SCRW larvae compared to WT. Cry3Bb.11221 has 4 amino acid changes in the $\alpha 3,4$ region. One of these changes, L158R, introduces an additional trypsin site adjacent to R159, the proteolytically sensitive site used to produce Cry3Bb.60 (example 4.1.1). Cry3Bb.11221 is produced by *B. thuringiensis* as a full length toxin protein but is presumably digested by insect gut proteases to the same size as Cry3Bb.60 (see Cry3A results from Carroll *et al.*, 1989). The additional protease recognition site

may make the α 3,4 region even more sensitive to digestion, thereby increasing activity.

5 **5.8 EXAMPLE 8 -- DESIGN METHOD 2: DETERMINATION AND MANIPULATION
 OF BOUND WATER**

There are several ways that water molecules can associate with a protein, including surface water that is easily removed and bound water that is more difficult to extract (Dunitz, 1994; Zhang and Matthews, 1994). The function of bound water has been the subject of significant academic extrapolation, but the precise
10 function has little experimental validation. Some of the most interesting bound or structural water is the water that participates in the protein structure from inside the protein itself.

The occupation of a site by a water molecule can indicate a stable pocket within a protein or a looseness of packing created by water-mediated salt bridges
15 and hydrogen bonding to water. This can reduce the degree of bonding between amino acids, possibly making the region more flexible. A different amino acid sequence around that same site could result in better packing, collapsing the pocket around polar or charged amino acids. This may result in decreased flexibility. Therefore, the degree of hydration of a region of a protein may determine the
20 flexibility or mobility of that region, and manipulation of the hydration may alter the flexibility. Methods of increasing the hydration of a water-exposed region include increasing the number of hydrophobic residues along that surface. It is taught in the art that exposed hydrophobic residues require significantly more water to hydrate than hydrophilic residues (CRC Handbook of Chemistry and Physics,
25 CRC Press, Inc.). It is not taught, however, that by doing this, improvements to the biological activity of a protein can be achieved.

Structural water has not previously been identified in *B. thuringiensis* δ -endotoxins including Cry3Bb. Furthermore, there are no reports of the function of this structural water in δ -endotoxins or bacterial toxins. In the analysis of Cry3Bb,
30 it was observed that a collection of water molecules are located around α 3,4, a site defined by the inventors as important for improvement of bioactivity. The loop

α 3,4 region is surface exposed and may define a hinge in the protein permitting either removal or movement of the first three helices of domain 1. The hydration found around this region may impart flexibility and mobility to this loop. The observation of structural water at the α 3,4 site provided an analytical tool for further structure analysis. If this important site is surrounded by water, then other important sites may also be completely or partially surrounded by water. Using this insight, structural water surrounding helices 5 and 6 was then identified. This structural water forms a column through the protein, effectively separating helices 5 and 6 from the rest of the molecule. The structures of Cry3A and Cry3Bb suggest that helices 5 and 6 are tightly associated, bound together by Van der Waals interactions. Alone, helix 5 from Cry3A, although insufficient for biological activity, has been demonstrated to have the ability to form ion channels in an artificial membrane (Gazit and Shai, 1993). The ion channels formed by helix 5 are 10-fold smaller than the channels of the full length toxin suggesting that significantly more toxin structure is required for the full-sized ion channels. In Cry3Bb, helix 5 as part of a cluster of α helices (domain 1) has been found to form ion channels (Von Tersch *et al.*, 1994). Unpublished experimental observations by the inventors demonstrate that helix 6 also crossed the biological membrane. Helices 5 and 6, therefore, are the putative channel-forming helices necessary for toxicity.

The hydration around these helices may indicate that flexibility of this region is necessary for toxicity. It is conceivable, therefore, that if it were possible to improve the hydration around helices 5 and 6, one could create a better toxin protein. Care must be taken, however, to avoid creating continuous hydrophobic surfaces between helices 5-6 and any other part of the protein which could, by hydrophobic interactions, act to restrict movement of the mobile helices. The mobility of helices 5 and 6 may also depend on the flexibility of the loops attached to them as well as on other regions of the Cry3Bb molecule, particularly in domain 1, which may undergo conformational changes to allow insertion of the 2 helices into the membrane. Altering the hydration of these regions of the protein may also affect its bioactivity.

5.8.1 CRY3BB.11032

A collection of bound water residues indicated the relative flexibility of the α 3,4 region. The flexibility of this loop can be increased by increasing the hydration of the region by substituting relatively hydrophobic residues for the exposed hydrophilic residues. An example of an improved, designed protein having this type of substitution is Cry3Bb.11032. Cry3Bb.11032 has the amino acid change D165G; glycine is more hydrophobic than aspartate (Kyte and Doolittle hydrophobicity score of -0.4 vs. -3.5 for aspartate). Cry3Bb.11032 is approximately 3 times more active than WT Cry3Bb.

5.8.2 CRY3BB.11051

To increase the hydration of the α 4,5 region of Cry3Bb, glycine was substituted for the surface exposed residue K189. Glycine is more hydrophobic than lysine (Kyte and Doolittle hydrophobicity score of -0.4 vs. -3.9 for lysine) and may result in an increase in bound water. The increase in bound water may impart greater flexibility to the loop region which precedes the channel-forming helix, α 5. The designed Cry3Bb protein with the K189G change, Cry3Bb.11051, exhibits a 3-fold increase in activity compared to WT Cry3Bb.

5.8.3 ALTERATIONS TO α 7, β 1 (CRY3BB.11241 AND 11242)

Amino acid changes made in the surface-exposed loop connecting α -helix 7 and β -strand 1 (α 7, β 1) resulted in the identification of 2 altered Cry3Bb proteins with increased bioactivities, Cry3Bb.11241 and Cry3Bb.11242. Analysis of the hydropathy index of 2 of these proteins over the 20 amino acid sequence 281-300, inclusive of the α 7, β 1 region, reveal that the amino acid substitutions in these proteins have made the α 7, β 1 region much more hydrophobic. The grand average of hydropathy value (GRAVY) was determined for each protein sequence using the PC\GENE[®] (IntelliGenetics, Inc., Mountain View, CA, release 6.85) protein sequence analysis computer program, SOAP, and a 7 amino acid interval. The SOAP program is based on the method of Kyte and Doolittle (1982). The increase

in hydrophobicity of the $\alpha 7, \beta 1$ region for each protein may increase the hydration of the loop and, therefore, the flexibility. The altered proteins, their respective amino acid changes, fold-increases over WT bioactivity, and GRAVY values are listed in Table 9.

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TABLE 9
HYDROPATHY VALUES FOR THE $\alpha 7, \beta 1$ REGION OF CRY3Bb AND 2 DESIGNED CRY3Bb PROTEINS SHOWING INCREASED SCRw BIOACTIVITY

Cry3Bb*	Amino Acid Changes	Fold Increase in	GRAVY
Protein		Bioactivity	(Amino Acids
		Over WT	281-300)
wildtype	—	—	4.50
Cry3Bb.11241	Y287F, D288N, R290L	2.6×	10.70
Cry3Bb.11242	R290V	2.5×	8.85

10 **5.8.4 ALTERATIONS TO $\beta 1, \alpha 8$ (CRY3Bb.11228, CRY3Bb.11229, CRY3Bb.11230, CRY3Bb.11233, CRY3Bb.11236, CRY3Bb.11237, CRY3Bb.11238 AND CRY3Bb.11239)**

The surface-exposed loop between β -strand 1 and α -helix 8 ($\beta 1, \alpha 8$) defines the boundary between domains 1 and 2 of Cry3Bb. The introduction of semi-random amino acid changes to this region resulted in the identification of several altered Cry3Bb proteins with increased bioactivity. Hydropathy index analysis of the amino acid substitutions found in the altered proteins shows that the changes have made the exposed region more hydrophobic which may result in increased hydration and flexibility. Table 10 lists the altered proteins, their respective amino acid changes and fold increases over WT Cry3Bb and the grand average of hydropathy value (GRAVY) determined using the PC\GENE[®] (IntelliGenetics, Inc., Mountain View, CA, release 6.85) protein sequence analysis program, SOAP, over the 20 amino acid sequence 305 - 324 inclusive of $\beta 1, \alpha 8$, using a 7 amino acid interval.

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TABLE 10
HYDROPATHY VALUES FOR THE L β 1, α 8 REGION OF CRY3Bb AND 8 DESIGNED
CRY3Bb* PROTEINS SHOWING INCREASED SCRWB BIOACTIVITY

Cry3Bb* Protein	Amino Acid Changes	Fold Increase in Bioactivity Over Wild Type	GRAVY (Amino Acids 305-324)
wildtype	—	—	0.85
Cry3Bb.11228	S311L, N313T, E317K	4.1×	4.35
Cry3Bb.11229	S311T, E317K, Y318C	2.5×	2.60
Cry3Bb.11230	S311A, L312V, Q316W	4.7×	3.65
Cry3Bb.11233	S311A, Q316D	2.2×	2.15
Cry3Bb.11236	S311I	3.1×	3.50
Cry3Bb.11237	S311I, N313H	5.4×	3.65
Cry3Bb.11238	N313V, T314N, Q316M, E317V	2.6×	9.85
Cry3Bb.11239	N313R, L315P, Q316L, E317A	2.8×	3.95

5 5.8.5 CRY3Bb.11227, CRY3Bb.11241 AND CRY3Bb.11242

Amino acid Q238, located in helix 6 of Cry3Bb, has been identified as a residue that, by its large size and hydrogen bonding to R290, blocks complete hydration of the space between helix 6 and helix 4. Substitution of R290 with amino acids that do not form hydrogen bonds or that have side chains that can not span the physical distance to hydrogen bond with Q238 may result in increased hydration around Q238. Q238, unable to hydrogen bond to R290, may now bind water. This may increase the flexibility of the channel-forming region. Designed proteins Cry3Bb.11227 (R290N), Cry3Bb.11241 (R290L) and Cry3Bb.11242 (R290V)

show increased activities of approximately 2-fold, 2.6-fold and 2.5-fold, respectively, against SCRW larvae compared to WT.

5.9 EXAMPLE 9 -- DESIGN METHOD 3: MANIPULATION OF HYDROGEN BONDS AROUND MOBILE REGIONS

Mobility of regions of a protein may be required for activity. The mobility of the α 5,6 region, the putative channel-forming region of Cry3Bb, may be improved by decreasing the number of hydrogen bonds, including salt bridges (hydrogen bonds between oppositely charged amino acid side chains), between helices 5-6 and any other part of the molecule or dimer structure. These hydrogen bonds may impede the movement of the two helices. Decreasing the number of hydrogen bonds and salt bridges may improve biological activity. Replacement of hydrogen-bonding amino acids with hydrophobic residues must be done with caution to avoid creating continuous hydrophobic surfaces between helices 5-6 and any other part of the dimer. This may decrease mobility by increasing hydrophobic surface interactions.

5.9.1 CRY3BB.11222 AND CRY3BB.11223

Tyr230 is located on helix 6 and, in the quaternary dimer structure of Cry3Bb, this amino acid is coordinated with Tyr230 from the adjacent molecule. Three hydrogen bonds are formed between the two helices 6 in the two monomers because of this single amino acid. In order to improve the flexibility of helices 5-6, the helices theoretically capable of penetrating the membrane and forming an ion channel, the hydrogen bonds across the dimer were removed by changing this amino acid and a corresponding increase in biological activity was observed. The designed Cry3Bb proteins, Cry3Bb.11222 and Cry3Bb.EG11223, show a 4-fold and 2.8-fold increase in SCRW activity, respectively, compared to WT.

5.9.2 CRY3BB.11051

Designed Cry3Bb protein Cry3Bb.11051 has amino acid change K189G in α 4,5 of domain 1. In the WT Cry3Bb structure, the exposed side chain of K189 is

close enough to the exposed side chain of E123, located in $\alpha 2b,3$, to form hydrogen bonds. Substitution of K189 with glycine, as found in this position in Cry3A, removes the possibility of hydrogen bond formation at this site and results in a protein with a bioactivity three-fold greater than WT Cry3Bb.

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5.9.3 CRY3Bb.11227, CRY3Bb.11241 AND CRY3Bb.11242

Amino acid Q238, located in helix 6 of Cry3Bb, has been identified as a residue that, by its large size and hydrogen bonding to R290, blocks complete hydration of the space between helix 6 and helix 4. Substitution of R290 with amino acids that do not form hydrogen bonds or that have side chains that can not span the physical distance to hydrogen bond with Q238 may increase the flexibility of the channel-forming region. Designed proteins Cry3Bb.11227 (R290N), Cry3Bb.11241 (R290L) and Cry3Bb.11242 (R290V) show increased activities of approximately 2-fold, 2.6-fold and 2.5-fold, respectively, against SCRW larvae compared to WT

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5.10 EXAMPLE 10 -- DESIGN METHOD 4: LOOP ANALYSIS AND LOOP DESIGN AROUND FLEXIBLE HELICES

Loop regions of a protein structure may be involved in numerous functions of the protein including, but not limited to, channel formation, quaternary structure formation and maintenance, and receptor binding. Cry3Bb is a channel-forming protein. The availability of the ion channel-forming helices of δ -endotoxins to move into the bilayer depend upon the absence of forces that hinder the process. One of the forces possibly limiting this process is the steric hindrance of amino acid side chains in loop regions around the critical helices. The literature suggests that in at least one other bacterial toxin, not a *B. thuringiensis* toxin, the toxin molecule opens up or, in scientific terms, loses some of the quaternary structure to expose a membrane-active region (Cramer *et al.*, 1990). This literature does not teach how to improve the probability of this event occurring and it is not known if *B. thuringiensis* toxins use this same process to penetrate the membrane. Reducing

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the steric hindrance of the amino acid side chains in these critical regions by reducing size or altering side chain positioning with the corresponding increase in biological activity was the inventive step.

5 5.10.1 ANALYSIS OF THE LOOP BETWEEN HELICES 3 AND 4 (CRY3Bb.11032)

The inventors have discovered that the first three helices of domain one could be cleaved from the rest of the toxin by proteolytic digestion of the loop between helices $\alpha 3$ and $\alpha 4$ (Cry3Bb.60). Initial efforts to truncate the *cry3Bb* gene to produce this shortened, though more active Cry3Bb molecule, failed. For unknown reasons, *B. thuringiensis* failed to synthesize this 60-kDa molecule. It was then reasoned that perhaps the first three helices of domain 1 did not have to be proteolytically removed, or equivalently, the protein did not have to be synthesized in this truncated form to take advantage of the Cry3Bb.60 design. It was observed that the protein Cry3A had a small amino acid near the $\alpha 3,4$ that might impart greater flexibility in the loop region thereby permitting the first three helices of domain 1 to move out of the way, exposing the membrane-active region. By designing a Cry3Bb molecule with a glycine residue near this loop, the steric hindrance of residues in the loop might be lessened. The redesigned protein, Cry3Bb.11032, has the amino acid change D165G, which replaces the larger aspartate residue (average mass of 115.09) with the smallest amino acid, glycine (average mass of 57.05). The activity of Cry3Bb.11032 is approximately 3-fold greater than that of the WT protein. In this way, the loop between helices $\alpha 3$ and $\alpha 4$ was rationally redesigned with a corresponding increase in the biological activity.

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5.10.2 CRY3Bb.11051

The loop region connecting helices $\alpha 4$ and $\alpha 5$ in Cry3Bb must be flexible so that the channel-forming helices $\alpha 5$ - $\alpha 6$ can penetrate into the membrane. It was noticed that Cry3A has a glycine residue in the middle of this loop that may impart greater flexibility. The corresponding change, K189G, was made in Cry3Bb and

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the resulting, designed protein, Cry3Bb.11051, exhibits a 3-fold increase in activity against SCRW larvae compare to WT Cry3Bb.

5 **5.10.3 ANALYSIS OF THE LOOP BETWEEN β -STRAND 1 AND HELIX 8**
(CRY3Bb.11228, CRY3Bb.11229, CRY3Bb.11230, CRY3Bb.11232,
CRY3Bb.11233, CRY3Bb.11236, CRY3Bb.11237, CRY3Bb.11238, AND
CRY3Bb.11239)

10 The loop region located between β strand 1 of domain 2 and α helix 8 in domain 2 is very close to the loop between α helices 6 and 7 in domain 1. Some of the amino acids side chains of $l\beta 1, \alpha 8$ appear as though they may sterically impede movement of $l\alpha 6, 7$. Since $l\alpha 6, 7$ must be flexible for the channel-forming helices $\alpha 5$ - $\alpha 6$ to insert into the membrane, it was thought that re-engineering this loop may change the positioning of the side chains resulting in less steric hindrance. This was accomplished creating proteins with increased biological activities ranging from 2.2 to 5.4 times greater than WT. These designed toxin proteins and their amino acid changes are listed in Table 2 as Cry3Bb.11228, Cry3Bb.11229, Cry3Bb.11230, Cry3Bb.11232, Cry3Bb.11233, Cry3Bb.11236, Cry3Bb.11237, Cry3Bb.11238, and Cry3Bb.11239.

20 **5.10.4 ANALYSIS OF THE LOOP BETWEEN HELIX 7 AND β -STRAND 1**
(CRY3Bb.11227, CRY3Bb.11234, CRY3Bb.11241, CRY3Bb.11242, AND
CRY3Bb.11036)

25 If Cry3Bb is similar to a bacterial toxin which must open up to expose a membrane active region for toxicity, it is possible that other helices in addition to the channel-forming helices must also change positions. It was reasoned that, if helices $\alpha 5$ - $\alpha 6$ insert into the membrane, than helix $\alpha 7$ may have to change positions also. It was shown in example 4.4.3 that increasing flexibility between helix $\alpha 6$ and $\alpha 7$ can increase activity, greater flexibility in the loop following helix $\alpha 7$, $l\alpha 7, \beta 1$ may also increase bioactivity. Alterations to the $l\alpha 7, \beta 1$ region of Cry3Bb resulted in the isolation of several proteins with increased activities ranging from

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1.9 to 4.3 times greater than WT. These designed proteins are listed in Table 7 as Cry3Bb.11227, Cry3Bb.11234, Cry3Bb.11241, Cry3Bb.11242, and Cry3Bb.11036.

5 **5.11 EXAMPLE 11 -- DESIGN METHOD 5: LOOP DESIGN AROUND β STRANDS AND β SHEETS**

Loop regions of a protein structure may be involved in numerous functions of the protein including, but not limited to, channel formation, quaternary structure formation and maintenance, and receptor binding. A binding surface is often defined by a number of loops, as is the case with immunoglobulin G (IgG) (see Branden and Tooze, 1991, for review). What can not be determined at this point, however, is what loops will be important for receptor interactions just by looking at the structure of the protein in question. Since a receptor has not been identified for Cry3Bb, it is not even possible to compare the structure of Cry3Bb with other proteins that have the same receptor for structural similarities. To identify Cry3Bb loops that contribute to receptor interactions, random mutagenesis was performed on surface-exposed loops.

As each loop was altered, the profile of the overall bioactivities of the resultant proteins were examined and compared. The loops, especially in domain 2 which appears to be unnecessary for channel activity, fall into two categories: (1) loops that could be altered without much change in the level of bioactivity of the resultant proteins and (2) loops where alterations resulted in overall loss of resultant protein bioactivity. Using this design method, it is possible to identify several loops important for activity.

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5.11.1 ANALYSIS OF LOOP β 2,3

Semi-random mutagenesis of the loop region between β strands 2 and 3 resulted in the production of structurally stable toxin proteins with significantly reduced activities against SCRW larvae. The l β 2,3 region is highly sensitive to amino acid changes indicating that specific amino acids or amino acid sequences

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are necessary for toxin protein activity. It is conceivable, therefore, that specific changes in the l β 2,3 region will increase the binding and, therefore, the activity of the redesigned toxin protein.

5 5.11.2 ANALYSIS OF LOOP β 6,7

Semi-random mutations introduced to the loop region between β strands 6 and 7 resulted in structurally stable proteins with an overall loss of SCRW bioactivity. The l β 6,7 region is highly sensitive to amino acid changes indicating that specific amino acids or amino acid sequences are necessary for toxin protein activity. It is conceivable, therefore, that specific changes in the l β 6,7 region will increase the binding and, therefore, the activity of the redesigned toxin protein.

5.11.3 ANALYSIS OF LOOP β 10,11

Random mutations to the loop region between β strands 10 and 11 resulted in proteins having an overall loss of SCRW bioactivity. Loop β 10,11 is structurally close to and interacts with loops β 2,3 and β 6,7. Specific changes to individual residues within the l β 10,11 region may also result in increased interaction with the insect membrane, increasing the bioactivity of the toxin protein.

20 5.11.4 CRY3BB.11095

Loops β 2,3, β 6,7 and β 10,11 have been identified as important for bioactivity of Cry3Bb. The 3 loops are surface-exposed and structurally close together. Amino acid Q348 in the WT structure, located in β -strand 2 just prior to l β 2,3, does not form any intramolecular contacts. However, replacing Q348 with arginine (Q348R) results in the formation of 2 new hydrogen-bonds between R348 and the backbone carbonyls of R487 and R488, both located in l β 10,11. The new hydrogen bonds may act to stabilize the structure formed by the 3 loops. The designed protein carrying this change, Cry3Bb.11095, is 4.6-fold more active than WT Cry3Bb.

5.12 EXAMPLE 12 -- DESIGN METHOD 6: IDENTIFICATION AND RE-DESIGN OF COMPLEX ELECTROSTATIC SURFACES

Interactions of proteins include hydrophobic interactions (*e.g.*, Van der Waals forces), hydrophilic interactions, including those between opposing charges on amino acid side chains (salt bridges), and hydrogen bonding. Very little is known about δ -endotoxin and receptor interactions. Currently, there are no literature reports identifying the types of interactions that predominate between *B. thuringiensis* toxins and receptors.

Experimentally, however, it is important to increase the strength of the *B. thuringiensis* toxin-receptor interaction and not permit the precise determination of the chemical interaction to stand in the way of improving it. To accomplish this, the electrostatic surface of Cry3Bb was defined by solving the Poisson-Boltzman distribution around the molecule. Once this electrically defined surface was solved, it could then be inspected for regions of greatest diversity. It was reasoned that these electrostatically diverse regions would have the greatest probability of participating in the specific interactions between the *B. thuringiensis* toxin proteins and the receptor, rather than more general and non-specific interactions. Therefore, these regions were chosen for redesign, continuing to increase the electrostatic diversity of the regions. In addition, examination of the electrostatic interaction around the putative channel forming region of the toxin created insights for redesign. This includes identification of an electropositive residue in an otherwise negatively charged conduit (see example 4.6.1).

5.12.1 R290 (Cry3Bb.11227, Cry3Bb.11241, and Cry3Bb.11242)

Examination of the Cry3Bb dimer interface along the domain 1 axis suggested that a pore or conduit for cations might be formed between the monomers. Electrostatic examination of this axis lent additional credibility to this suggestion. In fact, the hypothetical conduit is primarily negatively charged, an observation consistent with the biophysical analysis of cation-selective, δ -endotoxin channels. If a cation channel were formed along the axis of the dimer, then the cation could

move between the monomers relatively easily with only one significant hurdle. A positively charged arginine residue (R290) lies in the otherwise negatively charged conduit. This residue could impede the cation movement through the channel. Based on this analysis, R290 was changed to uncharged residues. The bioactivity
5 of redesigned proteins Cry3Bb.11227 (R290N), Cry3Bb.11241 (R290L) and Cry3Bb.11242 (R290V) was improved approximately 2-fold, 2.6-fold and 2.5-fold, respectively.

5.12.2 CRY3Bb.60

10 Trypsin digestion of solubilized Cry3Bb yields a stable, truncated protein with a molecular weight of 60 kDa (Cry3Bb.60). Trypsin digestion occurs on the carboxyl side of residue R159, effectively removing helices 1 through 3 from the native Cry3Bb structure. The cleavage of the first 3 helices exposes an electrostatic surface different than those found in the native structure. The new surface
15 has a combination of hydrophobic, polar and charged characteristics that may play a role in membrane interactions. The bioactivity of Cry3Bb.60 is 3.6-fold greater than that of WT Cry3Bb.

20 5.13 EXAMPLE 13 -- DESIGN METHOD 7: IDENTIFICATION AND REMOVAL OF METAL BINDING SITES

The literature teaches that the *in vitro* behavior of *B. thuringiensis* toxins can be increased by chelating divalent cations from the experimental system (Crawford and Harvey 1988). It was not known, however, how these divalent cations inhibited the *in vitro* activity. Crawford and Harvey (1988) demonstrated
25 that the short circuit current across the midgut was more severely inhibited by *B. thuringiensis* in the presence of EDTA, a chelator of divalent ions, than in the absence of this agent, thus suggesting that this step in the mode of action of *B. thuringiensis* could be potentiated by removing divalent ions. Similar observations were made using black-lipid membranes and measuring an increase in the
30 current created by the δ - endotoxins in the presence of EDTA to chelate divalent

ions. There were at least three possible explanations for these observations. The first explanation could be that the divalent ions are too large to move through a ion channel more suitable for monovalent ions, thereby blocking the channel. Second, the divalent ions may cover the protein in the very general way, thereby buffering the charge interactions required for toxin membrane interaction and limiting ion channel activity. The third possibility is that a specific metal binding site exists on the protein and, when occupied by divalent ions, the performance of the ion channel is impaired. Although the literature could not differentiate the value of one possibility over another, the third possibility led to an analysis of the Cry3Bb structure searching for a specific metal binding site that might alter the probability that a toxin could form an ion channel.

5.13.1 H231 (CRY3Bb.11222, CRY3Bb.11224, CRY3Bb.11225, AND CRY3Bb.11226)

A putative metal binding site is formed in the Cry3Bb dimer structure by the H231 residues of each monomer. The H231 residues, located in helix α_6 , lie adjacent to each other and close to the axis of symmetry of the dimer. Removal of this site by replacement of histidine with other amino acids was evaluated by the absence of EDTA-dependent ion channel activity. The bioactivities of the designed toxin proteins, Cry3Bb.11222, Cry3Bb.11224, Cry3Bb.11225 and Cry3Bb.11226, are increased 4-, 5-, 3.6- and 3-fold, respectively, over that of WT Cry3Bb. Their respective amino acid changes are listed in Table 2.

5.14 EXAMPLE 14 -- DESIGN METHOD 8: ALTERATION OF QUATERNARY STRUCTURE

Cry3Bb can exist in solution as a dimer similar to a related protein, Cry3A (Walters *et al.*, 1992). However, the importance of the dimer to biological activity is not known because the toxin as a monomer or as a higher order structure has not been seriously evaluated. It is assumed that specific amino acid residues contribute to the formation and stability of the quaternary structure. Once a contributing residue is identified, alterations can be made to diminish or enhance the effect of that

residue thereby affecting the interaction between monomers. Channel activity is a useful way, but by no means the only way, to assess quaternary structure of Cry3Bb and its derivatives. It has been observed that Cry3Bb creates gated conductances in membranes that grow in size with time, ultimately resulting in large pores in the membrane (the channel activity of WT Cry3Bb is described in Section 12.1). It also has been observed that Cry3A forms a more stable dimer than Cry3Bb and coincidentally forms higher level conductances faster (FIG. 10). This observation led the inventors to propose that oligomerization and ion channel formation (conductance size and speed of channel formation) were related. Based on this observation Cry3Bb was re-engineered to make larger and more stable oligomers at a faster rate. It is assumed in this analysis that the rate of ion channel formation and growth mirrors this process. It is also possible that changes in quaternary structure may not affect channel activity alone or at all. Alterations to quaternary structure may also affect receptor interactions, protein processing in the insect gut environment, as well as other aspects of bioactivity unknown.

5.14.1 CRY3BB.11048

Comparative structural analysis of Cry3A and Cry3Bb led to the identification of structural differences between the two toxins in the ion channel-forming domain; specifically, an insertion of one amino acid between helix 2a and helix 2b in Cry3Bb. Removal of this additional amino acid in Cry 3B2, A104, and a D103E substitution, as in Cry3A, resulted in loss of channel gating and the formation of symmetrical pores. Once the pores are formed they remain open and allow a steady conductance ranging from 25-130 pS. This designed protein, Cry3Bb.11048, is 4.3 times more active than WT Cry3Bb against SCRW larvae.

5.14.2 OLIGOMERIZATION OF CRY3BB.60

Individual molecules of Cry3Bb or Cry3Bb.60 can form a complex with another like molecule. Oligomerization of Cry3Bb is demonstrated by SDS-PAGE, where samples are not heated in sample buffer prior to loading on the gel. The lack of heat treatment allows some nondenatured toxin to remain. Oligomerization is

visualized following Coomassie staining by the appearance of a band at 2 times the molecular weight of the monomer. The intensity of the higher molecular weight band reflects the degree of oligomerization. The ability of Cry3Bb to form an oligomer is not reproducibly apparent. The complex cannot be repeatedly observed to form. Cry3Bb.60, however, forms a significantly greater amount of a higher molecular weight complex (120 kDa). These data suggest that Cry3Bb.60 more readily forms the higher order complex than Cry3Bb alone. Cry3Bb.60 also forms ion channels with greater frequency than WT Cry3Bb (see Section 5.12.9).

10 5.14.3 CRY3BB.11035

Changes were made in Cry3Bb to reflect the amino acid sequence in Cry3A at the end of $\alpha 3,4$ and in the beginning of helix 4. These changes resulted in the designed protein, Cry3Bb.11035, that, unlike wild type Cry3Bb, forms spontaneous channels with large conductances. Cry3Bb.11035 is also approximately three times more active against SCRW larvae than WT Cry3Bb. Cry3Bb.11035 and its amino acid changes are listed in Table 10.

5.14.4 CRY3BB.11032

Cry3Bb.11032 was altered at residue 165 in helix $\alpha 4$, changing an aspartate to glycine, as found in Cry3A. Cry3Bb.11032 is three-fold more active than WT Cry3Bb. The channel activity of Cry3Bb.11032 is much like Cry3Bb except when the designed protein is artificially incorporated into the membrane. A 16-fold increase in the initial channel conductances is observed compared to WT Cry3Bb (see Section 5.12.2). This increase in initial conductance presumably is due to enhanced quaternary structure, stability or higher-order structure.

5.14.5 EG11224

In the WT Cry3Bb dimer structure, histidine, at position 231 in domain 1, makes hydrogen bond contacts with D288 (domain 1), Y230 (domain 1), and, through a network of water molecules, also makes contacts to D610 (domain 3), all

of the opposite monomer. D610 and K235 (domain 1) also make contact. Replacing the histidine with an arginine, H231R, results, in one orientation, in the formation of a salt bridge to D610 of the neighboring monomer. In a second orientation, the contacts with D288 of the neighboring monomer, as appear in the WT structure, are retained. In either orientation, R231 does not hydrogen bond to Y230 of the opposite monomer but does make contact with K235 which retains its contacts to K610 (V. Cody, research communication). The shifting hydrogen bonds have changed the interactions between the different domains of the protein in the quaternary structure. Overall, fewer hydrogen bonds exist between domains 1 of the neighboring monomers and a much stronger bond has been formed between domains 1 and 3. Channel activity was found to be altered. Cry3Bb.11224 produces small, quickly gating channels like Cry3Bb. However, unlike WT Cry3Bb, Cry3Bb.11224 does not exhibit β -mercaptoethanol-dependent activation. Replacing H231 with arginine resulted in a designed Cry3Bb protein, Cry3Bb.11224, exhibiting a 5-fold increase in bioactivity.

5.14.6 CRY3BB.11226

Cry3Bb.11226 is similar to Cry3Bb.11224, discussed in Section 4.8.5, in that the histidine at position 231 has been replaced. The amino acid change, H231T, results in the loss of β -mercaptoethanol dependent activation seen with WT Cry3Bb (see Section 5.12.1). The replacement of H231, a putative metal binding site, changes the interaction of regions in the quaternary structure resulting in a different type of channel activity. Cry3Bb.11226 is three-fold more active than WT Cry3Bb.

5.14.7 CRY3BB.11221

Cry3Bb.11221 has been re-designed in the α 3,4 region of Cry3Bb. The channels formed by Cry3Bb.11221 are much more well resolved than the conductances formed by WT Cry3Bb (see Section 5.12.6). Cry3Bb.11221 exhibits a 6.4-fold increase in bioactivity over that of WT Cry3Bb. The amino acid changes found in Cry3Bb.11221 are listed in Table 2.

5.14.8 CRY3BB.11242

The designed protein, Cry3Bb.11242, carrying the alteration R290V, forms small conductances immediately which grow rapidly and steadily to large conductances in about 3 min (see Section 5.12.7). This is contrast to WT Cry3Bb channels which take 30-45 min to appear and grow slowly over hours to large conductances. Cry3Bb.11242 also exhibits a 2.5-fold increase in bioactivity compared to WT Cry3Bb.

10 5.14.9 CRY3BB.11230

Cry3Bb.11230, unlike WT Cry3Bb, forms well resolved channels with long open states. These channels reach a maximum conductance of 3000 pS but do not continue to grow with time. Cry3Bb.11230 has been re-designed in the I β 1, α 8 region of Cry3Bb and exhibits almost a 5-fold increase in activity against SCRW larvae (Table 9) and a 5.4-fold increase against WCRW larvae (Table 10) compared to WT Cry3Bb. The amino acid changes found in Cry3Bb.11230 are listed in Table 2.

5.15 EXAMPLE 15 -- DESIGN METHOD 9: DESIGN OF STRUCTURAL RESIDUES

20 The specific three-dimensional structure of a protein is held in place by amino acids that may be buried or otherwise removed from the surface of the protein. These structural determinants can be identified by inspection of forces responsible for the surface structure positioning. The impact of these structural residues can then be enhanced to restrict molecular motion or diminished to enhance
25 molecular flexibility.

5.15.1 CRY3BB.11095

Loops β 2,3, β 6,7 and β 10,11, located in domain 2 of Cry3Bb, have been identified as important for bioactivity. The three loops are surface-exposed and
30 structurally close together. Amino acid Q348 in the WT structure, located in β -

strand 2 just prior to l β 2,3, does not form any intramolecular contacts. However, replacing Q348 with arginine (Q348R) results in the formation of 2 new hydrogen-bonds between R348 and the backbone carbonyls of R487 and R488, both located in l β 10,11. The new hydrogen bonds may act to stabilize the structure formed by the three loops. Certainly, the structure around R348 is more tightly packed as determined by X-ray crystallography. The designed protein carrying this change, Cry3Bb.11095, is 4.6-fold more active than WT Cry3Bb.

5.16 EXAMPLE 16 -- DESIGN METHOD 10: COMBINATORIAL ANALYSIS AND MUTAGENESIS

Individual sites in the engineered Cry3Bb molecule can be used together to create a Cry3Bb molecule with activity even greater than the activity of any one site. This method has not been precisely applied to any δ -endotoxin. It is also not obvious that improvements in two sites can be pulled together to improve the biological activity of the protein. In fact, data demonstrates that improvements to 2 sites, when pulled together into a single construct, do not necessarily further improve the biological activity of Cry3Bb. In some cases, the combination resulted in decreased protein stability and/or activity. Examples of proteins with site combinations that resulted in improved activity compared to WT Cry3Bb but decreased activity compared to 1 or more of the "parental" proteins are Cry3Bb.11235, 11046, 11057 and 11058. Cry3Bb.11082, which contains designed regions from 4 parental proteins, retains the level of activity from the most active parental strain (Cry3Bb.11230) but does not show an increase in activity. These proteins are listed in Table 7. The following are examples of instances where combined mutations have significantly improved biological activity.

5.16.1 CRY3BB.11231

Designed protein Cry3Bb.11231 contains the alterations found in Cry3Bb.11224 (H231R) and Cry3Bb.11228 (changes in l β 1, α 8). The combination of amino acid changes found in Cry3Bb.11231 results in an increase in bioactivity

against SCRW larvae of approximately 8-fold over that of WT Cry3Bb (Table 2). This increase is greater than exhibited by either Cry3Bb.11224 (5.0×) or Cry3Bb.11228 (4.1×) alone. Cry3Bb.11231 also exhibits a 12.9-fold increase in activity compared to WT Cry3Bb against WCRW larvae (Table 10).

5

5.16.2 CRY3BB.11081

Designed Cry3Bb protein Cry3Bb.11081 was constructed by combining the changes found in Cry3Bb.11032 and Cry3Bb.11229 (with the exception of Y318C). Cry3Bb.11081 a 6.1-fold increase in activity over WT Cry3Bb; a greater increase in activity than either of the individual parental proteins, Cry3Bb.11032 (3.1-fold) and Cry3Bb.11229 (2.5-fold).

10

5.16.3 CRY3BB.11083

Designed Cry3Bb protein Cry3Bb.11083 was constructed by combining the changes found in Cry3Bb.11036 and Cry3Bb.11095. Cry3Bb.11083 exhibits a 7.4-fold increase in activity against SCRW larvae compared to WT Cry3Bb; a greater increase than either Cry3Bb.11036 (4.3×) or Cry3Bb.11095 (4.6×). Cry3Bb.11083 also exhibits a 5.4-fold increase in activity against WCRW larvae compared to WT Cry3Bb (Table 10).

15

20

5.16.4 CRY3BB.11084

Designed Cry3Bb protein Cry3Bb.11084 was constructed by combining the changes found in Cry3Bb.11032 and the S311L change found in Cry3Bb.11228. Cry3Bb.11084 exhibits a 7.2-fold increase in activity over that of WT Cry3Bb; a greater than either Cry3Bb.11032 (3.1×) or Cry3Bb.11228 (4.1×).

25

5.16.5 CRY3BB.11098

Designed Cry3Bb protein Cry3Bb.11098 was constructed to contain the following amino acid changes: D165G, H231R, S311L, N313T, and E317K. The

nucleic acid sequence is given in SEQ ID NO:107, and the encoded amino acid sequence is given in SEQ ID NO:108.

5.17 EXAMPLE 17 -- DESIGN STRATEGY 11: ALTERATION OF BINDING TO
5 GLYCOPROTEINS AND TO WCRW BRUSH BORDER MEMBRANES

While the identity of receptor(s) for Cry3Bb is unknown, it is nonetheless important to increase the interaction of the toxin with its receptor. One way to improve the toxin-receptor interaction with knowing the identity of the receptor is to reduce or eliminate non-productive binding to other biomolecules. The inventors
10 have observed that Cry3Bb binds non-specifically to bovine serum albumin (BSA) that has been glycosylated with a variety of sugar groups, but not to non-glycosylated BSA. Cry3A, which is not active on *Diabrotica* species, shows similar but even greater binding to glycosylated-BSA. Similarly, Cry3A shows greater binding to immobilized WCRW brush border membrane (BBM) than does
15 WT Cry3Bb, suggesting that much of the observed binding is non-productive. It was reasoned that the non-specific binding to WCRW BBM occurs via glycosylated proteins, and that binding to both glycosylated-BSA and WCRW BBM is non-productive in reaction pathway to toxicity. Therefore reduction or elimination of that binding would lead to enhanced binding to the productive receptor and to
20 enhanced toxicity. Potential binding sites for sugar groups were targeted for redesign to reduce the non-specific binding of Cry3Bb to glycoproteins and to immobilized WCRW BBM.

5.17.1 CRY3BB.60

25 Cry3Bb-60, in which Cry3Bb has been cleaved at R159 in 1 α 3,4, shows decreased binding to glycosylated-BSA and decreased binding to immobilized WCRW BBM. Cry3Bb-60 shows a 3.6-fold increase in bioactivity relative to WT Cry3Bb.

5.17.2 ALTERATIONS TO 1 α 3,4 (CRY3Bb.11221)

Cry3Bb.11221 has been redesigned in the 1 α 3,4 region of domain 1, which is the region in which Cry3Bb is cleaved to produce Cry3Bb-60. Cry3Bb.11221 also shows decreased binding to both glycosylated-BSA and immobilized WCRW
5 BBM, and exhibits a 6.4-fold increase in bioactivity over that of WT Cry3Bb. Together with data for Cry3Bb.60 (section 5.17.1) these data suggest that this loop region contributes substantially to non-productive binding of the toxin.

5.17.3 ALTERATION TO 1 β 1, α 8 (CRY3Bb.11228,11230,11237 AND 11231)

10 The 1 β 1, α 8 region of Cry3Bb has been re-engineered to increase hydration (section 4.2.4) and enhance flexibility (section 4.4.3). Several proteins altered in this region, Cry3Bb.11228,11230, and 11237 demonstrate substantially lower levels of binding both glycosylated-BSA and immobilized WCRW BBM, and also show between 4.1- and 4.5-fold increases in bioactivity relative to WT Cry3Bb.

15

5.17.4 BINDING ACTIVITY

The tendencies of Cry3Bb and some of its derivatives to bind to glycosylated-BSA and to WCRW BBM were determined using a BIAcore™ surface plasmon resonance biosensor. For glycosylated-BSA binding, the glycosylated protein
20 was immobilized using standard NHS chemistry to a CM5 chip (BIAcore), and the solubilized toxin was injected over the glycosylated-BSA surface. To measure binding to WCRW BBM, brush border membrane vesicles (BBMV) purified from WCRW midguts (English *et al.*, 1991) were immobilized on an HPA chip (BIAcore) then washed with either 10mM KOH or with 40mM β -octylglucoside.
25 The solubilized toxin was then injected over the resulting hybrid bilayer surface to detect binding. Protein concentration were determined by Protein Dye Reagent assay (BioRad) or BCA Protein Assay (Pierce).

Other methods may also be used to determine the same binding information. These include, but are not limited to, ligand blot experiments using labeled

toxin, labeled glycosylated protein, or anti-toxin antibodies, affinity chromatography, and *in vitro* binding of toxin to intact BBMV.

5 **5.18 EXAMPLE 18 -- CONSTRUCTION OF PLASMIDS WITH WT *CRY3Bb* SEQUENCES**

Standard recombinant DNA procedures were performed essentially as described by Sambrook *et al.*, (1989).

5.18.1 PEG1701

10 pEG1701 (FIG. 11), contained in EG11204 and EG11037, was constructed by inserting the *SphI*-*PstI* fragment containing the *cry3Bb* gene and the *cryIF* terminator from pEG911 (Baum, 1994) into the *SphI*-*PstI* site of pEG854.9 (Baum *et al.*, 1996), a high copy number *B. thuringiensis* - *E. coli* shuttle vector.

15 **5.18.2 PEG1028**

pEG1028 contains the *HindIII* fragment of *cry3Bb* from pEG1701 cloned into the multiple cloning site of pTZ18U at *HindIII*.

20 **5.19 EXAMPLE 19 -- CONSTRUCTION OF PLASMIDS WITH ALTERED *CRY3Bb* GENES**

Plasmid DNA from *E. coli* was prepared by the alkaline lysis method (Maniatis *et al.*, 1982) or by commercial plasmid preparation kits (examples: PERFECTprep™ kit, 5 Prime - 3 Prime, Inc., Boulder CO; QIAGEN plasmid prep kit, QIAGEN Inc.). *B. thuringiensis* plasmids were prepared from cultures grown in
25 brain heart infusion plus 0.5% glycerol (BHIG) to mid logarithmic phase by the alkaline lysis method. When necessary for purification, DNA fragments were excised from an agarose gel following electrophoresis and recovered by glass milk using a GeneClean II® kit (BIO 101 Inc., La Jolla, CA). Alteration of the *cry3Bb* gene was accomplished using several techniques including site-directed
30 mutagenesis, triplex PCR™, quasi-random PCR™ mutagenesis, DNA shuffling and standard recombinant techniques. These techniques are described in Sections

6.1, 6.2, 6.3, 6.4 and 6.5, respectively. The DNA sequences of primers used are listed in Section 7.

5.20 EXAMPLE 20 -- SITE-DIRECTED MUTAGENESIS

5 Site-directed mutagenesis was conducted by the protocols established by Kunkle (1985) and Kunkle *et al.* (1987) using the Muta-Gene™ M13 *in vitro* mutagenesis kit (Bio-Rad, Richmond, CA). Combinations of alterations to *cry3Bb* were accomplished by using the Muta-Gene™ kit and multiple mutagenic oligonucleotide primers.

10

5.20.1 pEG1041

pEG1041, contained in EG11032, was constructed using the Muta-Gene™ kit, primer C, and single-stranded pEG1028 as the DNA template. The resulting altered *cry3Bb* DNA sequence was excised as a *Pf*MI DNA fragment and used to
15 replace the corresponding DNA fragment in pEG1701.

5.20.2 pEG1046

pEG1046, contained in EG11035, was constructed using the Muta-Gene™ kit, primer D, and single-stranded pEG1028 as the DNA template. The resulting
20 altered *cry3Bb* DNA sequence was excised as a *Pf*MI DNA fragment and used to replace the corresponding DNA fragment in pEG1701.

5.20.3 pEG1047

pEG1047, contained in EG11036, was constructed using the Muta-Gene™
25 kit, primer E, and single-stranded pEG1028 as the DNA template. The resulting altered *cry3Bb* DNA sequence was excised as a *Pf*MI DNA fragment and used to replace the corresponding DNA fragment in pEG1701.

5.20.4 pEG1052

pEG1052, contained in EG11046, was constructed using the Muta-Gene™ kit, primers D and E, and single-stranded pEG1028 as the DNA template. The resulting altered *cry3Bb* DNA sequence was excised as a *Pf*MI DNA fragment and
5 used to replace the corresponding DNA fragment in pEG1701.

5.20.5 pEG1054

pEG1054, contained in EG11048, was constructed using the Muta-Gene™ kit, primer F, and single-stranded pEG1028 as the DNA template. The resulting
10 altered *cry3Bb* DNA sequence was excised as a *Pf*MI DNA fragment and used to replace the corresponding DNA fragment in pEG1701.

5.20.6 pEG1057

pEG1057, contained in EG11051, was constructed using the Muta-Gene™
15 kit, primer G, and single-stranded pEG1028 as the DNA template. The resulting altered *cry3Bb* DNA sequence was excised as a *Pf*MI DNA fragment and used to replace the corresponding DNA fragment in pEG1701.

5.21 EXAMPLE 21 -- TRIPLEX PCR™

20 Triplex PCR™ is described by Michael (1994). This method makes use of a thermostable ligase to incorporate a phosphorylated mutagenic primer into an amplified DNA fragment during PCR™. PCR™ was performed on a Perkin Elmer Cetus DNA Thermal Cycler (Perkin-Elmer, Norwalk, CT) using a AmpliTaq™ DNA polymerase kit (Perkin-Elmer) and *Sph*I-linearized pEG1701 as the template
25 DNA. PCR™ products were cleaned using commercial kits such as Wizard™ PCR™ Preps (Promega, Madison, WI) and QIAquick PCR™ Purification kit (QIAGEN Inc., Chatsworth, CA).

5.21.1 PEG1708 AND PEG1709

pEG1708 and pEG1709, contained in EG11222 and EG11223, respectively, were constructed by replacing the *Pfl*MI-*Pfl*MI fragment of *cry3Bb* in pEG1701 with *Pfl*MI-digested and gel purified PCR™ fragment altered at *cry3Bb* nucleotide positions 688-690, encoding amino acid Y230. Random mutations were introduced into the Y230 codon by triplex PCR™. Mutagenic primer MVT095 was phosphorylated and used together with outside primer pair FW001 and FW006. Primer MVT095 also contains a silent mutation at position 687, changing T to C, which, upon incorporation, introduces an additional *Eco*RI site into pEG1701.

5.21.2 PEG1710, PEG1711 AND PEG1712

Plasmids pEG1710, pEG1711 and pEG1712, contained in EG11224, EG11225 and EG11226, respectively, were created by replacing the *Pfl*MI-*Pfl*MI fragment of the *cry3Bb* gene in pEG1701 with *Pfl*MI-digested and gel purified PCR™ fragment altered at *cry3Bb* nucleotide positions 690-692, encoding H231. Random mutations were introduced into the H231 codon by triplex PCR™. Mutagenic primer MVT097 was phosphorylated and used together with outside primer pair FW001 and FW006. Primer MVT097 also contains a T to C sequence change at position 687 which, upon incorporation, results in an additional *Eco*RI site by silent mutation.

5.21.3 PEG1713 AND PEG1727

pEG1713 and pEG1727, contained in EG11227 and EG11242, respectively, were constructed by replacing the *Pfl*MI-*Pfl*MI fragment of the *cry3Bb* gene in pEG1701 with *Pfl*MI-digested and gel purified PCR™ fragment altered at *cry3Bb* nucleotide positions 868-870, encoding amino acid R290. Triplex PCR™ was used to introduce random changes into the R290 codon. The mutagenic primer, MVT091, was designed so that the nucleotide substitutions would result in

approximately 36% of the sequences encoding amino acids D or E. MVT091 was phosphorylated and used together with outside primer pair FW001 and FW006.

5.22 EXAMPLE 22 -- QUASI-RANDOM PCRTM MUTAGENESIS

5 Quasi-random mutagenesis combines the mutagenic PCRTM techniques described by Vallette *et al.* (1989), Tomic *et al.* (1990) and LaBean and Kauffman (1993). Mutagenic primers, sometimes over 70 nucleotides in length, were designed to introduce changes over nucleotide positions encoding for an entire structural region, such as a loop. Degenerate codons typically consisted of a ratio of
10 82% WT nucleotide plus 6% each of the other 3 nucleotides per position to semi-randomly introduce changes over the target region (LaBean and Kauffman, 1993). When possible, natural restriction sites were utilized; class 2s enzymes were used when natural sites were not convenient (Stemmer and Morris, 1992, list additional restriction enzymes useful to this technique). PCRTM was performed on a Perkin
15 Elmer Cetus DNA Thermal Cycler (Perkin-Elmer, Norwalk, CT) using a AmpliTaqTM DNA polymerase kit (Perkin-Elmer) and *SphI*-linearized pEG1701 as the template DNA. Quasi-random PCRTM amplification was performed using the following conditions: denaturation at 94°C for 1.5 min.; annealing at 50°C for 2 min. and extension at 72°C for 3 min., for 30 cycles. The final 14 extension cycles were
20 extended an additional 25 s per cycle. Primers concentration was 20 µM per reaction or 40 µM for long, mutagenic primers. PCRTM products were cleaned using commercial kits such as WizardTM PCRTM Preps (Promega, Madison, WI) and QIAquick PCRTM Purification kit (QIAGEN Inc., Chatsworth, CA). In some instances PCRTM products were treated with Klenow Fragment (Promega) following
25 the manufacturer's instructions to fill in any single base overhangs prior to restriction digestion.

5.22.1 pEG1707

EG1707, contained in EG11221, was constructed by replacing the *PfIMI*-
30 *PfIMI* fragment of the *cry3Bb* gene in pEG1701 with *PfIMI*-digested and gel purified PCRTM fragment altered at *cry3Bb* nucleotide positions 460-480, encoding

10 α 3,4 amino acids 154-160. Primer MVT075, which includes a recognition site for the class 2s restriction enzyme *Bsa*I, and primer FW006 were used to introduce changes into this region by quasi-random mutagenesis. Primers MVT076, also containing a *Bsa*I site, and primer FW001 were used to PCRTM amplify a "linker" fragment. Following PCRTM amplification, both products were cleaned, end-filled, digested with *Bsa*I and ligated to each other. Ligated fragment was gel purified and used as template for PCRTM amplification using primer pair FW001 and FW006. PCRTM product was cleaned, digested with *Pfl*MI, gel purified and ligated into *Pfl*MI-digested and purified pEG1701 vector DNA.

10

5.22.2 PEG1720 AND PEG1726

pEG1720 and pEG1726, contained in EG11234 and EG11241, respectively, were constructed by replacing the *Pfl*MI-*Pfl*MI fragment of the *cry3Bb* gene in pEG1701 with *Pfl*MI-digested and gel purified PCRTM fragment altered at *cry3Bb* nucleotide positions 859-885, encoding α 7, β 1 amino acids 287-295. Quasi-random PCRTM mutagenesis was used to introduce changes into this region. Mutagenic primer MVT111, designed with a *Bsa*I site, and primer FW006 were used to introduce the changes. Primer pair MVT094, also containing a *Bsa*I site, and FW001 were used to amplify the linker fragment. The PCRTM products were digested with *Bsa*I, gel purified then ligated to each other. Ligated product was PCRTM amplified using primer pair FW001 and FW006, digested with *Pfl*MI.

20

5.22.3 PEG1714, PEG1715, PEG1716, PEG1718, PEG1719, PEG1722, PEG1723, PEG1724 AND PEG1725

25 pEG1714, pEG1715, pEG1716, pEG1718, pEG1719, pEG1722, pEG1723, pEG1724 and pEG1725, contained in EG11228, EG11229, EG11230, EG11232, EG11233, EG11236, EG11237, EG11238 and EG11239, respectively, were constructed by replacing the *Pfl*MI-*Pfl*MI fragment of the *cry3Bb* gene in pEG1701 with *Pfl*MI-digested and gel purified PCRTM fragment altered at *cry3Bb* nucleotide positions 931-954, encoding β 1, α 8 amino acids 311-318. Quasi-random PCRTM

30

mutagenesis was used to introduce changes into this region using mutagenic primer MVT103 and primer FW006. Primers FW001 and FW006 were used to amplify a linker fragment. The PCRTM products were end-filled using Klenow and digested with *Bam*HI. The larger fragment from the FW001-FW006 digest was gel purified
5 then ligated to the digested MVT103-FW006 fragment. Ligated product was gel purified and amplified by PCRTM using primer pair FW001 and FW006. The amplified product was digested with *Pf*MI and gel purified prior to ligation into *Pf*MI-digested and purified pEG1701 vector DNA.

10 5.22.4 pEG1701.Lβ2,3

Plasmids carrying alterations of *cry3Bb* WT sequence at nucleotides 1051-1065, encoding structural region lβ2,3 of Cry3Bb, were constructed by replacing the *Mlu*I-*Spe*I fragment of pEG1701 with isolated *Mlu*I- and *Spe*I-digested PCRTM product. The PCRTM product was generated by quasi-random PCRTM mutagenesis
15 were mutagenic primer MVT081 was paired with FW006. These plasmids as a group are designated pEG1701.lβ2,3.

5.22.5 pEG1701.Lβ6,7

Plasmids containing mutations of the *cry3Bb* WT sequence at nucleotides
20 1234-1248, encoding structural region lβ6,7 of Cry3Bb, were constructed by replacing the *Mlu*I-*Spe*I fragment of pEG1701 with isolated *Mlu*I- and *Spe*I-digested PCRTM product. The PCRTM product was generated by quasi-random PCRTM mutagenesis where mutagenic primer MVT085 was paired with primer WD115. Primer pair MVT089 and WD112 were used to amplify a linker fragment. Both
25 PCRTM products were digested with *Taq*I and ligated to each other. The ligation product was gel purified and PCRTM amplified using primer pair MVT089 and FW006. The amplified product was digested with *Mlu*I and *Spe*I and ligated into *Mlu*I and *Spe*I digested and purified pEG1701 vector DNA. These plasmids as a group are designated pEG1701.lβ6,7.

5.22.6 pEG1701.L β 10,11

Plasmids containing mutated *cry3Bb* sequences at nucleotides 1450-1467, encoding structural region l β 10,11 of Cry3Bb, were constructed by replacing the *SpeI-PstI* fragment of pEG1701 with isolated *SpeI*- and *PstI*-digested PCRTM product. The PCRTM product was generated by quasi-random PCRTM mutagenesis where mutagenic primer MVT105 was paired with primer MVT070. Primer pair MVT092 and MVT083 were used to generate a linker fragment. (MVT083 is a mutagenic oligo designed for another region. The sequence changes introduced by MVT083 are removed following restriction digestion and do not impact the alteration of *cry3Bb* in the l β 10,11 region.) Both PCRTM products were digested with *BsaI*, ligated together, and the ligation product PCRTM amplified with primer pair MVT083 and MVT070. The resulting PCRTM product was digested with *SpeI* and *PstI*, and gel purified. These plasmids as a group are designated pEG1701.l β 10,11.

5.23 EXAMPLE 23 -- DNA SHUFFLING

DNA-shuffling, as described by Stemmer (1994), was used to combine individual alterations in the *cry3Bb* gene.

5.23.1 pEG1084, pEG1085, pEG1086 AND pEG1087

pEG1084, pEG1085, pEG1086, and pEG1087, contained in EG11081, EG11082, EG11083, and EG11084, respectively, were recovered from DNA-shuffling. Briefly, *Pfu*MI DNA fragments were generated using primer set A and B and each of the plasmids pEG1707, pEG1714, pEG1715, pEG1716, pEG1041, pEG1046, pEG1047, and pEG1054 as DNA templates. The resulting DNA fragments were pooled in equal-molar amounts and digested with DNaseI and 50-100 bp DNA fragments were recovered from an agarose gel by three successive freeze-thaw cycles: three min in a dry-ice ethanol bath followed by complete thawing at 50°C. The recovered DNA fragments were assembled by primerless-PCRTM and PCRTM-amplified using the primer set A and B as described by Stemmer (1994).

The final PCRTM-amplified DNA fragments were cut with *Pfl*MI and used to replace the corresponding *cry3Bb Pfl*MI DNA fragment in pEG1701.

5.24 EXAMPLE 24 -- RECOMBINANT DNA TECHNIQUES

5 Standard recombinant DNA procedures were performed essentially as described by Sambrook *et al.* (1989).

5.24.1 pEG1717

10 pEG1717, contained in EG11231, was constructed by replacing the small *Bgl*II fragment of pEG1710 with the small *Bgl*II fragment from pEG1714.

5.24.2 pEG1721

15 pEG1721, contained in EG11235, was constructed by replacing the small *Bgl*II fragment from pEG1710 with the small *Bgl*II fragment from pEG1087.

5.24.3 pEG1063

20 pEG1062, contained in EG11057, was constructed by replacing the *Nco*I DNA fragment containing *ori* 43 from pEG1054 with the isolated *Nco*I DNA fragment containing *ori* 43 and the alterations in *cry3Bb* from pEG1046.

5.24.4 pEG1063

25 pEG1063, contained in EG11058, was constructed by replacing the *Nco*I DNA fragment containing *ori* 43 from pEG1054 with the isolated *Nco*I DNA fragment containing *ori* 43 and the alterations in *cry3Bb* from pEG1707.

5.24.5 pEG1095

30 pEG1095, contained in EG11095, was constructed by replacing the *Mlu*I-*Spe*I DNA fragment in pEG1701 with the corresponding *Mlu*I-*Spe*I DNA fragment from pEG1086.

5.25.7 PRIMER MVT076 (SEQ ID NO:77):

5'-GGTCACCTAGGTCTCTCTTCCAGGAATTTAACGCATTAAC-3'

5.25.8 PRIMER MVT111 (SEQ ID NO:78):

5
5'-
AGCTATGCTGGTCTCCCATTTJEHIEJEJJEIHKRRJEHEIJEENIIIGTTAAAAC
AGAACTAAC-3'

5.25.9 PRIMER MVT094 (SEQ ID NO:79):

10 5'-ATCCAGTGGGGTCTCAAATGGGAAAAGTACAATTAG-3'

5.25.10 PRIMER MVT103 (SEQ ID NO:80):

5'-
CATTTTTACGGATCCAATTTTJFFFJNEEJEFNFJNFEILEIJEORGACCAAC
15 TTTTTTGAG-3'

5.25.11 PRIMER MVT081 (SEQ ID NO:81):

5'-
GAATTTTCATACGCGTCTTCAACCTGGTJEHJJJINMEEIEJTCTTTCAATTA
20 TTGGTCTGG-3'

5.25.12 PRIMER MVT085 (SEQ ID NO:82):

5'-
AAAAGTTTATCGAACTATAGCTAATACAGACGTAGCGGCTJQQFFNEEJII
25 JEEIGTATATTTAGGTGTTACG-3'

5.25.13 PRIMER A (SEQ ID NO:83) 3B2PFLM1:

5'-GGAGTTCCATTTGCTGGGGC-3'

5.25.14 PRIMER B (SEQ ID NO:84) 3B2PFLM2:

5'-ATCTCCATAAAAATGGGG-3'

5.25.15 PRIMER C (SEQ ID NO:85) 3B2165DG:

5 5'-GCGAAGTAAAAGAAGCCAAGGTCGAATAAGGG-3'

5.25.16 PRIMER D (SEQ ID NO:86) 3B2160SKRD:

5'-

CCTTTAAGTTTGCAGAAATCCACACAGCCAAGGTCGAATAAGGG-3'

10

5.25.17 PRIMER E (SEQ ID NO:87) 3B2290VP:

5'-CCCATTTTATGATGTTCGGTTATACCCAAAAGGGG-3'

5.25.18 PRIMER F (SEQ ID NO:88) 3B2EdA104:

15 5'-GGCCAAGTGAAGACCCATGGAAGGC-3'

5.25.19 PRIMER G (SEQ ID NO:89) 3B2KG189:

5'-GCAGTTTCCGGATTCGAAGTGC-3'

20 **5.25.20 PRIMER WD112 (SEQ ID NO:90):**

5'-CCGCTACGTCTGTATTA-3'

5.25.21 PRIMER WD115 (SEQ ID NO:91):

5'-ATAATGGAAGCACCTGA-3'

25

5.25.22 PRIMER MVT105 (SEQ ID NO:92):

5'-

AGCTATGCTGGTCTCTTCTTAEJIFEIIEFFIJFIJINACAATTCCATTTTTTAC
TTGG-3'

5.25.23 PRIMER MVT092 (SEQ ID NO:93):

5'-ATCCAGTTGGGTCTCTAAGAAACAAACCGCGTAATTAAGC-3'

5 5.25.24 PRIMER MVT070 (SEQ ID NO:94):

5'-CCTCAAGGGTTATAACATCC-3'

5.25.25 PRIMER MVT083 (SEQ ID NO:95):

5'-

10 GTACAAAAGCTAAGCTTTIEJIINPEEMEEIJNJESCGAACTATAGCTAATA
CAG-3'

5.26 EXAMPLE 26 -- SEQUENCE ANALYSIS OF ALTERED *CRY3Bb* GENES

E. coli DH5 α TM (GIBCO BRL, Gaithersburg, MD), JM110 and SureTM
15 (Stratagene, La Jolla, CA) cells were sometimes used amplify plasmid DNA for
sequencing. Plasmids were transformed into these cells using the manufacturers'
procedures. DNA was sequenced using the Sequenase® 2.0 DNA sequencing kit
purchased from U. S. Biochemical Corporation (Cleveland, Ohio). The plasmids
described in Section 6, their respective divergence from WT *cry3Bb* sequence, the
20 resulting amino acid changes and the protein structure site of the changes are listed
in Table 11.

TABLE 11
DNA SEQUENCE CHANGES OF *CRY3Bb** GENES AND RESULTING AMINO ACID SUBSTITUTIONS OF THE *CRY3Bb** PROTEINS

Plasmid	<i>cry3Bb</i> * DNA Sequence	<i>Cry3Bb</i> * Amino Acid Sequence	Structural Site of Alteration
pEG1707	A460T, C461T, A462T, C464A, T465C, T466C, T467A, A468T, A469T, G470C, T472C, T473G, G474T, A477T, A478T, G479C	T154F, P155H, L156H, L158R	1 α 3,4
pEG1708	T687C, T688C, A689T, C691A, A692G	Y230L, H231S	α 6
pEG1709	T667C, T687C, T688A, A689G, C691A, A692G	S223P, Y230S	α 6
pEG1710	T687C, A692G	H231R	α 6
pEG1711	T687C, C691A	H231N, T241S	α 6
pEG1712	T687C, C691A, A692C, T693C	H231T	α 6
pEG1713	C868A, G869A, G870T	R290N	1 α 7, β 1
pEG1714	C932T, A938C, T942G, G949A, T954C	S311L, N313T, E317K	1 β 1, α 8
pEG1715	T931A, A933C, T942A, T945A, G949A, A953G, T954C	S311T, E317K, Y318C	1 β 1, α 8
pEG1716	T931G, A933C, C934G, T945G, C946T, A947G, G951A, T954C	S311A, L312V, Q316W	1 β 1, α 8

TABLE 11 (CONT'D)

Plasmid	<i>cry3Bb</i> * DNA Sequence	Cry3Bb* Amino Acid Sequence	Structural Site of Alteration
pEG1717	T687C, A692G, C932T, A938C, T942G, G949A, T954C	H231R, S311L, N313T, E317K	$\alpha 6$, $\text{I}\beta 1$, $\alpha 8$
pEG1718	T931A, A933G, T935C, T936A, A938C, T939C, T942C, T945A, G951T, T954C	S311T, L312P, N313T, E317N	$\text{I}\beta 1$, $\alpha 8$
pEG1719	T931G, A933C, T936G, T942C, C943T, T945A, C946G, G948C, T954C	S311A, Q316D	$\text{I}\beta 1$, $\alpha 8$
pEG1720	T861C, T866C, C868A, T871C, T872G, A875T, T877A, C878G, A882G	I289T, L291R, Y292F, S293R	$\text{I}\alpha 7$, $\beta 1$
pEG1721	T687C, A692G, C932T	H231R, S311L	$\alpha 6$, $\text{I}\beta 1$, $\alpha 8$
pEG1722	T931A, C932T, A933C, T936C, T942G, T945A, T954C	S311I	$\text{I}\beta 1$, $\alpha 8$
pEG1723	T931A, C932T, A933C, T936C, A937G, A938T, C941A, T942C, T945A, C946A, A947T, A950T, T954C	S311I, N313H	$\text{I}\beta 1$, $\alpha 8$
pEG1724	A933C, T936C, A937G, A938T, C941A, T942C, T945A, C946A, A947T, A950T, T954C	N313V, T314N, Q316M, E317V	$\text{I}\beta 1$, $\alpha 8$

TABLE 11 (CONT'D)

Plasmid	<i>cry3Bb</i> * DNA Sequence	Cry3Bb* Amino Acid Sequence	Structural Site of Alteration
pEG1725	A933T, A938G, T939G, T942A, T944C, T945A, A947T, G948T, A950C, T954C	N313R, L315P, Q316L, E317A	I β 1, α 8
pEG1726	A860T, T861C, G862A, C868T, G869T, T871C, A873T, T877A, C878G, A879T	Y287F, D288N, R290L	I α 7, β 1
pEG1727	C868G, G869T	R290V	I α 7, β 1
pEG1041	A494G	D165G	α 4
pEG1046	G479A, A481C, A482C, A484C, G485A, A486C, A494G	S160N, K161P, P162H, D165G	α 4
pEG1047	A865G, T877C	I289V, S293P	I α 7, β 1
pEG1052	G479A, A481C, A482C, A484C, G485A, A486C, A494G, A865G, T877C	S160N, K161P, P162H, D165G, I289V, S293P	α 4, I α 7, β 1
pEG1054	T309A, Δ 310, Δ 311, Δ 312	D103E, Δ A104	I α 2a,2b
pEG1057	A565G, A566G	K189G	I α 4,5
pEG1062	T309A, Δ 310, Δ 311, Δ 312, G479A, A481C, A482C, A484C, G485A, A486C, A494G	D103E, Δ A104, S160N, K161P, P162H, D165G	I α 2a,2b α 4

TABLE 11 (CONT'D)

Plasmid	<i>cry3Bb</i> * DNA Sequence	Cry3Bb* Amino Acid Sequence	Structural Site of Alteration
pEG1063	T309A, Δ310, Δ311, Δ312, A460T, C461T, A462T, C464A, T465C, T466C, T467A, A468T, A469T, G470C, T472C, T473G, G474T, A477T, A478T, G479C	D103E, ΔA104, T154F, P155H, L156H, L158R	1α2a, 2b 1α3, 4
pEG1084	A494G, T931A, A933C, T942A, T945A, G949A, T954C	D165G, S311T, E317K	α4, 1β1, α8
pEG1085	A494G, A865G, T877C, T914C, T931G, A933C, C934G, T945G, C946T, A947G, G951A, T954C, A1043G, T1094C	D165G, I289V, S293P, F305S, S311A, L312V, Q316W, Q348R, V365A	α4, 1α7, β1 1β1, 1β1, α8 β2, β3b
pEG1086	A865G, T877C, A1043G	I289V, S293P, Q348R	1α7, β1, β2
pEG1087	A494G, C932T	D165G, S311L	α4, 1β1, α8
pEG1095	A1043G	Q348R	β2

5.27 EXAMPLE 27 -- EXPRESSION OF CRY3Bb* PROTEINS

5.27.1 CULTURE CONDITIONS

LB agar was prepared using a standard formula (Maniatis *et al.*, 1982). Starch agar was obtained from Difco Laboratories (Detroit, MI) and supplemented with an additional 5 g/l of agar. C2 liquid medium is described by Donovan *et al.* (1988). C2 medium was sometimes prepared without the phosphate buffer (C2-P). All cultures were incubated at 25°C to 30°C; liquid cultures were also shaken at 250 rpm, until sporulation and lysis had occurred.

10 5.27.2 TRANSFORMATION CONDITIONS

pEG1701 and derivatives thereof were introduced into acrySTALLIFEROUS *B. thuringiensis* var. *kurstaki* EG7566 (Baum, 1994) or EG10368 (U. S. Patent 5,322,687) by the electroporation method of Macaluso and Mettus (1991). In some cases, the method was modified as follows to maximize the number of transformants.

15 The recipient *B. thuringiensis* strain was inoculated from overnight growth at 30°C on LB agar into brain heart infusion plus 0.5% glycerol, grown to an optical density of approximately 0.5 at 600 nm, chilled on ice for 10 min, washed 2X with EB and re-suspended in a 1/50 volume of EB. Transformed cells were selected on LB agar or starch agar plus 5 µg/ml chloramphenicol. Visual screening of colonies was used to

20 identify transformants producing crystalline protein; those colonies were generally more opaque than colonies that did not produce crystalline protein.

5.27.3 STRAIN AND PROTEIN DESIGNATIONS

A transformant containing an altered *cry3Bb** gene encoding an altered

25 Cry3Bb* protein is designated by an "EG" number, *e.g.*, EG11231. The altered Cry3Bb* protein is designated Cry3Bb followed by the strain number, *e.g.*, Cry3Bb.11231. Collections of proteins with alterations at a structural site are designated Cry3Bb followed by the structural site, *e.g.*, Cry3Bb.lβ2,3. Table 12 lists the plasmids pertinent to this invention, the new *B. thuringiensis* strains containing the

plasmids, the acrySTALLIFEROUS *B. thuringiensis* recipient strain used, and the proteins produced by the new strains.

5.28 EXAMPLE 28 -- GENERATION AND CHARACTERIZATION OF CRY3BB-60

5 5.28.1 GENERATION OF CRY3BB-60

Cry3Bb-producing strain EG7231 (U. S. Patent 5,187,091) was grown in C2 medium plus 3 mg/ml chloramphenicol. Following sporulation and lysis, the culture was washed with water and Cry3Bb protein purified by the NaBr solubilization and recrystallization method of Cody *et al.* (1992). Protein concentration was determined
10 by BCA Protein Assay (Pierce, Rockford, IL). Recrystallized protein was solubilized in 10 ml of 50 mM KOH per 100 mg of Cry3Bb protein and buffered to pH 9.0 with 100 mM CAPS (3-[cyclohexylamino]-1-propanesulfonic acid), pH 9.0. The soluble toxin was treated with trypsin at a weight ratio of 50 mg toxin to 1 mg trypsin for 20 min to overnight at room temperature. Trypsin cleaves proteins on the carboxyl side
15 of available arginine and lysine residues. For 8-dose bioassay, the solubilization conditions were altered slightly to increase the concentration of protein: 50 mM KOH was added dropwise to 2.7 ml of a 12.77 mg/ml suspension of purified Cry3Bb* until crystal solubilization occurred. The volume was then adjusted to 7 ml with 100 mM CAPS, pH 9.0.

TABLE 12
PLASMIDS CARRYING ALTERED *CRY3Bb GENES TRANSFORMED INTO**
B. THURINGIENSIS
FOR EXPRESSION OF ALTERED *CRY3Bb PROTEINS**

Plasmid Designation	New BT Strain	Expressed Protein
pEG1701	EG11204	WT Cry3Bb
pEG1701	EG11037	WT Cry3Bb
pEG1707	EG11221	Cry3Bb.11221
pEG1708	EG11222	Cry3Bb.11222
pEG1709	EG11223	Cry3Bb.11223
pEG1710	EG11224	Cry3Bb.11224
pEG1711	EG11225	Cry3Bb.11225
pEG1712	EG11226	Cry3Bb.11226
pEG1713	EG11227	Cry3Bb.11227
pEG1714	EG11228	Cry3Bb.11228
pEG1715	EG11229	Cry3Bb.11229
pEG1716	EG11230	Cry3Bb.11230
pEG1717	EG11231	Cry3Bb.11231
pEG1718	EG11232	Cry3Bb.11232
pEG1719	EG11233	Cry3Bb.11233
pEG1720	EG11234	Cry3Bb.11234
pEG1721	EG11235	Cry3Bb.11235
pEG1722	EG11236	Cry3Bb.11236
pEG1723	EG11237	Cry3Bb.11237
pEG1724	EG11238	Cry3Bb.11238
pEG1725	EG11239	Cry3Bb.11239
pEG1726	EG11241	Cry3Bb.11241
pEG1727	EG11242	Cry3Bb.11242
pEG1041	EG11032	Cry3Bb.11032
pEG1046	EG11035	Cry3Bb.11035
pEG1047	EG11036	Cry3Bb.11036

TABLE 12 (CONT'D)

Plasmid Designation	New BT Strain	Expressed Protein
pEG1052	EG11046	Cry3Bb.11046
pEG1054	EG11048	Cry3Bb.11048
pEG1057	EG11051	Cry3Bb.11051
pEG1062	EG11057	Cry3Bb.11057
pEG1063	EG11058	Cry3Bb.11058
pEG1084	EG11081	Cry3Bb.11081
pEG1085	EG11082	Cry3Bb.11082
pEG1086	EG11083	Cry3Bb.11083
pEG1087	EG11084	Cry3Bb.11084
pEG1095	EG11095	Cry3Bb.11095
pEG1098	EG11098	Cry3Bb.11098
pEG1701.lβ2,3	collection of unnamed strains	Cry3Bb.lβ2,3
pEG1701.lβ6,7	collection of unnamed strains	Cry3Bb.lβ6,7
pEG1701.lβ10,11	collection of unnamed strains	Cry3Bb.lβ10,11

5.28.2 DETERMINATION OF MOLECULAR WEIGHT OF CRY3BB-60

The molecular weight of the predominant trypsin digestion fragment of Cry3Bb was determined to be 60 kDa by SDS-polyacrylamide gel electrophoresis (SDS-PAGE) analysis using commercial molecular weight markers. This digestion fragment is designated Cry3Bb-60. No further digestion of the 60 kDa cleavage product was observed.

10 5.28.3 DETERMINATION OF NH₂-TERMINUS OF CRY3BB-60

To determine the NH₂-terminal sequence of Cry3Bb-60, the trypsin digest was fractionated by SDS-PAGE and transferred to Immobilon™-P membrane (Millipore Corporation, Bedford, MA) following standard western blotting procedures. After transfer, the membrane was rinsed twice with water then stained with 0.025% Coomassie Brilliant Blue R-250 plus 40% methanol for 5 min, destained with 50%

methanol and rinsed in water. The Cry3Bb.60 band was excised with a razor blade. NH₂-terminal sequencing was performed at the Tufts Medical School, Department of Physiology (Boston, MA) using standard automated Edman degradation procedures. The NH₂-terminal amino acid sequence was determined to be SKRSQDR (SEQ ID
5 NO:96), corresponding to amino acids 160-166 of Cry3Bb. Trypsin digestion occurred on the carboxyl side of amino acid R159 resulting in the removal of helices 1-3.

5.29 EXAMPLE 29 -- BIOACTIVITY OF CRY3BB* PROTEINS

10 5.29.1 CULTURE CONDITIONS AND PROTEIN CONCENTRATION DETERMINATION

Cultures for 1-dose bioassays were grown in C2-P plus 5 µg/ml chloramphenicol (C2-P/cm5) then diluted with 3 volumes of 0.005% Triton X-100[®]. The protein concentrations of these cultures were not determined. Cultures for 8-dose bioassays were grown in C2/cm5, washed 1 - 2 times with 1 - 2 volumes of sterile water
15 and resuspended in 1/10 volume of sterile 0.005% Triton X-100[®]. The toxin protein concentration of each concentrate was determined as described by Brussock and Currier (1990), omitting the treatment with 3 M HEPES. The protein concentration was adjusted to 3.2 mg/ml in 0.005% Triton X-100[®] for the top dose of the assay. Cry3Bb.60 was produced and quantified for 8-dose assay as described in Section 9.1.

20

5.29.2 INSECT BIOASSAYS

Diabrotica undecimpunctata howardi Barber (southern corn rootworm or SCRW) and *Diabrotica virgifera virgifera* LeConte (western corn rootworm or WCRW) larvae were reared as described by Slaney *et al.* (1992). Eight-dose assays
25 and probit analyses were performed as described by Slaney *et al.* (1992). Thirty-two larvae were tested per dose at 50 µl of sample per well of diet (surface area of 175 mm²). Positive controls were WT Cry3Bb-producing strains EG11037 or EG11204. All bioassays were performed using 128-well trays containing approximately 1 ml of diet per well with perforated mylar sheet covers (C-D International Inc., Pitman, NJ).

One-dose assays were performed essentially the same except only 1 dose was tested per strain. All assay were replicated at least twice.

5.29.3 INSECT BIOASSAY RESULTS: 1-DOSE ASSAYS AGAINST SCRW

5 Results from 1-dose assays are expressed as the relative mortality (RM) of the experimental strain compared to WT (% mortality of experimental culture divided by % mortality of WT culture). Altered and improved Cry3Bb proteins derived from plasmids constructed using PCR™ methods introducing random or semi-random changes into the *cry3Bb* gene sequence were distinguished from other altered but not
10 improved Cry3Bb proteins by replicated, 1-dose assay against SCRW larvae. Those proteins showing increased activity (defined as $RM \geq 1.5$) compared to WT Cry3Bb or, in the case of proteins with combinations of altered sites, compared to a “parental” altered Cry3Bb protein were further characterized by 8-dose assay. The overall RM “pattern” produced by 1-dose assay results from a collection of proteins carrying ran-
15 dom or semi-random alterations within a single structural region, *e.g.*, in $\text{I}\beta 2,3$, can be used to determine if that structural region is important for bioactivity. Retention of WT levels of activity ($RM \approx 1$) indicate changes are tolerated in that region. Overall loss of activity ($RM < 1$) distinguishes the region as important for bioactivity.

20 5.29.4 CRY3BB. $\text{I}\beta 2,3$: RESULTS OF 1-DOSE BIOASSAYS AGAINST SCRW

Cry3Bb. $\text{I}\beta 2,3$ protein are a collection of proteins altered in the $\text{I}\beta 2,3$ region of Cry3Bb (see Section 5.3.4). Typical results of 1-dose assays of these altered proteins are shown in FIG. 12. The RM values for Cry3Bb. $\text{I}\beta 2,3$ proteins are less than 1, with a few exceptions of values close to 1, indicating that this region is important for tox-
25 icity.

5.29.5 CRY3BB. $\text{I}\beta 6,7$: RESULTS OF 1-DOSE BIOASSAYS AGAINST SCRW

Cry3Bb. $\text{I}\beta 6,7$ proteins are a collection of proteins altered in the $\text{I}\beta 6,7$ region of Cry3Bb (see Section 5.3.5). Typical results of 1-dose assays of these altered proteins
30 are shown in FIG. 13. With a few exceptions of values close to 1, the RM values for

Cry3Bb.lβ6,7 proteins are less than 1, indicating that this region is important for toxicity.

5.29.6 CRY3BB.lβ10,11: RESULTS OF 1-DOSE BIOASSAYS AGAINST SCR W

5 Cry3Bb.lβ10,11 proteins are a collection of proteins altered in the lβ10,11 region of Cry3Bb (see Section 5.3.6). Typical results of 1-dose assays of these altered proteins are shown in FIG. 14. With a few exceptions of values close to 1, the RM values for Cry3Bb.lβ10,11 proteins are less than 1, indicating that this region is important for bioactivity.

10

5.29.7 INSECT BIOASSAY RESULTS: RESULTS OF 8-DOSE ASSAYS AGAINST SCR W

Results from 8-dose assays are expressed as an LC₅₀ value (protein concentration giving 50% mortality) with 95% confidence intervals. The LC₅₀ values with 95% confidence intervals of altered Cry3Bb proteins showing improved activities against
15 SCR W larvae and LC₅₀ values of the WT Cry3Bb control determined at the same time are listed in Table 13 along with the fold increase over WT activity for each improved protein.

TABLE 13

20 DESIGNED CRY3BB PROTEINS WERE TESTED AGAINST SCR W LARVAE IN REPLICATED, 8-DOSE ASSAYS TO DETERMINE THE LC₅₀ VALUES

LC ₅₀ μg/well (95% C.I.)			
Improved Protein	Improved Protein	WT Cry3Bb Control	Fold Increase Over WT Activity
Cry3Bb.60	6.7 (5.3-8.4)	24.1 (15-39)	3.6×
Cry3Bb.11221	3.2 (2.5-4)	20.5 (14.5-29)	6.4×
Cry3Bb.11222	7.3 (6-9)	29.4 (23-37)	4.0×
Cry3Bb.11223	10.5 (9-12)	29.4 (23-37)	2.8×
Cry3Bb.11224	6.5 (5.1-8.2)	32.5 (25-43)	5.0×
Cry3Bb.11225	13.7 (11-16.8)	49.5 (39-65)	3.6×

TABLE 13 (CONT'D)

LC ₅₀ µg/well (95% C.I.)			
Improved Protein	Improved Protein	WT Cry3Bb Control	Fold Increase Over WT Activity
Cry3Bb.11226	16.7 (10.6-24.2)	49.5 (39-65)	3.0×
Cry3Bb.11227	11.1 (9.1-13.5)	21.3 (16-28)	1.9×
Cry3Bb.11228	8.0 (6.6-9.8)	32.9 (25-45)	4.1×
Cry3Bb.11229	7.2 (5.8-8.8)	18.2 (15-22)	2.5×
Cry3Bb.11230	7.0 (5.8-8.6)	32.9 (25-45)	4.7×
Cry3Bb.11231	3.3 (3.0-3.7)	26.1 (22-31)	7.9×
Cry3Bb.11232	6.4 (5.4-7.7)	32.9 (25-45)	5.1×
Cry3Bb.11233	15.7 (12-20)	32.9 (25-45)	2.2×
Cry3Bb.11234	7 (6-9)	29 (22-39)	4.1×
Cry3Bb.11235	4.2 (3.6-4.9)	13.3 (10-17)	3.2×
Cry3Bb.11236	11.6 (9-15)	36.4 (27-49)	3.1×
Cry3Bb.11237	6.8 (4-11)	36.4 (27-49)	5.4×
Cry3Bb.11238	13.9 (11-17)	36.4 (27-49)	2.6×
Cry3Bb.11239	13.0 (10-16)	36.4 (27-49)	2.8×
Cry3Bb.11241	11 (7-16)	29 (22-39)	2.6×
Cry3Bb.11242	11.9 (9.2-16)	30 (23-38)	2.5×
Cry3Bb.11032	4.2 (3.6-4.9)	13.3 (10-17)	3.1×
Cry3Bb.11035	10.3 (8-13)	27.9 (23-34)	2.7×
Cry3Bb.11036	6.5 (5.1-7.9)	27.9 (23-34)	4.3×
Cry3Bb.11046	12.1 (8-19)	31.2 (25-39)	2.6×
Cry3Bb.11048	8.3 (6-11)	35.4 (24-53)	4.3×
Cry3Bb.11051	11.8 (8-16)	35.4 (24-53)	3.0×
Cry3Bb.11057	8.8 (7-11)	29.5 (24-36)	3.4×
Cry3Bb.11058	9.6 (6-14)	33.4 (27-43)	3.5×
Cry3Bb.11081	8.5 (7-11)	51.5 (37-79)	6.1×
Cry3Bb.11082	10.6 (8-13)	51.5 (37-79)	4.9×
Cry3Bb.11083	7.0 (5-10)	51.5 (37-79)	7.4×

TABLE 13 (CONT'D)

Improved Protein	LC ₅₀ µg/well (95% C.I.)		
	Improved Protein	WT Cry3Bb Control	Fold Increase Over WT Activity
Cry3Bb.11084	7.2 (4-12)	51.5 (37-79)	7.2×
Cry3Bb.11095	11.1 (9-14)	51.5 (37-79)	4.6×
Cry3Bb.11098			

5.29.8 Insect Bioassay Results: 8-Dose Assays Against WCRW

WCRW larvae are delicate and difficult to work with. Therefore, only some of the designed Cry3Bb showing improved activity against SCRW larvae were also tested against WCRW larvae in 8-dose assays. The LC₅₀ determinations for the designed Cry3Bb proteins are shown in Table 14 along with the LC₅₀ values of the WT Cry3Bb control determined at the same time.

TABLE 14

**CRY3BB* PROTEINS SHOWING IMPROVED ACTIVITY AGAINST SCRW LARVAE
ALSO SHOW IMPROVED ACTIVITY AGAINST WCRW LARVAE**

Improved Protein	LC ₅₀ µg/well (95% C.I.)		
	Improved Protein	WT Cry3Bb Control	Fold Increase Over WT Activity
EG11083	6.3 (4.7-8.2)	63.5 (46-91)	10.1×
EG11230	24.2 (13-40)	4.5 (2.1-7.4)	5.4×
EG11231	32.2 (14-67)	2.5 (1.7-3.6)	12.9×

5.30 EXAMPLE 30 -- CHANNEL ACTIVITY

Ion channels produced by Cry3Bb and some of its derivatives were measured by the methods described by Slatin *et al.* (1990). In some instances, lipid bilayers were prepared from a mixture of 4:1 phosphatidylethanolamine (PE) : phosphatidylcholine (PC). Toxin protein was solubilized from washed, C2 medium,

B. thuringiensis cultures with 12 mM KOH. Following centrifugation to remove spores and other debris, 10 µg of soluble toxin protein was added to the *cis* compartment (4.5 ml volume) of the membrane chamber. Protein concentration was determined using the BCA Protein Assay (Pierce).

5

5.30.1 CHANNEL ACTIVITY OF WT CRY3Bb

Upon exposure to black lipid membranes, Cry3Bb forms ion channels with various conductance states. The channels formed by Cry3Bb are rarely discrete channels with well resolved open and closed states and usually require incubation of the toxin with the membrane for 30 - 45 min before any channel-like events are observed. After formation of the initial conductances, the size increases from approximately 200 pS to over 10,000 pS over 2 - 3 h. Only the small conductances (≤ 200 pS) are voltage dependent. Over 200 pS, the conductances are completely symmetric. Cry3Bb channels also exhibit β -mercaptoethanol-dependent activation, growing from small channel conductances of ~ 200 pS to several thousand pS within 2 min of the addition of β -mercaptoethanol to the *cis* compartment of the membrane chamber.

10

15

5.30.2 CRY3Bb.11032

The channel activity of Cry3Bb.11032 is much like WT Cry3Bb when the solubilized toxin protein is added to the *cis* compartment of the membrane chamber. However, when this protein is artificially incorporated into the membrane by forming or "painting" the membrane in the presence of the Cry3Bb.11032 protein, a 16-fold increase in the initial channel conductances is observed (~ 4000 pS). This phenomenon is not observed with WT Cry3Bb.

20

25

5.30.3 CRY3Bb.11035

Upon exposure to artificial membranes, the Cry3Bb.11035 protein spontaneously forms channels that grow to large conductances within a relatively short time span (~ 5 min). Conductance values range from 3000 - 6000 pS and, like WT Cry3Bb, are voltage dependent at low conductance values.

30

5.30.4 CRY3Bb.11048

The Cry3Bb.11048 protein is quite different than WT Cry3Bb in that it appears not to form channels at all, but, rather, forms symmetrical pores with respect to voltage. Once the pore is formed, it remains open and allows a steady conductance ranging from 25 to 130 pS.

5.30.5 CRY3Bb.11224 AND CRY3Bb.11226

The metal binding site of WT Cry3Bb formed by H231 in the dimer structure was removed in proteins Cry3Bb.11224 and Cry3Bb.11226. The conductances formed by both designed proteins are identical to that of WT Cry3Bb with the exception that neither of the designed proteins exhibits β -mercaptoethanol-dependent activation.

5.30.6 CRY3Bb.11221

Cry3Bb.11221 protein has been observed to immediately form small channels of 100 - 200 pS with limited voltage dependence. Some higher conductances were observed at the negative potential. In other studies, the onset of activity was delayed by 27 min, which is more typical for WT Cry3Bb. Unlike WT Cry3Bb, however, Cry3Bb.11221 forms well resolved, 600 pS channels with long open states. The protein eventually reaches conductances of 7000 pS.

5.30.7 CRY3Bb.11242

Cry3Bb.11242 protein forms small conductances immediately upon exposure to an artificial membrane. The conductances grow steadily and rapidly to 6000 pS in approximately 3 min. Some voltage dependence was noted with a preference for a negative imposed voltage.

5.30.8 CRY3Bb.11230

Unlike WT Cry3Bb, Cry3Bb.11230 forms well resolved channels with long open states that do not continue to grow in conductance with time. The maximum observed channel conductances reached 3000 pS. FIG. 15 illustrates the difference between the channels formed by Cry3Bb and Cry3Bb.11230.

5.30.9 CRY3Bb.60

Cry3Bb.60 forms well resolved ion channels within 20 min of exposure to an artificial membrane. These channels grow in conductance and frequency with time. The behavior of Cry3Bb.60 in a planar lipid bilayer differs from Cry3Bb in two significant ways. The conductances created by Cry3Bb.60 form more quickly than Cry3Bb and, unlike Cry3Bb, the conductances are stable, having well resolved open and closed states definitive of stable ion channels (FIG. 16).

5.31 EXAMPLE 31 -- PRIMER COMPOSITIONS**TABLE 15**

SEQ ID NO:83	% of Nucleotide in mixture			
Code	A	T	G	C
N	25	25	25	25

TABLE 16

SEQ ID NO:84	% of Nucleotide in mixture			
Code	A	T	G	C
N	25	25	25	25

TABLE 17

SEQ ID NO:85	% of Nucleotide in mixture			
Code	A	T	G	C
B	16	16	52	16
D	70	10	10	10
N	25	25	25	25

TABLE 18

SEQ ID NO:86	% of Nucleotide in mixture			
Code	A	T	G	C
E	82	6	6	6
F	6	6	6	82
J	6	82	6	6
I	6	6	82	6
N	25	25	25	25

5

TABLE 19

SEQ ID NO:88	% of Nucleotide in mixture			
Code	A	T	G	C
J	6	82	6	6
E	82	6	6	6
H	1	1	1	97
I	6	6	82	6
K	15	15	15	55
R	15	55	15	15

TABLE 20

SEQ ID NO:90	% of Nucleotide in mixture			
Code	A	T	G	C
J	6	82	6	6
F	6	6	6	82
N	25	25	25	25
E	82	6	6	6
I	6	6	82	6
L	8	1	83	8
O	1	1	1	97

TABLE 21

SEQ ID NO:91	% of Nucleotide in mixture			
Code	A	T	G	C
J	6	82	6	6
E	82	6	6	6
H	1	1	1	97
I	6	6	82	6
N	25	25	25	25
M	82	2	8	8

TABLE 22
SEQ ID NO:92

Code	% of Nucleotide in mixture			
	A	T	G	C
J	6	82	6	6
Q	0	9	82	9
F	6	6	6	82
N	25	25	25	25
E	82	6	6	6
I	6	6	82	6

TABLE 23
SEQ ID NO:92

Code	% of Nucleotide in mixture			
	A	T	G	C
J	6	82	6	6
F	6	6	6	82
N	25	25	25	25
E	82	6	6	6
I	6	6	82	6

TABLE 24
SEQ ID NO:95

Code	% of Nucleotide in mixture			
	A	T	G	C
J	6	82	6	6
N	25	25	25	25
E	82	6	6	6
I	6	6	82	6
M	82	2	8	8
P	8	2	8	82
S	1	97	1	1

5.32 EXAMPLE 32 -- ATOMIC COORDINATES FOR CRY3Bb

5 The atomic coordinates of the Cry3Bb protein are given in the Appendix included in Section 9.1

5.33 EXAMPLE 33 -- ATOMIC COORDINATES FOR CRY3A

10 The atomic coordinates of the Cry3A protein are given in the Appendix included in Section 9.2

5.34 EXAMPLE-34 -- MODIFICATION OF *CRY* GENES FOR EXPRESSION IN PLANTS

15 Wild-type *cry* genes are known to be expressed poorly in plants as a full length gene or as a truncated gene. Typically, the G+C content of a *cry* gene is low (37%) and often contains many A+T rich regions, potential polyadenylation sites and numerous ATTTA sequences. Table 25 shows a list of potential polyadenylation sequences which should be avoided when preparing the "plantized" gene construct.

TABLE 25
LIST OF SEQUENCES OF THE POTENTIAL POLYADENYLATION SIGNALS

AATAAA*	AAGCAT
AATAAT*	ATTAAT
AACCAA	ATACAT
ATATAA	AAAATA
AATCAA	ATTAAA**
ATACTA	AATTAA**
ATAAAA	AATACA**
ATGAAA	CATAAA**

* indicates a potential major plant polyadenylation site.

** indicates a potential minor animal polyadenylation site.

5 All others are potential minor plant polyadenylation sites.

The regions for mutagenesis may be selected in the following manner. All regions of the DNA sequence of the *cry* gene are identified which contained five or more consecutive base pairs which were A or T. These were ranked in terms of length and highest percentage of A+T in the surrounding sequence over a 20-30 base pair region. The DNA is analysed for regions which might contain polyadenylation sites or ATTTA sequences. Oligonucleotides are then designed which maximize the elimination of A+T consecutive regions which contained one or more polyadenylation sites or ATTTA sequences. Two potential plant polyadenylation sites have been shown to be more critical based on published reports. Codons are selected which increase G+C content, but do not generate restriction sites for enzymes useful for cloning and assembly of the modified gene (*e.g.*, *Bam*HI, *Bgl*II, *Sac*I, *Nco*I, *Eco*RV, *etc.*). Likewise condons are avoided which contain the doublets TA or GC which have been reported to be infrequently-found codons in plants.

20 Although the CaMV35S promoter is generally a high level constitutive promoter in most plant tissues, the expression level of genes driven the CaMV35S promoter is low in floral tissue relative to the levels seen in leaf tissue. Because the eco-

nomically important targets damaged by some insects are the floral parts or derived from floral parts (*e.g.*, cotton squares and bolls, tobacco buds, tomato buds and fruit), it is often advantageous to increase the expression of crystal proteins in these tissues over that obtained with the CaMV35S promoter.

5 The 35S promoter of Figwort Mosaic Virus (FMV) is analogous to the CaMV35S promoter. This promoter has been isolated and engineered into a plant transformation vector. Relative to the CaMV promoter, the FMV 35S promoter is highly expressed in the floral tissue, while still providing similar high levels of gene
10 expression in other tissues such as leaf. A plant transformation vector, may be constructed in which the full length synthetic *cry* gene is driven by the FMV 35S promoter. Tobacco plants may be transformed with the vector and compared for expression of the crystal protein by Western blot or ELISA immunoassay in leaf and floral tissue. The FMV promoter has been used to produce relatively high levels of crystal protein in floral tissue compared to the CaMV promoter.

15

5.35 EXAMPLE 35 -- EXPRESSION OF SYNTHETIC *CRY* GENES WITH SS RUBISCO PROMOTERS AND CHLOROPLAST TRANSIT PEPTIDES

 The genes in plants encoding the small subunit of RUBISCO (SSU) are often highly expressed, light regulated and sometimes show tissue specificity. These ex-
20 pression properties are largely due to the promoter sequences of these genes. It has been possible to use SSU promoters to express heterologous genes in transformed plants. Typically a plant will contain multiple SSU genes, and the expression levels and tissue specificity of different SSU genes will be different. The SSU proteins are encoded in the nucleus and synthesized in the cytoplasm as precursors that contain an
25 N-terminal extension known as the chloroplast transit peptide (CTP). The CTP directs the precursor to the chloroplast and promotes the uptake of the SSU protein into the chloroplast. In this process, the CTP is cleaved from the SSU protein. These CTP sequences have been used to direct heterologous proteins into chloroplasts of transformed plants.

The SSU promoters might have several advantages for expression of heterologous genes in plants. Some SSU promoters are very highly expressed and could give rise to expression levels as high or higher than those observed with the CaMV35S promoter. The tissue distribution of expression from SSU promoters is different from that of the CaMV35S promoter, so for control of some insect pests, it may be advantageous to direct the expression of crystal proteins to those cells in which SSU is most highly expressed. For example, although relatively constitutive, in the leaf the CaMV35S promoter is more highly expressed in vascular tissue than in some other parts of the leaf, while most SSU promoters are most highly expressed in the mesophyll cells of the leaf. Some SSU promoters also are more highly tissue specific, so it could be possible to utilize a specific SSU promoter to express the protein of the present invention in only a subset of plant tissues, if for example expression of such a protein in certain cells was found to be deleterious to those cells. For example, for control of Colorado potato beetle in potato, it may be advantageous to use SSU promoters to direct crystal protein expression to the leaves but not to the edible tubers.

Utilizing SSU CTP sequences to localize crystal proteins to the chloroplast might also be advantageous. Localization of the *B. thuringiensis* crystal proteins to the chloroplast could protect these from proteases found in the cytoplasm. This could stabilize the proteins and lead to higher levels of accumulation of active toxin. *cry* genes containing the CTP could be used in combination with the SSU promoter or with other promoters such as CaMV35S.

5.36 EXAMPLE 36 -- TARGETING OF CRY* PROTEINS TO THE EXTRACELLULAR SPACE OR VACUOLE THROUGH THE USE OF SIGNAL PEPTIDES

The *B. thuringiensis* proteins produced from the synthetic genes described here are localized to the cytoplasm of the plant cell, and this cytoplasmic localization results in plants that are insecticidally effective. It may be advantageous for some purposes to direct the *B. thuringiensis* proteins to other compartments of the plant cell. Localizing *B. thuringiensis* proteins in compartments other than the cytoplasm may result in less exposure of the *B. thuringiensis* proteins to cytoplasmic proteases lead-

ing to greater accumulation of the protein yielding enhanced insecticidal activity. Extracellular localization could lead to more efficient exposure of certain insects to the *B. thuringiensis* proteins leading to greater efficacy. If a *B. thuringiensis* protein were found to be deleterious to plant cell function, then localization to a noncyto-
5 plasmic compartment could protect these cells from the protein.

In plants as well as other eukaryotes, proteins that are destined to be localized either extracellularly or in several specific compartments are typically synthesized with an N-terminal amino acid extension known as the signal peptide. This signal peptide directs the protein to enter the compartmentalization pathway, and it is typi-
10 cally cleaved from the mature protein as an early step in compartmentalization. For an extracellular protein, the secretory pathway typically involves cotranslational insertion into the endoplasmic reticulum with cleavage of the signal peptide occurring at this stage. The mature protein then passes through the Golgi body into vesicles that fuse with the plasma membrane thus releasing the protein into the extracellular space.
15 Proteins destined for other compartments follow a similar pathway. For example, proteins that are destined for the endoplasmic reticulum or the Golgi body follow this scheme, but they are specifically retained in the appropriate compartment. In plants, some proteins are also targeted to the vacuole, another membrane bound compartment in the cytoplasm of many plant cells. Vacuole targeted proteins diverge from the
20 above pathway at the Golgi body where they enter vesicles that fuse with the vacuole.

A common feature of this protein targeting is the signal peptide that initiates the compartmentalization process. Fusing a signal peptide to a protein will in many cases lead to the targeting of that protein to the endoplasmic reticulum. The efficiency of this step may depend on the sequence of the mature protein itself as well.
25 The signals that direct a protein to a specific compartment rather than to the extracellular space are not as clearly defined. It appears that many of the signals that direct the protein to specific compartments are contained within the amino acid sequence of the mature protein. This has been shown for some vacuole targeted proteins, but it is not yet possible to define these sequences precisely. It appears that secretion into the
30 extracellular space is the "default" pathway for a protein that contains a signal sequence but no other compartmentalization signals. Thus, a strategy to direct

B. thuringiensis proteins out of the cytoplasm is to fuse the genes for synthetic *B. thuringiensis* genes to DNA sequences encoding known plant signal peptides. These fusion genes will give rise to *B. thuringiensis* proteins that enter the secretory pathway, and lead to extracellular secretion or targeting to the vacuole or other compartments.

Signal sequences for several plant genes have been described. One such sequence is for the tobacco pathogenesis related protein PR1b has been previously described (Cornelissen *et al.*, 1986). The PR1b protein is normally localized to the extracellular space. Another type of signal peptide is contained on seed storage proteins of legumes. These proteins are localized to the protein body of seeds, which is a vacuole like compartment found in seeds. A signal peptide DNA sequence for the β -subunit of the 7S storage protein of common bean (*Phaseolus vulgaris*), PvuB has been described (Doyle *et al.*, 1986). Based on the published these published sequences, genes may be synthesized chemically using oligonucleotides that encode the signal peptides for PR1b and PvuB. In some cases to achieve secretion or compartmentalization of heterologous proteins, it may be necessary to include some amino acid sequence beyond the normal cleavage site of the signal peptide. This may be necessary to insure proper cleavage of the signal peptide.

5.37 EXAMPLE 37 -- ISOLATION OF TRANSGENIC MAIZE RESISTANT TO *DIABROTICA* SPP. USING CRY3BB VARIANTS

5.37.1 PLANT GENE CONSTRUCTION

The expression of a plant gene which exists in double-stranded DNA form involves transcription of messenger RNA (mRNA) from one strand of the DNA by RNA polymerase enzyme, and the subsequent processing of the mRNA primary transcript inside the nucleus. This processing involves a 3' non-translated region which adds polyadenylate nucleotides to the 3' end of the RNA. Transcription of DNA into mRNA is regulated by a region of DNA usually referred to as the "promoter". The promoter region contains a sequence of bases that signals RNA polymerase to associ-

ate with the DNA and to initiate the transcription of mRNA using one of the DNA strands as a template to make a corresponding strand of RNA.

A number of promoters which are active in plant cells have been described in the literature. Such promoters may be obtained from plants or plant viruses and include, but are not limited to, the nopaline synthase (NOS) and octopine synthase (OCS) promoters (which are carried on tumor-inducing plasmids of *Agrobacterium tumefaciens*), the cauliflower mosaic virus (CaMV) 19S and 35S promoters, the light-inducible promoter from the small subunit of ribulose 1,5-bisphosphate carboxylase (ssRUBISCO, a very abundant plant polypeptide), and the Figwort Mosaic Virus (FMV) 35S promoter. All of these promoters have been used to create various types of DNA constructs which have been expressed in plants (see *e.g.*, U. S. Patent No. 5,463,175, specifically incorporated herein by reference).

The particular promoter selected should be capable of causing sufficient expression of the enzyme coding sequence to result in the production of an effective amount of protein. One set of preferred promoters are constitutive promoters such as the CaMV35S or FMV35S promoters that yield high levels of expression in most plant organs (U. S. Patent No. 5,378,619, specifically incorporated herein by reference). Another set of preferred promoters are root enhanced or specific promoters such as the CaMV derived 4 as-1 promoter or the wheat POX1 promoter (U. S. Patent No. 5,023,179, specifically incorporated herein by reference; Hertig *et al.*, 1991). The root enhanced or specific promoters would be particularly preferred for the control of corn rootworm (*Diabroticus* spp.) in transgenic corn plants.

The promoters used in the DNA constructs (*i.e.* chimeric plant genes) of the present invention may be modified, if desired, to affect their control characteristics. For example, the CaMV35S promoter may be ligated to the portion of the ssRUBISCO gene that represses the expression of ssRUBISCO in the absence of light, to create a promoter which is active in leaves but not in roots. The resulting chimeric promoter may be used as described herein. For purposes of this description, the phrase "CaMV35S" promoter thus includes variations of CaMV35S promoter, *e.g.*, promoters derived by means of ligation with operator regions, random or controlled

mutagenesis, *etc.* Furthermore, the promoters may be altered to contain multiple “enhancer sequences” to assist in elevating gene expression.

The RNA produced by a DNA construct of the present invention also contains a 5' non-translated leader sequence. This sequence can be derived from the promoter
5 selected to express the gene, and can be specifically modified so as to increase translation of the mRNA. The 5' non-translated regions can also be obtained from viral RNA's, from suitable eucaryotic genes, or from a synthetic gene sequence. The present invention is not limited to constructs wherein the non-translated region is derived from the 5' non-translated sequence that accompanies the promoter sequence.

10 For optimized expression in monocotyledenous plants such as maize, an intron should also be included in the DNA expression construct. This intron would typically be placed near the 5' end of the mRNA in untranslated sequence. This intron could be obtained from, but not limited to, a set of introns consisting of the maize *hsp70* intron (U. S. Patent No. 5,424,412; specifically incorporated herein by reference) or the rice
15 *Act1* intron (McElroy *et al.*, 1990). As shown below, the maize *hsp70* intron is useful in the present invention.

As noted above, the 3' non-translated region of the chimeric plant genes of the present invention contains a polyadenylation signal which functions in plants to cause the addition of adenylate nucleotides to the 3' end of the RNA. Examples of preferred
20 3' regions are (1) the 3' transcribed, non-translated regions containing the polyadenylate signal of *Agrobacterium* tumor-inducing (Ti) plasmid genes, such as the nopaline synthase (NOS) gene and (2) plant genes such as the pea ssRUBISCO E9 gene (Fischhoff *et al.*, 1987).

25 5.37.2 PLANT TRANSFORMATION AND EXPRESSION

A chimeric plant gene containing a structural coding sequence of the present invention can be inserted into the genome of a plant by any suitable method. Suitable plant transformation vectors include those derived from a Ti plasmid of *Agrobacterium tumefaciens*, as well as those disclosed, *e.g.*, by Herrera-Estrella (1983), Bevan
30 (1983), Klee (1985) and Eur. Pat. Appl. Publ. No. EP0120516. In addition to plant transformation vectors derived from the Ti or root-inducing (Ri) plasmids of *Agrobac-*

terium, alternative methods can be used to insert the DNA constructs of this invention into plant cells. Such methods may involve, for example, the use of liposomes, electroporation, chemicals that increase free DNA uptake, free DNA delivery *via* microprojectile bombardment, and transformation using viruses or pollen (Fromm *et al.*, 1986; Armstrong *et al.*, 1990; Fromm *et al.*, 1990).

5.37.3 CONSTRUCTION OF MONOCOT PLANT EXPRESSION VECTORS FOR *CRY3BB* VARIANTS

5.37.3.1 DESIGN OF *CRY3BB* VARIANT GENES FOR PLANT EXPRESSION

For efficient expression of the *cry3Bb* variants in transgenic plants, the gene encoding the variants must have a suitable sequence composition (Diehn *et al.*, 1996). One example of such a sequence is shown for the v11231 gene (SEQ ID NO:99) which encodes the Cry3Bb11231 variant protein (SEQ ID NO:100) with *Diabrotica* activity. This gene was derived *via* mutagenesis (Kunkel, 1985) of a *cry3Bb* synthetic gene (SEQ ID NO:101) encoding a protein essentially homologous to the protein encoded by the native *cry3Bb* gene (Gen Bank Accession Number m89794, SEQ ID NO:102). The following oligonucleotides were used in the mutagenesis of the original *cry3Bb* synthetic gene (SEQ ID NO:101) to create the v11231 gene (SEQ ID NO:99):

- Oligo #1:
5'-TAGGCCTCCATCCATGGCAAACCCTAACAATC-3' (SEQ ID NO:103)
- Oligo #2:
5'-TCCCATCTTCCTACTTACGACCCTGCAGAAATACGGTCCAAC -3'
(SEQ ID NO:104)
- Oligo #3:
5'-GACCTCACCTACCAAACATTCGATCTTG -3' (SEQ ID NO:105)
- Oligo #4:
5'-CGAGTTCTACCGTAGGCAGCTCAAG-3' (SEQ ID NO:106)

5.37.3.2 CONSTRUCTION OF CRY3Bb MONOCOT PLANT EXPRESSION VECTOR

To place the *cry3Bb* variant gene v11231 in a vector suitable for expression in monocotyledonous plants (*i.e.* under control of the enhanced Cauliflower Mosaic Virus 35S promoter and link to the *hsp70* intron followed by a nopaline synthase polyadenylation site as in U. S. Patent No. 5,424,412, specifically incorporated herein by reference), the vector pMON19469 was digested with *NcoI* and *EcoRI*. The larger vector band of approximately 4.6 kb was electrophoresed, purified, and ligated with T4 DNA ligase to the *NcoI-EcoRI* fragment of approximately 2 kb containing the v11231 gene (SEQ ID NO:99). The ligation mix was transformed into *E. coli*, carbenicillin resistant colonies recovered and plasmid DNA recovered by DNA miniprep procedures. This DNA was subjected to restriction endonuclease analysis with enzymes such as *NcoI* and *EcoRI* (together), *NotI*, and *PstI* to identify clones containing pMON33708 (the v11231 coding sequence fused to the *hsp70* intron under control of the enhanced CaMV35S promoter).

To place the v11231 gene in a vector suitable for recovery of stably transformed and insect resistant plants, the 3.75-kb *NotI* restriction fragment from pMON33708 containing the lysine oxidase coding sequence fused to the *hsp70* intron under control of the enhanced CaMV35S promoter was isolated by gel electrophoresis and purification. This fragment was ligated with pMON30460 treated with *NotI* and calf intestinal alkaline phosphatase (pMON30460 contains the neomycin phosphotransferase coding sequence under control of the CaMV35S promoter). Kanamycin resistant colonies were obtained by transformation of this ligation mix into *E. coli* and colonies containing pMON33710 identified by restriction endonuclease digestion of plasmid miniprep DNAs. Restriction enzymes such as *NotI*, *EcoRV*, *HindIII*, *NcoI*, *EcoRI*, and *BglII* can be used to identify the appropriate clones containing the *NotI* fragment of pMON33708 in the *NotI* site of pMON30460 (*i.e.* pMON33710) in the orientation such that both genes are in tandem (*i.e.* the 3' end of the v11231 expression cassette is linked to the 5' end of the *nptII* expression cassette). Expression of the v11231 protein by pMON33710 in corn protoplasts was confirmed by electroporation of pMON33710 DNA into protoplasts followed by protein blot and ELISA analysis. This vector can be introduced into the genomic DNA of corn em-

bryos by particle gun bombardment followed by paromomycin selection to obtain corn plants expressing the v11231 gene essentially as described in U. S. Patent No. 5,424,412, specifically incorporated herein by reference.

5 In this example, the vector was introduced *via* cobombardment with a hygromycin resistance conferring plasmid into immature embryo scutella (IES) of maize, followed by hygromycin selection, and regeneration. Transgenic corn lines expressing the v11231 protein were identified by ELISA analysis. Progeny seed from these events were subsequently tested for protection from *Diabrotica* feeding.

10 5.37.3.3 *IN PLANTA* PERFORMANCE OF CRY3BB.11231

Transformed corn plants expressing Cry3Bb.11231 protein were challenged with western corn rootworm (WCR) larvae in both a seedling and 10 inch pot assay. The transformed genotype was A634, where the progeny of the R0 cross by A634 was evaluated. Observations included effect on larval development (weight), root damage
15 rating (RDR), and protein expression. The transformation vector containing the *cry3Bb* gene was pMON33710. Treatments included the positive and negative isopopulations for each event and an A634 check.

The seedling assay consisted of the following steps: (i) single seeds were placed in 1 oz cups containing potting soil; (ii) at spiking, each seedling was infested
20 with 4 neonate larvae; and (iii) after infestation, seedlings were incubated for 7 days at 25°C, 50% RH, and 14:10 (L:D) photo period. Adequate moisture was added to the potting soil during the incubation period to maintain seedling vigor.

The 10 inch pot assay consisted of the following steps: (i) single seeds were placed in 10 inch pots containing potting soil; (ii) at 14 days post planting, each pot
25 was infested with 800 eggs which have been pre-incubated such that hatch would occur 5-7 days post infestation; and (iii) after infestation, plants were incubated for 4 weeks under the same environmental conditions as the seedling assay. Pots were both sub and top irrigated daily.

For the seedling assay, on day 7 plants were given a root damage rating, and
30 surviving larvae were weighed. Also at this time, Cry3Bb protein concentrations in the roots were determined by ELISA. The scale used for the seedling assay to assess

root damage is as follows: RDR (root damage rating) 0 = no visible feeding; RDR 1 = very light feeding; RDR 2 = light feeding; RDR 3 = moderate feeding; RDR 4 = heavy feeding; and RDR 5 = very heavy feeding.

Results of the seedling assay are shown in Table 26. Plants expressing
 5 Cry3Bb protein were completely protected by WCR feeding, where surviving larvae within this treatment had not grown. Mean larval weights ranged from 2.03-2.73 mg for the nonexpressing treatments, where the surviving larval average weight was 0.11 mg on the expressing cry3Bb treatment. Root damage ratings were 3.86 and 0.33 for the nonexpressing and expressing isopopulations, respectively. Larval survival
 10 ranged from 75-85% for the negative and check treatments, where only 25% of the larvae survived on the Cry3Bb treatment.

TABLE 26
EFFECT OF CRY3BB EXPRESSING PLANTS ON
WCR LARVAE IN A SEEDLING ASSAY

		Plants			Larvae		
		N	Root		N	%	Mean±SD
Event	Treatment		(ppm)	RDR±SD			Wt. (mg)
16	Negative	7	0.0	3.86±0.65	21	75	2.73±1.67
16	Positive	3	29.01	0.33±0.45	3	25	0.11±0.07
A634	Check	4	0.0	--	13	81	2.03±0.83

For the 10 inch pot assay, at 4 weeks post infestation plant height was recorded and a root damage rating (Iowa 1-6 scale; Hills and Peters, 1971) was given.

Results of the 10 inch pot assay are shown in Table 27. Plants expressing
 20 Cry3Bb protein had significantly less feeding damage and were taller than the non-expressing plants. Event 16, the higher of the two expressing events provided nearly complete control. The negative treatments had very high root damage ratings indicating very high insect pressure. The positive mean root damage ratings were 3.4 and 2.2 for event 6 and 16, respectively. Mean RDR for the negative treatment was 5.0
 25 and 5.6.

TABLE 27
EFFECT OF CRY3BB EXPRESSING CORN IN CONTROLLING
WCR LARVAL FEEDING IN A 10 INCH POT ASSAY

Event	Treatment	N	Root		Plant
			(ppm)	RDR±SD	Height (cm)
6	Negative	7	0.0	5.0±1.41	49.7±18.72
6	Positive	5	7.0	3.4±1.14	73.9±8.67
16	Negative	5	0.0	5.6±0.89	61.2±7.75
16	Positive	5	55.0	2.2±0.84	83.8±7.15

5

In summary, corn plants expressing Cry3Bb protein have a significant biological effect on WCR larval development as seen in the seedling assay. When challenged with very high infestation levels, plants expressing the Cry3Bb protein were protected from WCR larval feeding damage as illustrated in the 10 inch pot assay.

10

6.0 BRIEF DESCRIPTION OF THE SEQUENCE IDENTIFIERS

- SEQ ID NO:1 DNA sequence of *cry3Bb.11221* gene.
- SEQ ID NO:2 Amino acid sequence of Cry3Bb.11221 polypeptide.
- SEQ ID NO:3 DNA sequence of *cry3Bb.11222* gene.
- 15 SEQ ID NO:4 Amino acid sequence of Cry3Bb.11222 polypeptide.
- SEQ ID NO:5 DNA sequence of *cry3Bb.11223* gene.
- SEQ ID NO:6 Amino acid sequence of Cry3Bb.11223 polypeptide.
- SEQ ID NO:7 DNA sequence of *cry3Bb.11224* gene.
- SEQ ID NO:8 Amino acid sequence of Cry3Bb.11224 polypeptide.
- 20 SEQ ID NO:9 DNA sequence of *cry3Bb.11225* gene.
- SEQ ID NO:10 Amino acid sequence of Cry3Bb.11225 polypeptide.
- SEQ ID NO:11 DNA sequence of *cry3Bb.11226* gene.
- SEQ ID NO:12 Amino acid sequence of Cry3Bb.11226 polypeptide.
- SEQ ID NO:13 DNA sequence of *cry3Bb.11227* gene.

	SEQ ID NO:14	Amino acid sequence of Cry3Bb.11227 polypeptide.
	SEQ ID NO:15	DNA sequence of <i>cry3Bb.11228</i> gene.
	SEQ ID NO:16	Amino acid sequence of Cry3Bb.11228 polypeptide.
	SEQ ID NO:17	DNA sequence of <i>cry3Bb.11229</i> gene.
5	SEQ ID NO:18	Amino acid sequence of Cry3Bb.11229 polypeptide.
	SEQ ID NO:19	DNA sequence of <i>cry3Bb.11230</i> gene.
	SEQ ID NO:20	Amino acid sequence of Cry3Bb.11230 polypeptide.
	SEQ ID NO:21	DNA sequence of <i>cry3Bb.11231</i> gene.
	SEQ ID NO:22	Amino acid sequence of Cry3Bb.11231 polypeptide.
10	SEQ ID NO:23	DNA sequence of <i>cry3Bb.11232</i> gene.
	SEQ ID NO:24	Amino acid sequence of Cry3Bb.11232 polypeptide.
	SEQ ID NO:25	DNA sequence of <i>cry3Bb.11233</i> gene.
	SEQ ID NO:26	Amino acid sequence of Cry3Bb.11233 polypeptide.
	SEQ ID NO:27	DNA sequence of <i>cry3Bb.11234</i> gene.
15	SEQ ID NO:28	Amino acid sequence of Cry3Bb.11234 polypeptide.
	SEQ ID NO:29	DNA sequence of <i>cry3Bb.11235</i> gene.
	SEQ ID NO:30	Amino acid sequence of Cry3Bb.11235 polypeptide.
	SEQ ID NO:31	DNA sequence of <i>cry3Bb.11236</i> gene.
	SEQ ID NO:32	Amino acid sequence of Cry3Bb.11236 polypeptide.
20	SEQ ID NO:33	DNA sequence of <i>cry3Bb.11237</i> gene.
	SEQ ID NO:34	Amino acid sequence of Cry3Bb.11237 polypeptide.
	SEQ ID NO:35	DNA sequence of <i>cry3Bb.11238</i> gene.
	SEQ ID NO:36	Amino acid sequence of Cry3Bb.11238 polypeptide.
	SEQ ID NO:37	DNA sequence of <i>cry3Bb.11239</i> gene.
25	SEQ ID NO:38	Amino acid sequence of Cry3Bb.11239 polypeptide.
	SEQ ID NO:39	DNA sequence of <i>cry3Bb.11241</i> gene.
	SEQ ID NO:40	Amino acid sequence of Cry3Bb.11241 polypeptide.
	SEQ ID NO:41	DNA sequence of <i>cry3Bb.11242</i> gene.
	SEQ ID NO:42	Amino acid sequence of Cry3Bb.11242 polypeptide.
30	SEQ ID NO:43	DNA sequence of <i>cry3Bb.11032</i> gene.
	SEQ ID NO:44	Amino acid sequence of Cry3Bb.11032 polypeptide.

	SEQ ID NO:45	DNA sequence of <i>cry3Bb.11035</i> gene.
	SEQ ID NO:46	Amino acid sequence of Cry3Bb.11035 polypeptide.
	SEQ ID NO:47	DNA sequence of <i>cry3Bb.11036</i> gene.
	SEQ ID NO:48	Amino acid sequence of Cry3Bb.11036 polypeptide.
5	SEQ ID NO:49	DNA sequence of <i>cry3Bb.11046</i> gene.
	SEQ ID NO:50	Amino acid sequence of Cry3Bb.11046 polypeptide.
	SEQ ID NO:51	DNA sequence of <i>cry3Bb.11048</i> gene.
	SEQ ID NO:52	Amino acid sequence of Cry3Bb.11048 polypeptide.
	SEQ ID NO:53	DNA sequence of <i>cry3Bb.11051</i> gene.
10	SEQ ID NO:54	Amino acid sequence of Cry3Bb.11051 polypeptide.
	SEQ ID NO:55	DNA sequence of <i>cry3Bb.11057</i> gene.
	SEQ ID NO:56	Amino acid sequence of Cry3Bb.11057 polypeptide.
	SEQ ID NO:57	DNA sequence of <i>cry3Bb.11058</i> gene.
	SEQ ID NO:58	Amino acid sequence of Cry3Bb.11058 polypeptide.
15	SEQ ID NO:59	DNA sequence of <i>cry3Bb.11081</i> gene.
	SEQ ID NO:60	Amino acid sequence of Cry3Bb.11081 polypeptide.
	SEQ ID NO:61	DNA sequence of <i>cry3Bb.11082</i> gene.
	SEQ ID NO:62	Amino acid sequence of Cry3Bb.11082 polypeptide.
	SEQ ID NO:63	DNA sequence of <i>cry3Bb.11083</i> gene.
20	SEQ ID NO:64	Amino acid sequence of Cry3Bb.11083 polypeptide.
	SEQ ID NO:65	DNA sequence of <i>cry3Bb.11084</i> gene.
	SEQ ID NO:66	Amino acid sequence of Cry3Bb.11084 polypeptide.
	SEQ ID NO:67	DNA sequence of <i>cry3Bb.11095</i> gene.
	SEQ ID NO:68	Amino acid sequence of Cry3Bb.11095 polypeptide.
25	SEQ ID NO:69	DNA sequence of <i>cry3Bb.60</i> gene.
	SEQ ID NO:70	Amino acid sequence of Cry3Bb.60 polypeptide.
	SEQ ID NO:71	Primer FW001.
	SEQ ID NO:72	Primer FW006.
	SEQ ID NO:73	Primer MVT095.
30	SEQ ID NO:74	Primer MVT097.
	SEQ ID NO:75	Primer MVT091.

	SEQ ID NO:76	Primer MVT075.
	SEQ ID NO:77	Primer MVT076.
	SEQ ID NO:78	Primer MVT111.
	SEQ ID NO:79	Primer MVT094.
5	SEQ ID NO:80	Primer MVT103.
	SEQ ID NO:81	Primer MVT081.
	SEQ ID NO:82	Primer MVT085.
	SEQ ID NO:83	Primer A.
	SEQ ID NO:84	Primer B.
10	SEQ ID NO:85	Primer C.
	SEQ ID NO:86	Primer D.
	SEQ ID NO:87	Primer E.
	SEQ ID NO:88	Primer F.
	SEQ ID NO:89	Primer G.
15	SEQ ID NO:90	Primer WD112.
	SEQ ID NO:91	Primer WD115.
	SEQ ID NO:92	Primer MVT105.
	SEQ ID NO:93	Primer MVT092.
	SEQ ID NO:94	Primer MVT070.
20	SEQ ID NO:95	Primer MVT083.
	SEQ ID NO:96	N-terminal amino acid of Cry3Bb polypeptide.
	SEQ ID NO:97	DNA sequence of wild-type <i>cry3Bb</i> gene.
	SEQ ID NO:98	Amino acid sequence of wild-type Cry3Bb polypeptide.
	SEQ ID NO:99	Plantized DNA sequence for <i>cry3Bb.11231</i> gene.
25	SEQ ID NO:100	Amino acid sequence of plantized Cry3Bb.11231 polypeptide.
	SEQ ID NO:101	DNA sequence of <i>cry3Bb</i> gene used to prepare SEQ ID NO:99.
	SEQ ID NO:102	DNA sequence of wild-type <i>cry3Bb</i> gene, Genbank
30	#M89794.	
	SEQ ID NO:103	DNA sequence of Oligo #1.

SEQ ID NO:104 DNA sequence of Oligo #2.

SEQ ID NO:105 DNA sequence of Oligo #3.

SEQ ID NO:106 DNA sequence of Oligo #4.

SEQ ID NO:107 DNA sequence of *cry3Bb.11098* gene.

5 SEQ ID NO:108 Amino acid sequence of Cry3Bb.11098 polypeptide.

7.0 REFERENCES

The following references, to the extent that they provide exemplary procedural or other details supplementary to those set forth herein, are specifically incorporated
10 herein by reference.

U. S. Patent 4,237,224, issued December 2, 1980.

U. S. Patent 4,332,898, issued Jun. 1, 1982.

U. S. Patent 4,342,832, issued Aug. 3, 1982.

U. S. Patent 4,356,270, issued Oct. 26, 1982.

15 U. S. Patent 4,362,817, issued Dec. 7, 1982.

U. S. Patent 4,371,625, issued Feb. 1, 1983.

U. S. Patent 4,448,885, issued May 15, 1984.

U. S. Patent 4,467,036, issued Aug. 21, 1984.

U. S. Patent 4,554,101, issued Nov. 19, 1985.

20 U. S. Patent 4,683,195, issued Jul. 28, 1987.

U. S. Patent 4,683,202, issued Jul. 28, 1987.

U. S. Patent 4,757,011, issued Jul. 12, 1988.

U. S. Patent 4,766,203, issued August 23, 1988.

U. S. Patent 4,769,061, issued September 6, 1988.

25 U. S. Patent 4,797,279, issued January 10, 1989.

U. S. Patent 4,800,159, issued January 24, 1989.

U. S. Patent 4,883,750, issued November 28, 1989.

U. S. Patent 4,910,016, issued March 20, 1990.

U. S. Patent 4,940,835, issued Feb. 23, 1990.

30 U. S. Patent 4,965,188, issued Oct. 23, 1990.

U. S. Patent 4,971,908, issued Nov. 20, 1990.

- U. S. Patent 4,987,071, issued Jan. 22, 1991.
- U. S. Patent 5, 380, 831, issued Jan. 10, 1995.
- U. S. Patent 5,023,179, issued June 11, 1991.
- U. S. Patent 5,024,837, issued June 18, 1991.
- 5 U. S. Patent 5,126,133, issued June 30, 1992.
- U. S. Patent 5,176,995, issued Oct. 15, 1991.
- U. S. Patent 5,187,091, issued XXXXX, 1993.
- U. S. Patent 5,322,687, issued Jun. 21, 1994.
- U. S. Patent 5,334,711, issued Aug. 2, 1994.
- 10 U. S. Patent 5,378,619, issued January 3, 1995.
- U. S. Patent 5,424,412, issued June 13, 1995.
- U. S. Patent 5,441,884, issued Aug. 15, 1995.
- U. S. Patent 5,463,175, issued October 31, 1995.
- U. S. Patent 5,500,365, issued Mar 19, 1996.
- 15 U. S. Patent 5,591,616, issued January 7, 1997.
- U. S. Patent 5,631,359, issued May 20, 1997.
- U. S. Patent 5,659,123, issued August 19, 1997.
- Eur. Pat. No. EP 0120516.
- Eur. Pat. No. EP 0360257.
- 20 Eur. Pat. Appl. No. 92110298.4.
- Eur. Pat. Appl. No. 295156A1.
- Great Britain Patent 2202328.
- Int. Pat. Appl. Publ. No. WO 91/03162.
- Int. Pat. Appl. Publ. No. WO 92/07065.
- 25 Int. Pat. Appl. Publ. No. WO 93/15187.
- Int. Pat. Appl. Publ. No. WO 93/23569.
- Int. Pat. Appl. Publ. No. WO 94/02595.
- Int. Pat. Appl. Publ. No. WO 94/13688.
- Intl. Pat. Appl. Publ. No. PCT/US87/00880.
- 30 Intl. Pat. Appl. Publ. No. PCT/US89/01025.
- Intl. Pat. Appl. Publ. No. WO 88/09812.

- Intl. Pat. Appl. Publ. No. WO 88/10315.
Intl. Pat. Appl. Publ. No. WO 89/06700.
Intl. Pat. Appl. Publ. No. WO 93/07278.
Abbott, "A method for computing the effectiveness of an insecticide," *J. Econ. Entomol.*, 18:265-267, 1925.
- 5 Abdullah *et al.*, *Biotechnology*, 4:1087, 1986.
Almond and Dean, *Biochemistry*, 32:1040-1046, 1993.
An *et al.*, *EMBO J.*, 4:277-287, 1985.
Angsuthanasamvat *et al.*, *FEMS Microbiol. Lett.*, 111:255-262, 1993.
- 10 Armstrong *et al.*, *Plant Cell Rep.*, 9:335-339, 1990.
Aronson, Wu, Zhang, "Mutagenesis of specificity and toxicity regions of a *Bacillus thuringiensis* protoxin gene." *J. Bacteriol.*, 177:4059-4065, 1995.
Bagdasarian *et al.*, *Gene*, 16:237, 1981.
Baum *et al.*, *Appl. Environ. Microbiol.*, 56:3420-3428, 1990.
- 15 Baum, "Tn5401, a new class II transposable element from *Bacillus thuringiensis*," *J. Bacteriol.*, 176:2835-2845, 1994.
Baum, *J. Bacteriol.*, 177:4036-4042, 1995.
Baum, Kakefuda, Gawron-Burke, "Engineering *Bacillus thuringiensis* Bioinsecticides with an Indigenous Site-Specific Recombination System," *Appl. Environ. Microbiol.*, 62:XXX-XXX, 1996.
- 20 Benbrook *et al.*, *In: Proceedings Bio Expo 1986*, Butterworth, Stoneham, MA, pp. 27-54, 1986.
Bevan *et al.*, *Nature*, 304:184, 1983.
Bolivar *et al.*, *Gene*, 2:95, 1977.
- 25 Branden and Tooze, "Introduction to Protein Structure," Garland Publishing, Inc., New York, NY, 1991.
Brussock and Currier, "Use of sodium dodecyl sulfate-polacryamide gel electrophoresis to quantify *Bacillus thuringiensis* δ -endotoxins," *In: "Analytical Chemistry of Bacillus thuringiensis,"* L.A. Hickie and W.L. Fitch, (Eds), American
30 Chemical Society, Washington D. C., pp. 78-87, 1990.

- Capecchi, "High efficiency transformation by direct microinjection of DNA into cultured mammalian cells," *Cell*, 22(2):479-488, 1980.
- Caramori, Albertini, Galizzi, "In vivo generation of hybrids between two *Bacillus thuringiensis* insect-toxin-encoding genes," *Gene*, 98:37-44, 1991.
- 5 Cashmore *et al.*, *Gen. Eng. of Plants*, Plenum Press, New York, 29-38, 1983.
- Chambers *et al.*, *Appl. Environ. Microbiol.*, 173:3966-3976, 1991.
- Chau *et al.*, *Science*, 244:174-181, 1989.
- Chen *et al.*, *Nucl. Acids Res.*, 20:4581-9, 1992.
- Chen, Curtiss, Alcantara, Dean, "Mutations in domain I of *Bacillus thuringiensis* δ -endotoxin CryIAb reduce the irreversible binding of toxin to *Manduca sexta* brush border membrane vesicles," *J. Biol. Chem.*, 270:6412-6419, 1995.
- 10 Chen, Lee, Dean, "Site-directed mutations in a highly conserved region of *Bacillus thuringiensis* δ -endotoxin affect inhibition of short circuit current across *Bombyx mori* midguts," *Proc. Natl. Acad. Sci. USA*, 90:9041-9045, 1993.
- 15 Chowrira and Burke, *Nucl. Acids Res.*, 20:2835-2840, 1992.
- Clapp, "Somatic gene therapy into hematopoietic cells. Current status and future implications," *Clin. Perinatol.*, 20(1):155-168, 1993.
- Cody, Luft, Jensen, Pangborn English, "Purification and crystallization of insecticidal δ -endotoxin CryIIIB2 from *Bacillus thuringiensis*," *Proteins: Struct. Funct. Genet.*, 14:324, 1992.
- 20 Collins and Olive, *Biochem.*, 32:2795-2799, 1993.
- Conway and Wickens, *In: RNA Processing*, p. 40, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1988.
- Cornelissen *et al.*, "A tobacco mosaic virus-induced tobacco protein is homologous to the sweet-tasting protein thaumatin," *Nature*, 321(6069):531-532, 1986.
- 25 Cramer, Cohen, Merrill, Song, "Structure and dynamics of the colicin E1 channel," *Molec. Microbiol.*, 4:519-526, 1990.
- CRC Handbook of Chemistry and Physics, 58th edition, CRC Press, Inc., Cleveland, Ohio, p. C-769, 1977.
- 30 Cristou *et al.*, *Plant Physiol.*, 87:671-674, 1988.

- Curiel, Agarwal, Wagner, Cotten, "Adenovirus enhancement of transferrin-polylysine-mediated gene delivery," *Proc. Natl. Acad. Sci. USA*, 88(19):8850-8854, 1991.
- Curiel, Wagner, Cotten, Birnstiel, Agarwal, Li, Loechel, Hu, "High-efficiency gene transfer mediated by adenovirus coupled to DNA-polylysine complexes," *Hum. Gen. Ther.*, 3(2):147-154, 1992.
- Daum, "Revision of two computer programs for probit analysis," *Bull. Entomol. Soc. Amer.*, 16:10-15, 1970.
- De Maagd, Kwa, van der Klei, Yamamoto, Schipper, Vlak, Stiekema, Bosch, "Domain III substitution in *Bacillus thuringiensis* delta-endotoxin CryIA(b) results in superior toxicity for *Spodoptera exigua* and altered membrane protein recognition," *Appl. Environ. Microbiol.*, 62:1537-1543, 1996.
- Dean *et al.*, *Nucl. Acids Res.*, 14(5):2229, 1986.
- Dhir *et al.*, *Plant Cell Reports*, 10:97, 1991.
- Diehn *et al.*, *Genet. Engineer.*, 18:83-99, 1996.
- Donovan, Dankocsik, Gilbert, Groat, Gawron-Burke, Carlton, "The P2 protein of *Bacillus thuringiensis* var. *kurstaki*: nucleotide sequence and entomocidal activity," *J. Biol. Chem.*, 263:561-567, 1988.
- Doyle *et al.*, *J. Biol. Chem.*, 261(20):9228-9236, 1986.
- Dropulic *et al.*, *J. Virol.*, 66:1432-41, 1992.
- Dunitz, "The entropic cost of bound water in crystals and biomolecules," *Science*, 264:670-68x, 1994.
- Earp and Ellar, *Nucl. Acids Res.*, 15:3619, 1987.
- Eglitis and Anderson, "Retroviral vectors for introduction of genes into mammalian cells," *Biotechniques*, 6(7):608-614, 1988.
- Eglitis, Kantoff, Kohn, Karson, Moen, Lothrop, Blaese, Anderson, "Retroviral-mediated gene transfer into hemopoietic cells," *Adv. Exp. Med. Biol.*, 241:19-27, 1988.
- Elroy-Stein and Moss, *Proc. Natl. Acad. Sci. USA*, 87:6743-7, 1990.
- English and Slatin, *Insect Biochem. Mol. Biol.*, 22:1-7, 1992.

- English, Readdy, Bastian, "Delta-endotoxin-induced leakage of $^{86}\text{Rb}^+ - \text{K}^+$ and H_2O from phospholipid vesicles is catalyzed by reconstituted midgut membrane," *Insect Biochem.*, 21:177-184, 1991.
- Fischhoff *et al.*, *Bio/Technology*, 5:807-813, 1987.
- 5 Fraley *et al.*, *Bio/Technology*, 3:629-635, 1985.
- Fraley *et al.*, *Proc. Natl. Acad. Sci. USA*, 80:4803, 1983.
- Frohman, PCR™ Protocols, a Guide to Methods and Applications XVIII Ed., Academic Press, New York, 1990.
- Fromm *et al.*, *Bio/Technology*, 8:833-839, 1990.
- 10 Fromm *et al.*, *Nature*, 319:791-793, 1986.
- Fromm, Taylor, Walbot, "Expression of genes transferred into monocot and dicot plant cells by electroporation," *Proc. Natl. Acad. Sci. USA*, 82(17):5824-5828, 1985.
- Fujimura *et al.*, *Plant Tissue Cult. Lett.*, 2:74, 1985.
- 15 Fynan, Webster, Fuller, Haynes, Santoro, Robinson, "DNA vaccines: protective immunizations by parenteral, mucosal, and gene gun inoculations," *Proc. Natl. Acad. Sci. USA*, 90(24):11478-11482, 1993.
- Galitsky, Cody, Wojtczak, Ghosh, Luft, Pangborn, Wawrzak, English, "Crystal and Molecular Structure of the Insecticidal Bacterial δ -Endotoxin CryIIIB2 of
- 20 *Bacillus thuringiensis*," Research Communication to Ecogen Inc., Langhorne, PA, 1993.
- Gao and Huang, *Nucl. Acids Res.*, 21:2867-72, 1993.
- Gazit and Shai, "Structural and Functional Characterization of the α -5 segment of *Bacillus thuringiensis* δ -endotoxin," *Biochemistry*, 32:3429-3436, 1993.
- 25 Gazit and Shai, "The assembly and organization of the α 5 and α 7 helices from the pore-forming domain of *Bacillus thuringiensis* δ -endotoxin," *J. Biol. Chem.*, 270:2571-2578, 1995.
- Ge, Rivers, Milne, Dean, "Functional domains of *Bacillus thuringiensis* insecticidal crystal proteins: refinement of *Heliothis virescens* and *Trichoplusia ni* specificity domains on CryIA(c)," *J. Biol. Chem.*, 266:17954-17958, 1991.
- 30

- Genovese and Milcarek, *In: RNA Processing*, p. 62, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1988.
- Gil and Proudfoot, *Nature*, 312:473, 1984.
- Gonzalez Jr. *et al.*, *Proc. Natl. Acad. Sci USA*, 79:6951-6955, 1982.
- 5 Graham and van der Eb, "Transformation of rat cells by DNA of human adenovirus 5," *Virology*, 54(2):536-539, 1973.
- Grochulski, Masson, Borisova, Pusztai-Carey, Schwartz, Brousseau, Cygler, "*Bacillus thuringiensis* CryIA(a) insecticidal toxin: crystal structure and channel formation," *J. Mol. Biol.*, 254:447-464, 1995.
- 10 Guerrier-Takada *et al.*, *Cell*, 35:849, 1983.
- Hampel and Tritz, *Biochem.*, 28:4929, 1989.
- Hampel *et al.*, *Nucl. Acids Res.*, 18:299, 1990.
- Harlow and Lane, "Antibodies: A Laboratory Manual," Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1988.
- 15 Herrera-Estrella *et al.*, *Nature*, 303:209, 1983.
- Hertel *et al.*, *Nucl. Acids Res.*, 20:3252, 1992.
- Hertig *et al.*, *Plant Mol. Biol.*, 16:171-174, 1991.
- Hess, *Intern Rev. Cytol.*, 107:367, 1987.
- Hills and Peters, *J. Econ. Entomol.*, 64:764-765, 1971.
- 20 Hockema, *In: The Binary Plant Vector System*, Offset-durkkerij, Kanters B.V., Alblasserdam, Chapter 5.
- Höfte and Whitely, *Microbiol. Rev.*, 53:242-255, 1989.
- Holland *et al.*, *Biochemistry*, 17:4900, 1978.
- Holsters *et al.*, *Mol. Gen. Genet.*, 163:181-187, 1978.
- 25 Honee, van der Salm, Visser, *Nucl. Acids Res.*, 16:6240, 1988.
- Horsch *et al.*, *Science*, 227:1229-1231, 1985.
- Humason, *In: Animal Tissue Techniques*, W.H. Freeman and Company, 1967.
- Jaeger *et al.*, *Proc. Natl. Acad. Sci. USA*, 86: 7706-7710, 1989.
- Johnston and Tang, "Gene gun transfection of animal cells and genetic immunization," *Methods Cell. Biol.*, 43(A):353-365, 1994.
- 30 Jorgensen *et al.*, *Mol. Gen. Genet.*, 207:471, 1987.

- Kaiser and Kezdy, *Science*, 223:249-255, 1984.
- Kashani-Saber *et al.*, *Antisense Res. Dev.*, 2:3-15, 1992.
- Keller *et al.*, *EMBO J.*, 8:1309-14, 1989.
- Klee *et al.*, *Bio/Technology*, 3:637-642, 1985.
- 5 Klein *et al.*, *Nature*, 327:70, 1987.
- Klein *et al.*, *Proc. Natl. Acad. Sci. USA*, 85:8502-8505, 1988.
- Kozak, *Nature*, 308:241-246, 1984.
- Krieg *et al.*, *Anzeiger fur Schadlingskunde Pflanzenschutz Umweltschutz*, 57:145-150, 1984.
- 10 Krieg *et al.*, *Z. ang Ent.*, 96:500-508, 1983.
- Kuby, *Immunology 2nd Edition*, W. H. Freeman & Company, NY, 1994
- Kunkle, "Rapid and efficient site-specific mutagenesis without phenotypic selection," *Proc. Natl. Acad. Sci. USA*, 82:488-492, 1985.
- Kunkle, Roberts, Zabour, *Methods Enzymol.*, 154:367-382, 1987.
- 15 Kwak, Lu, Dean, "Exploration of receptor binding of *Bacillus thuringiensis* toxins," *Mem. Inst. Oswaldo*, 90:75-79, 1995.
- Kwoh *et al.*, *Proc. Natl. Acad. Sci. USA*, 86(4):1173-1177, 1989.
- Kyte and Doolittle, *J. Mol. Biol.*, 157:105-132, 1982.
- L'Huillier *et al.*, *EMBO J.*, 11:4411-8, 1992.
- 20 LaBean and Kauffman, "Design of synthetic gene libraries encoding random sequence proteins with desired ensemble characteristics," *Prot. Sci.*, 2:1249-1254, 1993.
- Lambert, Buysse, Decock, Jansens, Piens, Saey, Seurinck, Van Audenhove, Van Rie, Van Vliet, Peferoen, "A *Bacillus thuringiensis* insecticidal crystal protein with a high activity against members of the family *Noctuidae*," *Appl. Environ. Microbiol.*, 62:80-86, 1996.
- 25 Lee, Milne, Ge, Dean, "Location of a *Bombyx mori* receptor binding region on a *Bacillus thuringiensis* δ -endotoxin," *J. Biol. Chem.*, 267:3115-3121, 1992.
- Lee, Young, Dean, "Domain III exchanges of *Bacillus thuringiensis* CryIA toxins affect binding to different gypsy moth midgut receptors," *Biochem. Biophys. Res. Commun.*, 216:306-312, 1995.
- 30

- Li, Carroll, Ellar, "Crystal structure of insecticidal δ -endotoxin from *Bacillus thuringiensis* at 2.5Å resolution," *Nature* (London), 353:815-821, 1991.
- Lieber *et al.*, *Methods Enzymol.*, 217:47-66, 1993.
- Lindstrom *et al.*, *Developmental Genetics*, 11:160, 1990.
- 5 Lisziewicz *et al.*, *Proc. Natl. Acad. Sci. U.S.A.*, 90:8000-4, 1993.
- Lorz *et al.*, *Mol. Gen. Genet.*, 199:178, 1985.
- Lu, Rajamohan, Dean, "Identification of amino acid residues of *Bacillus thuringiensis* δ -endotoxin CryIAa associated with membrane binding and toxicity to *Bombyx mori*," *J. Bacteriol.*, 176:5554-5559, 1994.
- 10 Lu, Xiao, Clapp, Li, Broxmeyer, "High efficiency retroviral mediated gene transduction into single isolated immature and replatable CD34(3+) hematopoietic stem/progenitor cells from human umbilical cord blood," *J. Exp. Med.*, 178(6):2089-2096, 1993.
- Macaluso and Mettus, *J. Bacteriol.*, 173:1353-1356, 1991.
- 15 Maddock *et al.*, *Third International Congress of Plant Molecular Biology*, Abstract 372, 1991.
- Maloy *et al.*, "Microbial Genetics" 2nd Edition. Jones and Bartlett Publishers, Boston, MA, 1994.
- Maloy, "Experimental Techniques in Bacterial Genetics" Jones and Bartlett Publishers, Boston, MA, 1990.
- 20 Maniatis, Fritsch, Sambrook, *In: Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1982.
- Marcotte *et al.*, *Nature*, 335:454, 1988.
- McDevitt *et al.*, *Cell*, 37:993-999, 1984.
- 25 McElroy *et al.*, *Plant Cell*, 2:163-171, 1990.
- Mettus and Macaluso, *Appl. Environ. Microbiol.*, 56:1128-1134, 1990.
- Michael, "Mutagenesis by Incorporation of a Phosphorylated Oligo During PCR™ Amplification," *BioTechniques*, 16(3):410-412, 1994.
- Neuhaus *et al.*, *Theor. Appl. Genet.*, 75:30, 1987.
- 30 Odell *et al.*, *Nature*, 313:810, 1985.
- Ohara *et al.*, *Proc. Natl. Acad. Sci. USA*, 86(15):5673-5677, 1989.

- Ohkawa *et al.*, *Nucl. Acids Symp. Ser.*, 27:15-6, 1992.
- Ojwang *et al.*, *Proc. Natl. Acad. Sci. USA*, 89:10802-6, 1992.
- Olson *et al.*, *J. Bacteriol.*, 150:6069, 1982.
- Omirulleh *et al.*, *Plant Molecular Biology*, 21:415-428, 1993.
- 5 Pandey and Marzluff, *In "RNA Processing,"* p. 133, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1987.
- Pena *et al.*, *Nature*, 325:274, 1987.
- Perrault *et al.*, *Nature*, 344:565, 1990.
- Perrotta and Been, *Biochem.*, 31:16, 1992.
- 10 Pieken *et al.*, *Science*, 253:314, 1991.
- Poszkowski *et al.*, *EMBO J.*, 3:2719, 1989.
- Potrykus *et al.*, *Mol. Gen. Genet.*, 199:183, 1985.
- Poulsen *et al.*, *Mol. Gen. Genet.*, 205:193-200, 1986.
- Prokop and Bajpai, "Recombinant DNA Technology I," *Ann. N. Y. Acad. Sci.*,
15 646:xxx-xxx, 1991.
- Rajamohan, Alcantara, Lee, Chen, Curtiss, Dean, "Single amino acid changes in domain II of *Bacillus thuringiensis* CryIAb δ -endotoxin affect irreversible binding to *Manduca sexta* midgut membrane vesicles," *J. Bacteriol.*, 177:2276-2282, 1995.
- 20 Rajamohan, Cotrill, Gould, Dean, "Role of domain II, loop 2 residues of *Bacillus thuringiensis* CryIAb δ -endotoxin in reversible and irreversible binding to *Manduca sexta* and *Heliothis virescens*," *J. Biol. Chem.*, 271:2390-2397, 1996.
- Rogers *et al.*, *In: Methods For Plant Molecular Biology*, A. Weissbach and H. Weissbach, eds., Academic Press Inc., San Diego, CA 1988.
- 25 Rogers *et al.*, *Methods Enzymol.*, 153:253-277, 1987.
- Rossi *et al.*, *Aids Res. Hum. Retrovir.*, 8:183, 1992.
- Sadofsky and Alwine, *Molec. Cell. Biol.*, 4(8):1460-1468, 1984.
- Sambrook *et al.*, "Molecular Cloning: A Laboratory Manual," Cold Spring Harbor
30 Laboratory, Cold Spring Harbor, NY, 1989.

- Sanchis, Lereclus, Menou, Chaufaux, Guo, Lecadet, *Mol. Microbiol.*, 3:229-238, 1989.
- Sanchis, Lereclus, Menou, Chaufaux, Lecadet, *Mol. Microbiol.*, 2:393-404, 1988.
- Sarver *et al.*, *Science*, 247:1222-1225, 1990.
- 5 Saville and Collins, *Cell*, 61:685-696, 1990.
- Saville and Collins, *Proc. Natl. Acad. Sci. USA*, 88:8826-8830, 1991.
- Scanlon *et al.*, *Proc. Natl. Acad. Sci. USA*, 88:10591-5, 1991.
- Scaringe *et al.*, *Nucl. Acids Res.*, 18:5433-5441, 1990.
- Schnepf and Whitely, *Proc. Natl. Acad. Sci. USA*, 78:2893-2897, 1981.
- 10 Schnepf *et al.*, *J. Biol. Chem.*, 260:6264-6272, 1985.
- Segal, "Biochemical Calculations" 2nd Edition, John Wiley & Sons, New York, 1976.
- Shaw and Kamen, *Cell*, 46:659-667, 1986.
- Shaw and Kamen, *In*: "RNA Processing", p. 220, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1987.
- 15 Simpson, *Science*, 233:34, 1986.
- Slaney, Robbins, English, "Mode of action of *Bacillus thuringiensis* toxin CryIIIA: An analysis of toxicity in *Leptinotarsa decemlineata* (Say) and *Diabrotica undecimpunctata howardi* Barber," *Insect Biochem. Molec. Biol.*, 22:9-18, 1992.
- Slatin, Abrams, English, "Delta-endotoxins form cation-selective channels in planar lipid bilayers," *Biochem. Biophys. Res. Comm.*, 169(2):765-772, 1990.
- 20 Smedley and Ellar, "Mutagenesis of three surface-exposed loops of a *Bacillus thuringiensis* insecticidal toxin reveals residues important for toxicity, receptor recognition and possibly membrane insertion," *Microbiology*, 142:1617-1624, 1996.
- 25 Smith and Ellar, "Mutagenesis of two surface-exposed loops of the *Bacillus thuringiensis* CryIC δ -endotoxin affects insecticidal specificity," *Biochem. J.*, 302:611-616, 1994.
- Smith, Merrick, Bone, Ellar, *Appl. Environ. Microbiol.*, 62:680-684, 1996.
- Spielmann *et al.*, *Mol. Gen. Genet.*, 205:34, 1986.

- Stemmer and Morris, "Enzymatic Inverse PCR™: A Restriction Site Independent, Single-Fragment Method for High-Efficiency, Site-Directed Mutagenesis," *BioTechniques*, 13(2):214-220, 1992.
- Stemmer, *Proc. Natl. Acad. Sci. USA*, 91:10747-1075, 1994.
- 5 Taira *et al.*, *Nucl. Acids Res.*, 19:5125-30, 1991.
- Tomic *et al.*, *Nucl. Acids Res.*, 12:1656, 1990.
- Tomic, Sunjevaric, Savtchenko, Blumenberg, "A rapid and simple method for introducing specific mutations into any position of DNA leaving all other positions unaltered," *Nucleic Acids Res.*, 18(6):1656, 1990.
- 10 Toriyama *et al.*, *Theor Appl. Genet.*, 73:16, 1986.
- Uchimiya *et al.*, *Mol. Gen. Genet.*, 204:204, 1986.
- Upender *et al.*, *Biotechniques*, 18:29-31, 1995.
- Usman and Cedergren, *TIBS*, 17:34, 1992.
- Usman and Cedergren, *Trends in Biochem. Sci.*, 17:334, 1992.
- 15 Usman *et al.*, *J. Am. Chem. Soc.*, 109:7845-7854, 1987.
- Vallette, Merge, Reiss, Adesnik, "Construction of mutant and chimeric genes using the polymerase chain reaction," *Nucl. Acids Res.*, 17:723-733, 1989.
- Vasil *et al.*, "Herbicide-resistant fertile transgenic wheat plants obtained by microprojectile bombardment of regenerable embryogenic callus," *Biotechnology*,
20 10:667-674, 1992.
- Vasil, *Biotechnology*, 6:397, 1988.
- Velten and Schell, *Nucl. Acids Res.*, 13:6981-6998, 1985.
- Velten *et al.*, *EMBO J.*, 3:2723-2730, 1984.
- Ventura *et al.*, *Nucl. Acids Res.*, 21:3249-55, 1993.
- 25 Vodkin *et al.*, *Cell*, 34:1023, 1983.
- Vogel *et al.*, *J. Cell Biochem.*, Suppl. 13D:312, 1989.
- Von Tersch, Slatin, Kulesza, English, "Membrane permeabilizing activity of *Bacillus thuringiensis* Coleopteran-active toxins CryIIIB2 and CryIIIB2 domain 1 peptides," *Appl. Env Microbiol.*, 60:3711-3717, 1994.
- 30 Wagner, Zatloukal, Cotten, Kirlappos, Mechtler, Curiel, Birnstiel, "Coupling of adenovirus to transferrin-polylysine/DNA complexes greatly enhances receptor-

- mediated gene delivery and expression of transfected genes," *Proc. Natl. Acad. Sci. USA*, 89(13):6099-6103, 1992.
- Walker *et al.*, *Proc. Natl. Acad. Sci. USA*, 89(1):392-396, 1992.
- Walters *et al.*, *Biochem. Biophys. Res. Commun.*, 196:921-926, 1993.
- 5 Watson *et al.*, *Molecular Biology of the Gene*, 4th Ed., W. A. Benjamin, Inc., Menlo Park, CA, 1987.
- Weerasinghe *et al.*, *J. Virol.*, 65:5531-4, 1991.
- Weissbach and Weissbach, *Methods for Plant Molecular Biology*, (eds.), Academic Press, Inc., San Diego, CA, 1988.
- 10 Wenzler *et al.*, *Plant Mol. Biol.*, 12:41-50, 1989.
- Wickens and Stephenson, *Science*, 226:1045, 1984.
- Wickens *et al.*, In: "RNA Processing," p. 9, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1987.
- Wolfersberger *et al.*, *Appl. Environ. Microbiol.*, 62:279-282, 1996.
- 15 Wong and Neumann, "Electric field mediated gene transfer," *Biochim. Biophys. Res. Commun.*, 107(2):584-587, 1982.
- Woolf *et al.*, *Proc. Natl. Acad. Sci. USA*, 89:7305-7309, 1992.
- Wu and Aronson, "Localized mutagenesis defines regions of the *Bacillus thuringiensis* δ -endotoxin involved in toxicity and specificity," *J. Biol. Chem.*, 267:2311-2317, 1992.
- 20 Wu and Dean, "Functional significance of loops in the receptor binding domain of *Bacillus thuringiensis* CryIIIA δ -endotoxin," *J. Mol. Biol.*, 255:628-640, 1996.
- Yamada *et al.*, *Plant Cell Rep.*, 4:85, 1986.
- Yang *et al.*, *Proc. Natl. Acad. Sci. USA*, 87:4144-48, 1990.
- 25 Yu *et al.*, *Proc. Natl. Acad. Sci. USA*, 90:6340-4, 1993.
- Zatloukal, Wagner, Cotten, Phillips, Plank, Steinlein, Curiel, Birnstiel, "Transferrin infection: a highly efficient way to express gene constructs in eukaryotic cells," *Ann. N. Y. Acad. Sci.*, 660:136-153, 1992.
- Zhang and Matthews, "Conservations of solvent-binding sites in 10 crystal forms of T4 lysozyme," *Prot. Sci.*, 3:1031-1039, 1994.
- 30 Zhou *et al.*, *Mol. Cell Biol.*, 10:4529-37, 1990.

All of the compositions and methods disclosed and claimed herein can be made and executed without undue experimentation in light of the present disclosure. While the compositions and methods of this invention have been described in terms of preferred embodiments, it will be apparent to those of skill in the art that variations may be applied to the compositions and methods and in the steps or in the sequence of steps of the method described herein without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain agents which are both chemically and physiologically related may be substituted for the agents described herein while the same or similar results would be achieved. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention as defined by the appended claims.

CLAIMS:

1. An isolated *B. thuringiensis* Cry3Bb polypeptide modified to have improved insecticidal activity or enhanced insecticidal specificity against a target insect,
5 said polypeptide comprising at least one amino acid substitution, one amino acid addition, or one amino acid deletion in the primary sequence of the native or unmodified Cry3Bb polypeptide, wherein said substitution or deletion occurs at a position corresponding to from about amino acid 1 to about amino acid 365 of the unmodified polypeptide's amino acid sequence.
10
2. The polypeptide of claim 1, wherein Asp103 is replaced by glutamic acid; Ala104 is deleted; Thr154 is replaced by glycine or phenylalanine; Pro155 is replaced by histidine; Leu156 is replaced by histidine; Leu158 is replaced by
15 arginine; Ser160 is replaced by asparagine; Lys161 is replaced by proline; Pro162 is replaced by histidine; Asp165 is replaced by glycine; Lys189 is replaced by glycine; Ser223 is replaced by proline; Tyr230 is replaced by leucine or serine; His231 is replaced by arginine, asparagine, serine, or threonine; Thr241 is replaced by serine; Tyr287 is replaced by phenylalanine; Asp288 is replaced by asparagine; Ile289 is replaced by threonine or valine; Arg290 is
20 replaced by asparagine, leucine or valine; Leu291 is replaced by arginine; Tyr292 is replaced by phenylalanine; Ser293 is replaced by arginine or proline; Phe305 is replaced by serine; Ser311 is replaced by alanine, isoleucine, leucine,

or threonine; Leu312 is replaced by proline or valine; Asn313 is replaced by arginine, histidine, threonine or valine; Thr314 is replaced by asparagine; Leu315 is replaced by proline; Gln316 is replaced by aspartic acid, leucine, methionine, or tryptophan; Glu317 is replaced by alanine, asparagine, lysine or valine; Tyr318 is replaced by cysteine; Gln348 is replaced by arginine; or Val365 is replaced by alanine.

3. The polypeptide of claim 1 or 2, wherein Thr154 is replaced by phenylalanine, Pro155 is replaced by histidine, Leu156 is replaced by histidine, and Leu158 is replaced by arginine.

4. The polypeptide of claim 1 or 2, wherein Tyr230 is replaced by leucine, and His231 is replaced by serine.

5. The polypeptide of claim 1 or 2, wherein Ser223 is replaced by proline, and Tyr230 is replaced by serine.

20

6. The polypeptide of claim 1 or 2, wherein His231 is replaced by arginine.

7. The polypeptide of claim 1 or 2, wherein His231 is replaced by asparagine, and Thr241 is replaced by serine.

5

8. The polypeptide of claim 1 or 2, wherein His231 is replaced by threonine.

9. The polypeptide of claim 1 or 2, wherein Arg290 is replaced by asparagine.

10

10. The polypeptide of claim 1 or 2, wherein Ser311 is replaced by leucine, Asn313 is replaced by threonine, and Glu317 is replaced by lysine.

15

11. The polypeptide of claim 1 or 2, wherein Ser311 is replaced by threonine, Glu317 is replaced by lysine, and Tyr318 is replaced by cysteine.

- 20 12. The polypeptide of claim 1 or 2, wherein Ser311 is replaced by alanine, Leu312 is replaced by valine, and Gln316 is replaced by tryptophan.

13. The polypeptide of claim 1 or 2, wherein His231 is replaced by arginine, Ser311 is replaced by leucine, Asn313 is replaced by threonine, and Glu317 is replaced by lysine.
- 5
14. The polypeptide of claim 1 or 2, wherein Ser311 is replaced by threonine, Leu312 is replaced by proline, Asn313 is replaced by threonine, and Glu317 is replaced by asparagine.
- 10
15. The polypeptide of claim 1 or 2, wherein Ser311 is replaced by alanine, and Gln316 is replaced by aspartic acid.
- 15 16. The polypeptide of claim 1 or 2, wherein Ile289 is replaced by threonine, Leu291 is replaced by arginine, Tyr292 is replaced by phenylalanine, and Ser293 is replaced by arginine.
- 20 17. The polypeptide of claim 1 or 2, wherein His231 is replaced by arginine, and Ser311 is replaced by leucine.

18. The polypeptide of claim 1 or 2, wherein Ser311 is replaced by isoleucine.
19. The polypeptide of claim 1 or 2, wherein Ser311 is replaced by isoleucine, and
5 Asn313 is replaced by histidine.
20. The polypeptide of claim 1 or 2, wherein Asn313 is replaced by valine, Thr314
is replaced by asparagine, Gln316 is replaced by methionine, and Glu317 is
10 replaced by valine.
21. The polypeptide of claim 1 or 2, wherein Asn313 is replaced by arginine,
Leu315 is replaced by proline, Gln316 is replaced by leucine, and Glu317 is
15 replaced by alanine.
22. The polypeptide of claim 1 or 2, wherein Tyr287 is replaced by phenylalanine,
Asp288 is replaced by asparagine, and Arg290 is replaced by leucine.
20
23. The polypeptide of claim 1 or 2, wherein Arg290 is replaced by valine.

24. The polypeptide of claim 1 or 2, wherein Asp165 is replaced by glycine.
- 5 25. The polypeptide of claim 1 or 2, wherein Ser160 is replaced by asparagine, Lys161 is replaced by proline, Pro162 is replaced by histidine, and Thr154 is replaced by glycine.
- 10 26. The polypeptide of claim 1 or 2, wherein Ile289 is replaced by valine, and Ser293 is replaced by proline.
- 15 27. The polypeptide of claim 1 or 2, wherein Ser160 is replaced by asparagine, Lys161 is replaced by proline, Pro162 is replaced by histidine, Asp165 is replaced by glycine, Ile289 is replaced by valine, and Ser293 is replaced by proline.
- 20 28. The polypeptide of claim 1 or 2, wherein Asp103 is replaced by glutamic acid, and Ala104 is deleted.

29. The polypeptide of claim 1 or 2, wherein Lys189 is replaced by glycine.
30. The polypeptide of claim 1 or 2, wherein Asp103 is replaced by glutamic acid,
5 Ala104 is deleted, Ser160 is replaced by asparagine, Lys161 is replaced by proline, Pro162 is replaced by histidine, and Asp165 is replaced by glycine.
31. The polypeptide of claim 1 or 2, wherein Asp103 is replaced by glutamic acid,
10 Ala104 is deleted, Thr154 is replaced by phenylalanine, Pro155 is replaced by histidine, Leu156 is replaced by histidine, and Leu158 is replaced by arginine.
32. The polypeptide of claim 1 or 2, wherein Asp165 is replaced by glycine, Ser311
15 is replaced by threonine, and Glu317 is replaced by lysine.
33. The polypeptide of claim 1 or 2, wherein Asp165 is replaced by glycine, Ile289
is replaced by valine, Ser293 is replaced by proline, Phe305 is replaced by
20 serine, Ser311 is replaced by alanine, Leu312 is replaced by valine, Gln316 is replaced by tryptophan, Gln348 is replaced by arginine, and Val365 is replaced by alanine.

34. The polypeptide of claim 1 or 2, wherein Ile289 is replaced by valine, Ser293 is replaced by proline, and Gln348 is replaced by arginine.

5

35. The polypeptide of claim 1 or 2, wherein Asp165 is replaced by glycine, and Ser311 is replaced by leucine.

- 10 36. The polypeptide of claim 1 or 2, wherein the first 159 amino acids are deleted.

37. The polypeptide of claim 1 or 2, wherein Gln348 is replaced by arginine.

15

38. The polypeptide of claim 1 or 2, wherein Asp165 is replaced by glycine, His231 is replaced by arginine, Ser311 is replaced by leucine, Asn313 is replaced by threonine, and Glu317 is replaced by lysine.

20

39. The polypeptide of any preceding claim, wherein said polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ

ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22,
SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID
NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ
ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50,
5 SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID
NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ
ID NO:70, SEQ ID NO:100 and SEQ ID NO:108.

10 40. The polypeptide of any preceding claim, wherein said polypeptide is encoded by
a contiguous nucleic acid sequence selected from the group consisting of SEQ
ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID
NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ
ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29,
15 SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID
NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ
ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57,
SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID
NO:67, SEQ ID NO:69, SEQ ID NO:99 and SEQ ID NO:107.

20

41. A composition comprising an insecticidally-effective amount of the Cry3Bb
polypeptide of claim 1.

42. The composition of claim 41, comprising from about 0.5% to about 99% by weight of the polypeptide of claim 1.

5

43. The composition of claim 41 or 42, wherein said polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:72, SEQ ID NO:100, and SEQ ID NO:108.

44. The composition of any one of claims 41 to 43, wherein said polypeptide is encoded by a nucleic acid sequence having the sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ

ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39,
SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID
NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ
ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67,
5 SEQ ID NO:69, SEQ ID NO:71, SEQ ID NO:99, or SEQ ID NO:107.

45. The composition of any one of claims 41 to 44, prepared by a process comprising the steps of:
- 10
- (a) culturing a *Bacillus thuringiensis* NRRL B-21744, NRRL B-21745, NRRL B-21746, NRRL B-21747, NRRL B-21748, NRRL B-21749, NRRL B-21750, NRRL B-21751, NRRL B-21752, NRRL B-21753, NRRL B-21754, NRRL B-21755, NRRL B-21756, NRRL B-21757, NRRL B-21758, NRRL B-21759, NRRL B-21760, NRRL B-21761, NRRL B-21762, NRRL B-21763, NRRL B-21764, NRRL B-21765, NRRL B-21766, NRRL B-21767, NRRL B-21768, NRRL B-21769, NRRL B-21770, NRRL B-21771, NRRL B-21772, NRRL B-21773, NRRL B-21774, NRRL B-21775, NRRL B-21776, NRRL B-21777, NRRL B-21778, NRRL B-21779, or EG11098 cell under conditions
15 effective to produce an insecticidal polypeptide; and
- 20

(b) obtaining said insecticidal polypeptide from said cell.

46. The composition of any one of claims 41 to 45, comprising a *Bacillus thuringiensis* NRRL B-21744, NRRL B-21745, NRRL B-21746, NRRL B-21747, NRRL B-21748, NRRL B-21749, NRRL B-21750, NRRL B-21751, NRRL B-21752, NRRL B-21753, NRRL B-21754, NRRL B-21755, NRRL B-21756, NRRL B-21757, NRRL B-21758, NRRL B-21759, NRRL B-21760, NRRL B-21761, NRRL B-21762, NRRL B-21763, NRRL B-21764, NRRL B-21765, NRRL B-21766, NRRL B-21767, NRRL B-21768, NRRL B-21769, NRRL B-21770, NRRL B-21771, NRRL B-21772, NRRL B-21773, NRRL B-21774, NRRL B-21775, NRRL B-21776, NRRL B-21777, NRRL B-21778, or NRRL B-21779 cell.
47. The composition of any one of claims 41 to 46, wherein said composition comprises a cell extract, cell suspension, protein fraction, crystal fraction, cell culture, cell homogenate, cell lysate, cell supernatant, cell filtrate, or cell pellet of a *Bacillus thuringiensis* NRRL B-21744, NRRL B-21745, NRRL B-21746, NRRL B-21747, NRRL B-21748, NRRL B-21749, NRRL B-21750, NRRL B-21751, NRRL B-21752, NRRL B-21753, NRRL B-21754, NRRL B-21755, NRRL B-21756, NRRL B-21757, NRRL B-21758, NRRL B-21759, NRRL B-21760, NRRL B-21761, NRRL B-21762, NRRL B-21763, NRRL B-21764, NRRL B-21765, NRRL B-21766, NRRL B-21767, NRRL B-21768, NRRL B-21769, NRRL B-21770, NRRL B-21771, NRRL B-21772, NRRL B-21773,

NRRL B-21774, NRRL B-21775, NRRL B-21776, NRRL B-21777, NRRL B-21778, NRRL B-21779, or an EG11098 cell.

- 5 48. The composition of any one of claims 41 to 47, formulated as a powder, granule, spray, emulsion, colloid, or solution.
49. The composition of any one of claims 41 to 48, wherein said composition is
10 prepared by desiccation, lyophilization, homogenization, freeze drying, emulsification, evaporation, separation, extraction, filtration, centrifugation, sedimentation, dilution, crystallization, or concentration.
- 15 50. A polynucleotide comprising an isolated sequence region that encodes the polypeptide of any one of claims 1 to 40.
51. The polynucleotide of claim 50, comprising an isolated sequence region that
20 encodes a polypeptide that comprises an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26,

SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:72, SEQ ID NO:100, and SEQ ID NO:108.

52. The polynucleotide of claim 50 or 51, comprising a contiguous nucleotide sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65, SEQ ID NO:67, SEQ ID NO:69, SEQ ID NO:99, SEQ ID NO:99, and SEQ ID NO:107.

20

53. The polynucleotide of any one of claims 50 to 53, characterized as DNA, cDNA, rRNA, or mRNA.

54. The polynucleotide of any one of claims 50 to 53, wherein said polynucleotide is from about 2000 to about 10,000 nucleotides in length.

5

55. The polynucleotide of any one of claims 50 to 54, wherein said nucleic acid segment is from about 3000 to about 8,000 nucleotides in length.

- 10 56. The polynucleotide of any one of claims 50 to 55, wherein said isolated sequence region is operably linked to a promoter, said promoter expressing said sequence region.

- 15 57. The polynucleotide of any one of claims 50 to 56, wherein said isolated sequence region is operably linked to a heterologous promoter.

- 20 58. The polynucleotide of any one of claims 50 to 57, wherein said isolated sequence region is operably linked to a plant-expressible promoter.

59. The polynucleotide of any one of claims 50 to 58, wherein said isolated sequence region is operably linked to a constitutive, inducible, or tissue-specific promoter.

5

60. A vector comprising the polynucleotide of any one of claims 50 to 59, or a polynucleotide that encodes the polypeptide of any one of claims 1 to 40.

10 61. The vector of claim 60, defined as a plasmid, a cosmid, a phagemid, a phage, a virus, or a baculovirus.

62. The vector of claim 60 or 61, transformed and replicated in a prokaryotic or
15 eukaryotic host.

63. A virus comprising the polypeptide of any one of claims 1 to 40, or the polynucleotide of any one of claims 50 to 59.

20

64. A transformed host cell comprising the polypeptide of any one of claims 1 to 40, the polynucleotide of any one of claims 50 to 59, the vector of any one of claims 60 to 62, or the virus of claim 63.

5

65. The transformed host cell of claim 64, further defined as a prokaryotic or eukaryotic cell.

- 10 66. The transformed host cell of claim 64 or 65, wherein said prokaryotic cell is a eubacterial, archaeobacterial or cyanobacterial cell, or wherein said eukaryotic cell is an animal, fungal, or plant cell.

- 15 67. The transformed host cell of any one of claims 64 to 66, wherein said cell is an *E. coli*, *B. thuringiensis*, *A. tumefaciens*, *B. subtilis*, *B. megaterium*, *B. cereus*, *Salmonella* spp., or *Pseudomonas* spp. cell.

- 20 68. The transformed host cell of any one of claims 64 to 67, wherein said cell is selected from the group consisting of *B. thuringiensis* NRRL B-21744, NRRL B-21745, NRRL B-21746, NRRL B-21747, NRRL B-21748, NRRL B-21749, NRRL B-21750, NRRL B-21751, NRRL B-21752, NRRL B-21753, NRRL B-

21754, NRRL B-21755, NRRL B-21756, NRRL B-21757, NRRL B-21758,
NRRL B-21759, NRRL B-21760, NRRL B-21761, NRRL B-21762, NRRL B-
21763, NRRL B-21764, NRRL B-21765, NRRL B-21766, NRRL B-21767,
NRRL B-21768, NRRL B-21769, NRRL B-21770, NRRL B-21771, NRRL B-
5 21772, NRRL B-21773, NRRL B-21774, NRRL B-21775, NRRL B-21776,
NRRL B-21777, NRRL B-21778, and NRRL B-21779.

69. The transformed host cell of claim 66, wherein said plant cell is a grain, tree,
10 legume, fiber, vegetable, fruit, berry, nut, citrus, grass, cactus, succulent, or
ornamental plant cell.

70. The transformed host cell of claim 69, wherein said plant cell is a corn, rice,
15 tobacco, alfalfa, soybean, sorghum, potato, tomato, flax, canola, sunflower,
cotton, flax, kapok, wheat, oat, barley, or rye cell.

71. The transformed host cell of any one of claims 64 to 70, wherein said
20 polynucleotide is introduced into said cell by a vector, virus, cosmid, phagemid,
phage, plasmid, or by electroporation, transformation, conjugation,
microprojectile bombardment, direct DNA injection, naked DNA transfer,
transformation, or transfection.

72. A transgenic plant comprising the polypeptide of any one of claims 1 to 40,
the polynucleotide of any one of claims 50 to 59, the vector of any one of
5 claims 60 to 62, the virus of claim 63, or the host cell of any one of claims 64
to 71.
- 73 The transgenic plant of claim 72, having incorporated into its genome a selected
10 polynucleotide that encodes the polypeptide of any one of claims 1 to 40.
74. The transgenic plant of claim 72 or 73, wherein said polypeptide comprises an
amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ
15 ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ
ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22,
SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID
NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ
ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50,
20 SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID
NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ
ID NO:70, SEQ ID NO:100, and SEQ ID NO:108.

75. The transgenic plant of any one of claims 72 to 74, wherein said plant is a grain, tree, legume, fiber, vegetable, fruit, berry, nut, citrus, grass, cactus, succulent, or ornamental plant.

5

76. The transgenic plant of any one of claims 62 to 75, wherein said plant is a corn, rice, tobacco, alfalfa, soybean, sorghum, potato, tomato, flax, canola, sunflower, cotton, flax, kapok, wheat, oat, barley, or rye plant.

10

77. A progeny of any generation of the transgenic plant of any one of claims 72 to 76.

15

78. A seed of any generation of the transgenic plant of any one of claims 72 to 76.

79. A seed of any generation of the progeny of claim 77.

20

80. A plant grown from the seed of claim 78 or or 79.

81. A method of killing a coleopteran insect, said method comprising the step of contacting said insect with an insecticidally-effective amount of the polypeptide of claim 1.

5

82. A method of controlling a coleopteran insect population, said method comprising the step of providing to the environment of said insect population, an insecticidally-effective amount of the polypeptide of claim 1.

10

83. The method of claim 81 or 82, wherein said polypeptide is obtained from a cell extract, cell suspension, protein fraction, crystal fraction, cell culture, cell homogenate, cell lysate, cell supernatant, cell filtrate, or cell pellet of a *Bacillus thuringiensis* NRRL B-21744, NRRL B-21745, NRRL B-21746, NRRL B-21747, NRRL B-21748, NRRL B-21749, NRRL B-21750, NRRL B-21751, NRRL B-21752, NRRL B-21753, NRRL B-21754, NRRL B-21755, NRRL B-21756, NRRL B-21757, NRRL B-21758, NRRL B-21759, NRRL B-21760, NRRL B-21761, NRRL B-21762, NRRL B-21763, NRRL B-21764, NRRL B-21765, NRRL B-21766, NRRL B-21767, NRRL B-21768, NRRL B-21769, NRRL B-21770, NRRL B-21771, NRRL B-21772, NRRL B-21773, NRRL B-21774, NRRL B-21775, NRRL B-21776, NRRL B-21777, NRRL B-21778, or NRRL B-21779 cell.

84. The method of any one of claims 81 to 83, wherein said polypeptide is provided to said environment by spraying, dusting, sprinkling, soaking, aerating,
5 misting, atomizing, soil injection, soil tilling, seed coating, or seedling coating.

85. The method of any one of claims 81 to 84, wherein said polypeptide is formulated as a powder, granule, spray, emulsion, colloid, or solution.
10

86. The method of any one of claims 81 to 85, wherein said polypeptide is prepared by desiccation, lyophilization, homogenization, freeze drying, emulsification, evaporation, separation, extraction, filtration, centrifugation, sedimentation, dilution, crystallization, or concentration.
15

87. A method of preparing a Coleopteran-resistant transgenic plant, comprising the steps of:

20

(a) transforming a plant cell with a polynucleotide comprising a selected sequence region that encodes the polypeptide of claim 1, wherein said sequence

region is operably linked to a promoter which expresses said sequence region;
and

- 5 (b) generating from said plant cell a transgenic plant that comprises said selected sequence region and that expresses said polypeptide.

88. The method of claim 87, wherein said sequence region encodes a polypeptide
10 that comprises an amino acid sequence selected from the group consisting of
SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10,
SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID
NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ
ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38,
15 SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID
NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ
ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66,
SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, and SEQ ID NO:108.

20

89. A method of killing a Coleopteran insect, comprising feeding to said insect a
plant cell transformed with a polynucleotide that encodes an amino acid
sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4,

SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14,
SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID
NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ
ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42,
5 SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID
NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ
ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70,
SEQ ID NO:100, and SEQ ID NO:108.

10

90. The method of claim 89, wherein said insect is killed by ingesting a portion of a transgenic plant that comprises said transformed cell.

15 91. A method of preparing a plant seed resistant to Coleopteran insect attack, said method comprising the steps of:

(a) transforming a plant cell with a nucleic acid segment comprising a polynucleotide that encodes an amino acid sequence selected from the group
20 consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ
ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18,
SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID
NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ

ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:100, and SEQ ID NO:108 to produce a transformed plant cell;

- (b) growing said transformed plant cell under conditions effective to produce a transgenic plant from said cell; and
- (c) obtaining from said transgenic plant, a seed resistant to attack by said Coleopteran insect.

92. The method of claim 91, wherein step (a) comprises transforming said plant cell by electroporation, transfection, naked DNA uptake, protoplast generation, direct transfer of DNA into pollen, embryo or pluripotent plant cell, *Agrobacterium*-mediated transformation, particle bombardment, or microprojectile bombardment.

20

93. The method of claim 91 or 92, wherein step (b) comprises generation of pluripotent plant cells from said transformed plant cell.

94. A method for producing a modified Cry3Bb polypeptide having improved insecticidal activity or specificity, comprising:

5 (a) obtaining a high-resolution three-dimensional crystal structure of said polypeptide;

(b) locating in said crystal structure of said polypeptide one or more regions of bound water, wherein said bound water forms a contiguous hydrated
10 surfaces separated by no more than about 16Å;

(c) increasing the hydrophobicity of one or more amino acids of said polypeptide in said region; and

15 (d) obtaining the modified Cry3Bb polypeptide so produced.

95. A method for producing a modified Cry3Bb polypeptide having improved insecticidal activity, or enhanced insecticidal specificity, comprising

20

(a) obtaining a high-resolution three-dimensional crystal structure of said polypeptide;

- (b) identifying a loop region in said polypeptide;
- (c) modifying one or more amino acids in said loop region to increase the hydrophobicity of one or more of said amino acids; and
- (d) obtaining the modified Cry3Bb polypeptide so produced.

- 5
96. A method for increasing the mobility of channel forming helices of a Cry3Bb polypeptide, comprising disrupting one or more hydrogen bonds formed between a first amino acid of one or more of said channel forming helices and a second amino acid of said polypeptide.
- 10
97. The method of claim 96, wherein said hydrogen bonds are formed inter- or intramolecularly.
- 15
98. The method of claim 96, wherein said disrupting comprises replacing said first amino acid or said second amino acid with a third amino acid whose spatial distance bond angle is greater than about 3Å, or whose spatial orientation is not equal to 180 ± 60 degrees relative to the hydrogen bonding site of said first or said second amino acid.
- 20

99. A method for increasing the flexibility of a loop region in a channel forming domain of a Cry3Bb polypeptide, comprising:

5

(a) obtaining a crystal structure of a Cry3Bb polypeptide having one or more loop regions between adjacent α -helices;

(b) identifying the amino acids comprising said loop region; and

10

(c) altering one or more of said amino acids in said loop region to reduce the steric hindrance in said region, wherein said altering increases flexibility of said loop region in said polypeptide.

15

100. A method of increasing the insecticidal activity of a Cry3Bb polypeptide, comprising reducing or eliminating binding of said polypeptide to a carbohydrate in a target insect gut.

20

101. The method of claim 100, wherein said reducing or eliminating is accomplished by removal of one or more α -helices of domain 1 of said polypeptide.

102. The method claim 100, wherein said reducing or eliminating is accomplished by removal of α -helices $\alpha 1$, $\alpha 2a/b$, or $\alpha 3$.

5

103. The method of claim 102, wherein said reducing or eliminating is accomplished by replacing one or more amino acids within loop $\beta 1, \alpha 8$, with one or more amino acids having increased hydrophobicity.

10

104. The method of claim 103, wherein said reducing or eliminating is accomplished by replacing with any other amino acid, one or more amino acids selected from the group consisting of threonine 154, proline 155, leucine 156, and leucine 158.

15

105. A method of preparing a modified Cry3Bb polypeptide having improved insecticidal activity or enhanced insecticidal specificity when compared to an unmodified Cry3Bb polypeptide, said method comprising the steps of:

20

(a) obtaining a crystal structure of said polypeptide;

(b) identifying from said crystal structure one or more surface-exposed amino acids in said polypeptide;

(c) randomly substituting one or more of said surface-exposed amino acids to obtain a plurality of mutated polypeptides, wherein at least 50% of said mutated polypeptides have diminished insecticidal activity, or reduced insecticidal specificity;

(d) identifying from said plurality of mutated polypeptides a region of said Cry3Bb polypeptide for targeted mutagenesis; and

(e) mutagenizing said region to obtain said Cry3Bb polypeptide having improved insecticidal activity or enhanced insecticidal specificity.

106. The method of claim 105, further comprising determining the amino acid sequences of a plurality of mutated polypeptides having diminished activity or reduced insecticidal specificity, and identifying one or more amino acid residues required for said activity or specificity.

107. A method for producing a Cry3Bb polypeptide having improved insecticidal activity, comprising:

- (a) obtaining a high-resolution three-dimensional crystal structure of said polypeptide;
- 5 (b) determining the electrostatic surface distribution of said polypeptide;
- (c) identifying one or more regions of high electrostatic diversity;
- (d) modifying the electrostatic diversity of said region by altering one or
10 more amino acids in said region; and
- (e) obtaining said Cry3Bb polypeptide having improved insecticidal activity.

15

108. The method of claim 107, wherein said electrostatic diversity is decreased relative to the electrostatic diversity of a native Cry3Bb polypeptide.
- 20 109. The method of claim 107, wherein said electrostatic diversity is increased relative to the electrostatic diversity of a native Cry3Bb polypeptide.

110. A method of producing a Cry3Bb polypeptide having improved insecticidal activity, comprising:

(a) obtaining a high-resolution three-dimensional crystal structure;

5

(b) identifying the presence of one or more metal binding sites in said polypeptide;

(c) altering one or more amino acids in said binding site; and

10

(d) obtaining an altered polypeptide, wherein said polypeptide has improved insecticidal activity.

15 111. The method of claim 110, wherein said altering eliminates metal binding.

112. A method of identifying a Cry3Bb polypeptide having improved channel activity, comprising:

20

(a) obtaining a Cry3Bb polypeptide suspected of having improved channel activity;

(b) determining one or more of the following characteristics in said polypeptide, and in a wild-type polypeptide: the rate of channel formation, the rate of growth of channel conductance or the duration of open channel state;

5 (c) comparing said characteristics of said mutant and said wild-type; and

(d) identifying said polypeptide having an increased rate of channel formation compared to said wildtype polypeptide.

10

113. A method for producing a modified Cry3Bb polypeptide, having improved insecticidal activity, comprising altering one or more non-surface amino acids located at or near the point of greatest convergence of two or more loop regions of said Cry3Bb polypeptide, wherein said altering decreases the mobility
15 of one or more of said loop regions.

114. The method of claim 113, wherein said mobility is determined by comparing the thermal denaturation of said modified protein to a wild-type Cry3Bb
20 polypeptide.

115. A method of improving the insecticidal activity of a Cry3 polypeptide, said method comprising inserting one or more protease sensitive sites into one or more loop regions of domain 1 of said polypeptide.

5

116. The method of claim 115, wherein said loop region is $\alpha 3,4$.



FIG. 1



FIG. 2

FIG. 3A

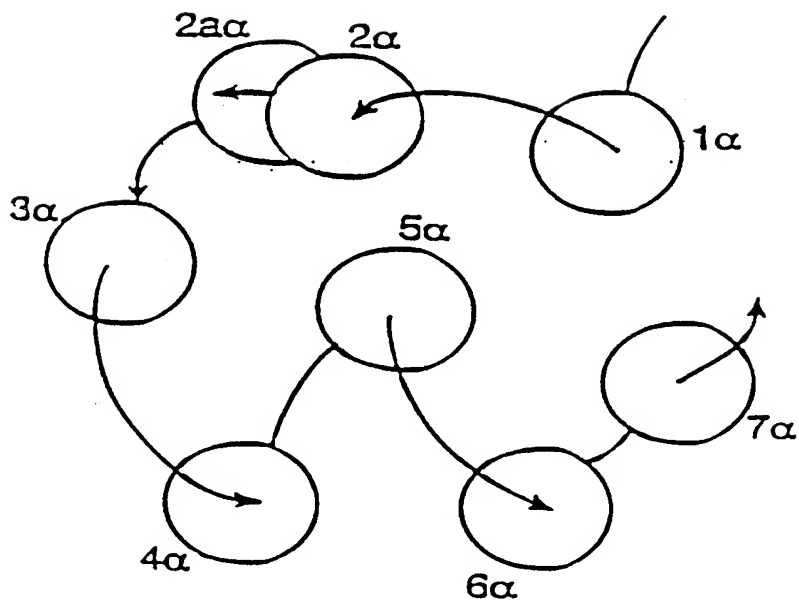
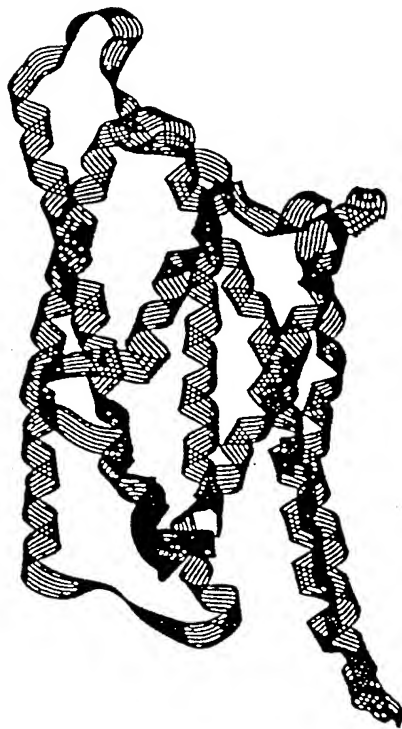


FIG. 3B

alpha helix	Amino acid Residues
$\alpha 1$	63-79
$\alpha 2a$	85-98
$\alpha 2b$	105-118
$\alpha 3$	124-153
$\alpha 4$	161-186
$\alpha 5$	194-215
$\alpha 6$	223-255
$\alpha 7$	260-286

FIG. 4

FIG. 5A

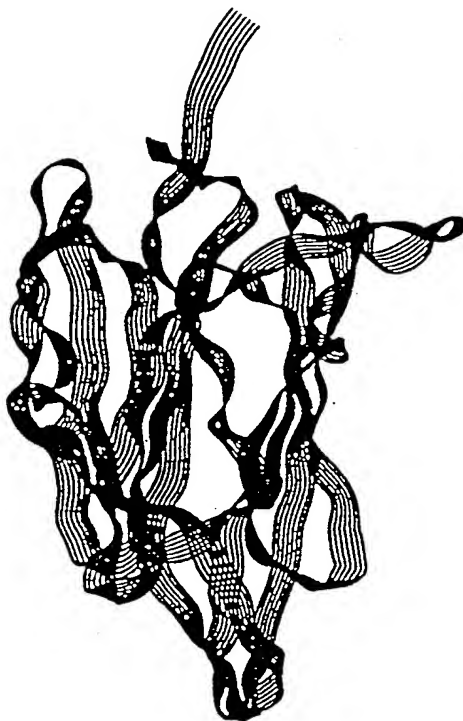
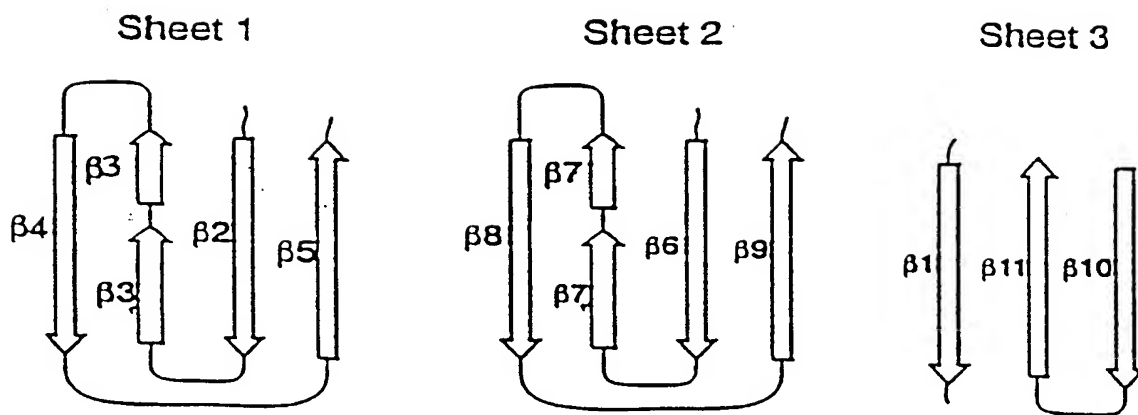


FIG. 5B



Sheet 1

β Strand	Amino Acid Residue
$\beta 2$	339-350
$\beta 3a$	256-360
$\beta 3b$	362-368
$\beta 4$	375-379
$\beta 5$	390-395

Sheet 2

β Strand	Amino Acid Residue
$\beta 6$	402-412
$\beta 7a$	416-419
$\beta 7b$	423-430
$\beta 8$	435-442
$\beta 9$	452-456

Sheet 3

β.Strand	Amino Acid Residue
$\beta 1$	296-306
$\beta 10$	472-483
$\beta 11$	492-498

FIG. 6

FIG. 7A

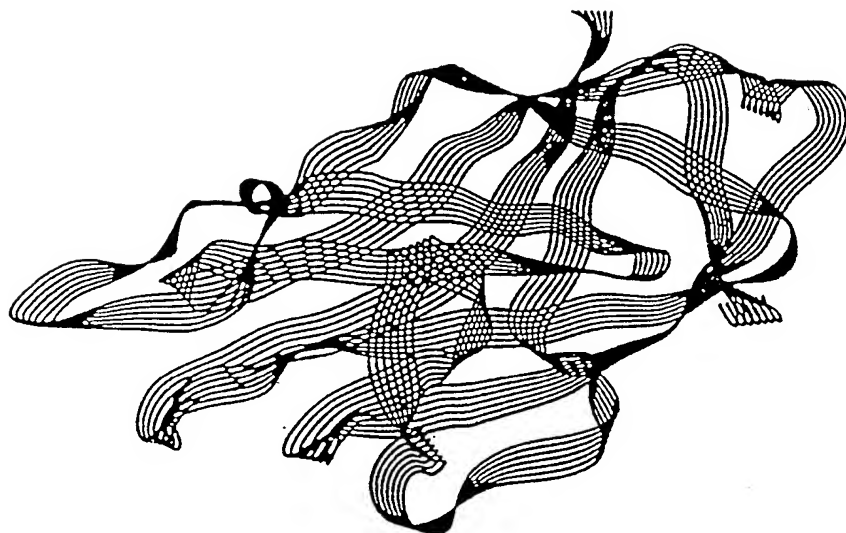
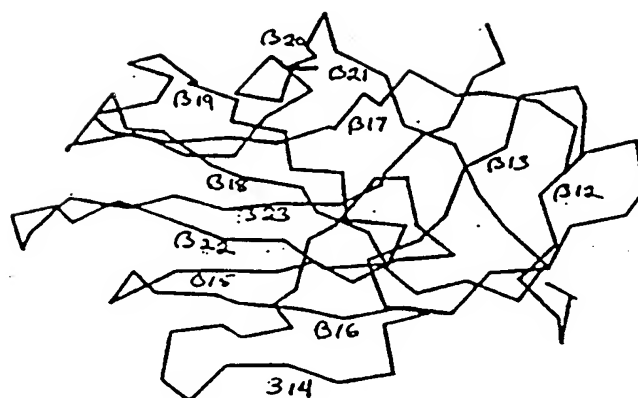


FIG. 7B



Strand Number	Amino Acid Residues
β 12	505-509
β 13	512-515
β 14	522-528
β 15	539-544
β 16	550-557
β 17	563-574
β 18	578-584
β 19	590-596
β 20	609-614
β 21	616-619
β 22	626-636
β 23	638-650

FIG. 8

FIG. 9A

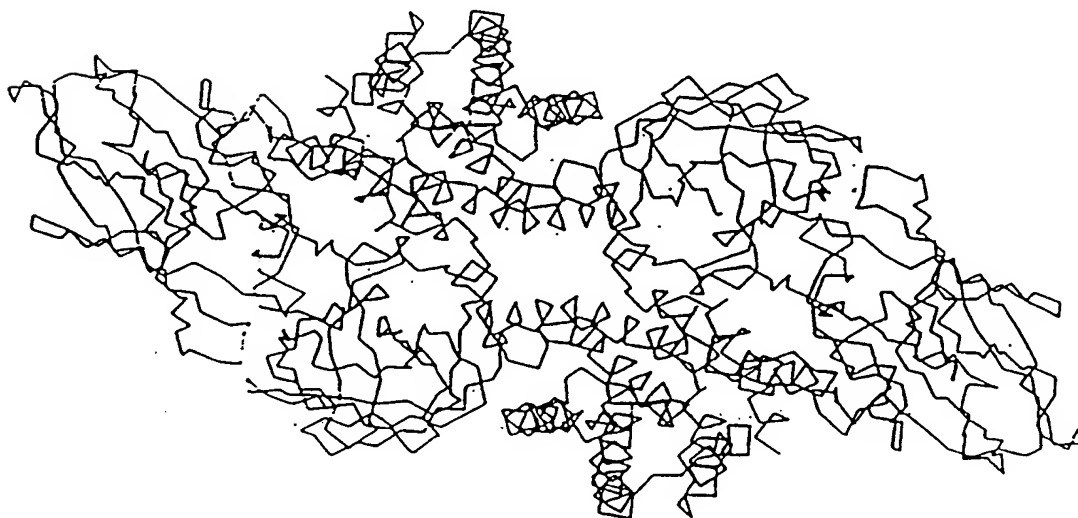
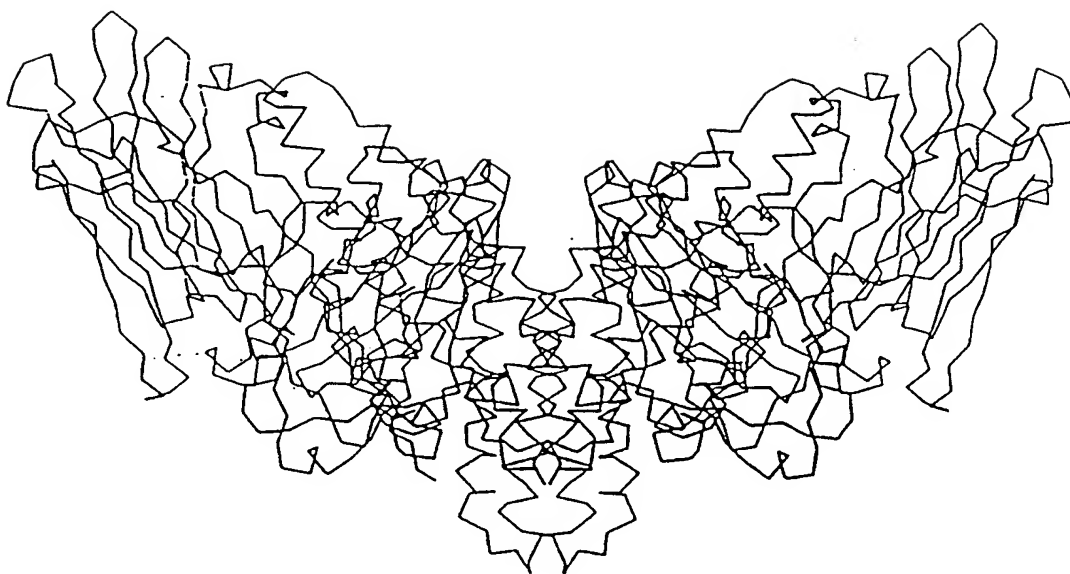
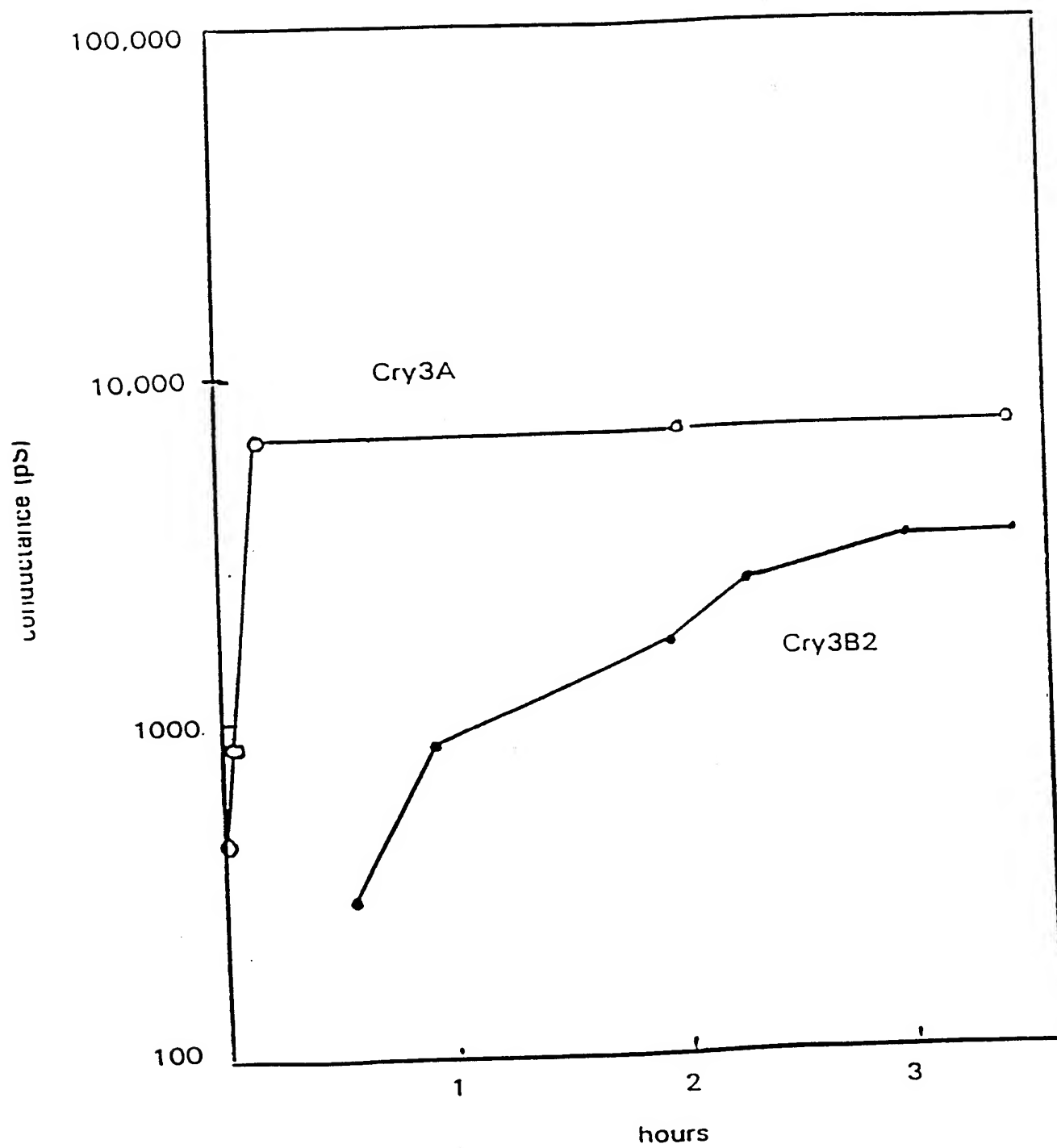


FIG. 9B

FIG. 10



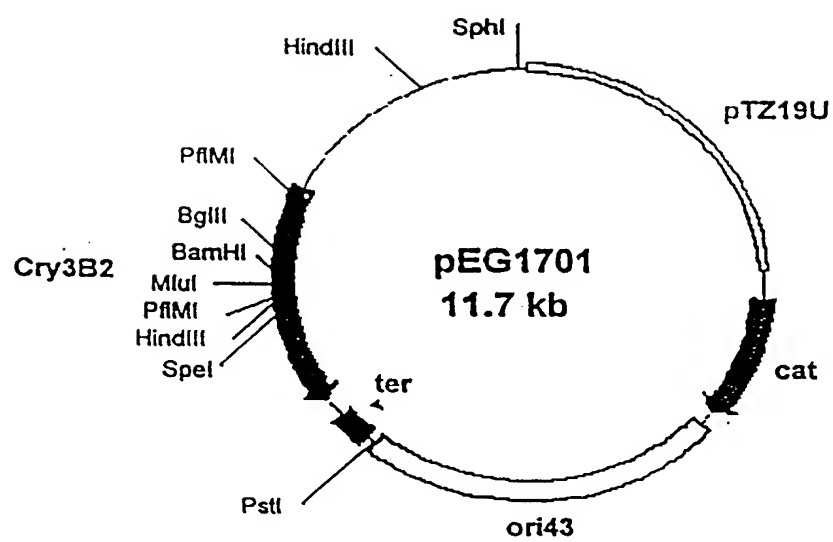


FIG. 11

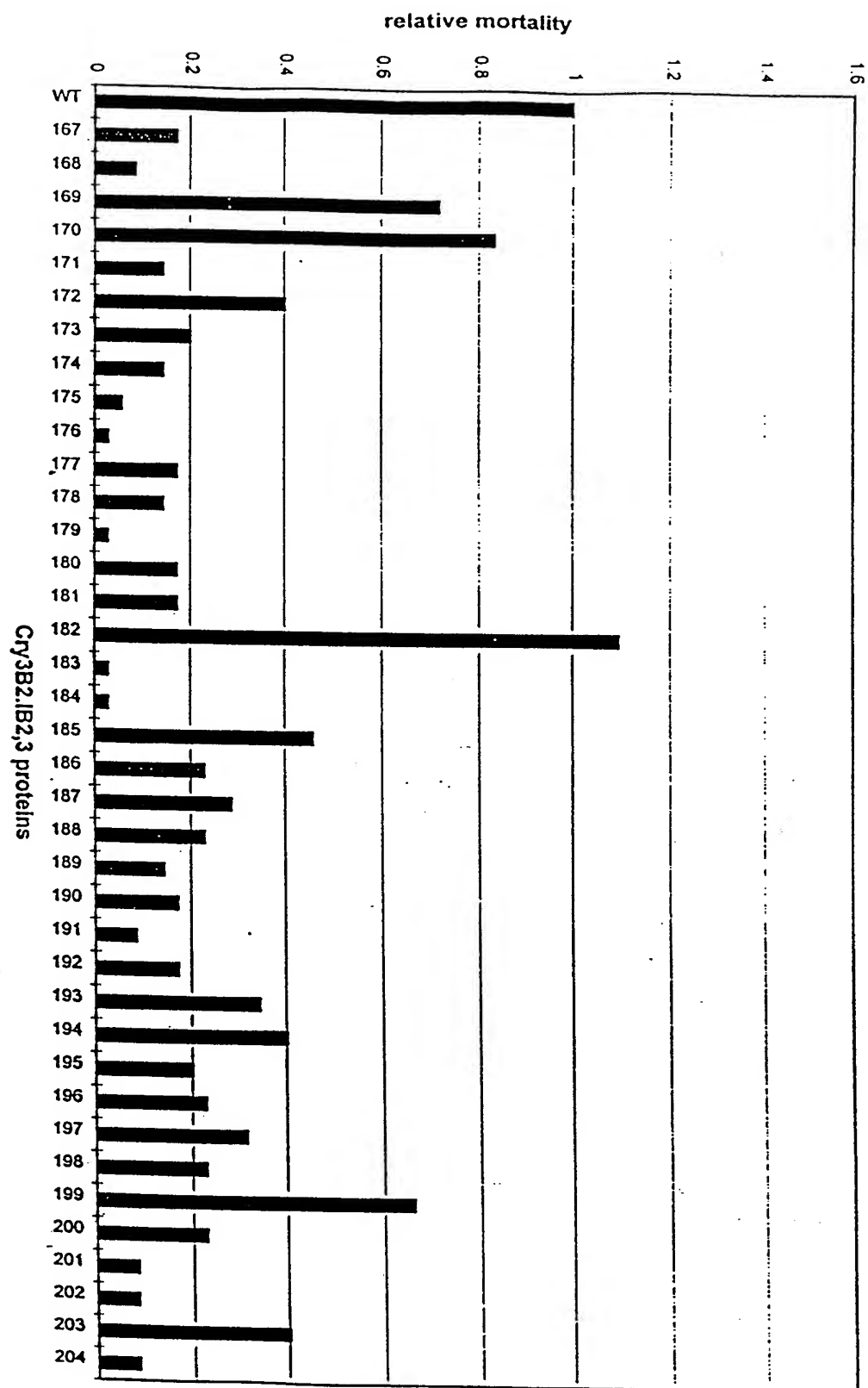


FIG. 12

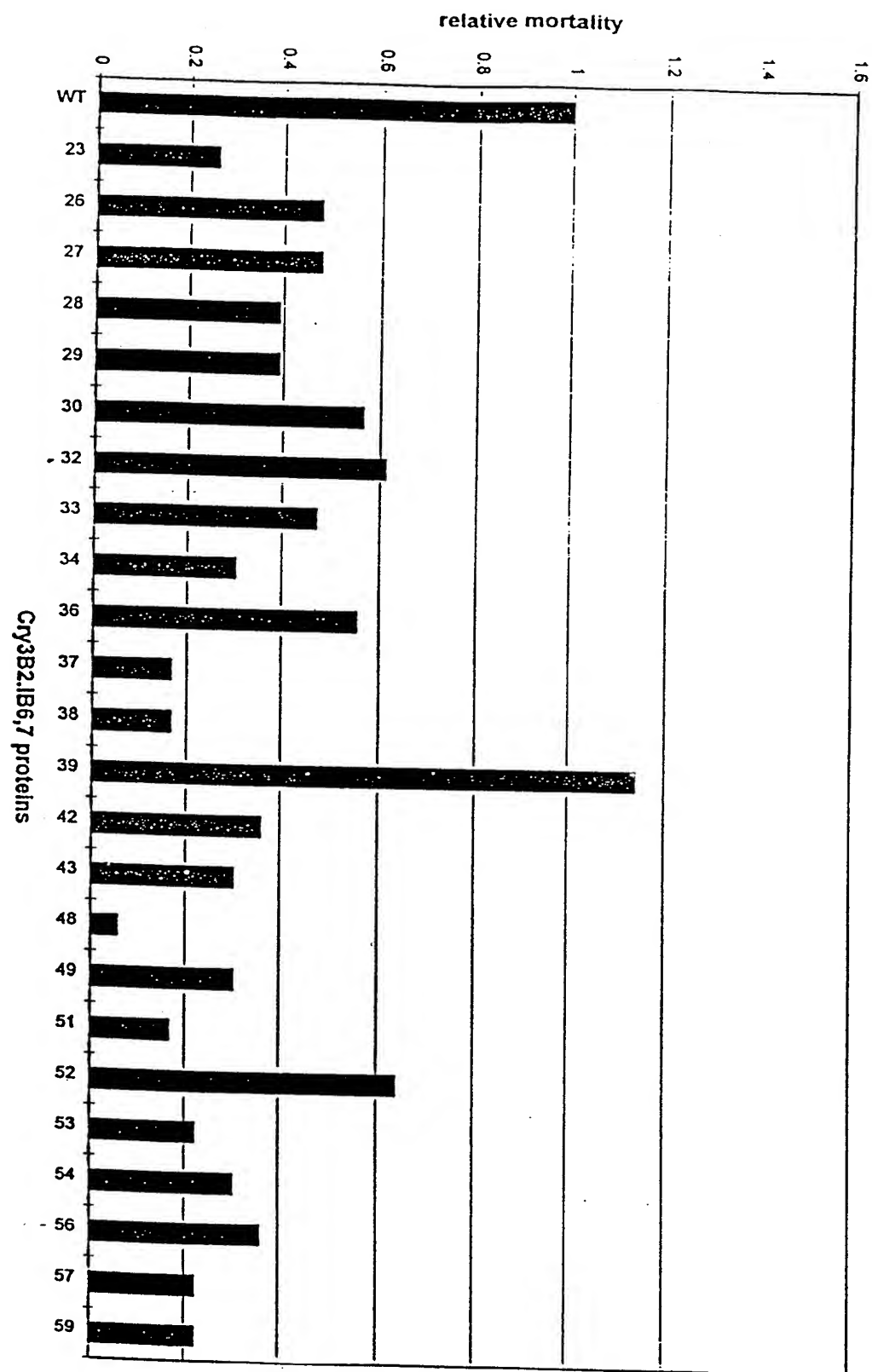


FIG. 13

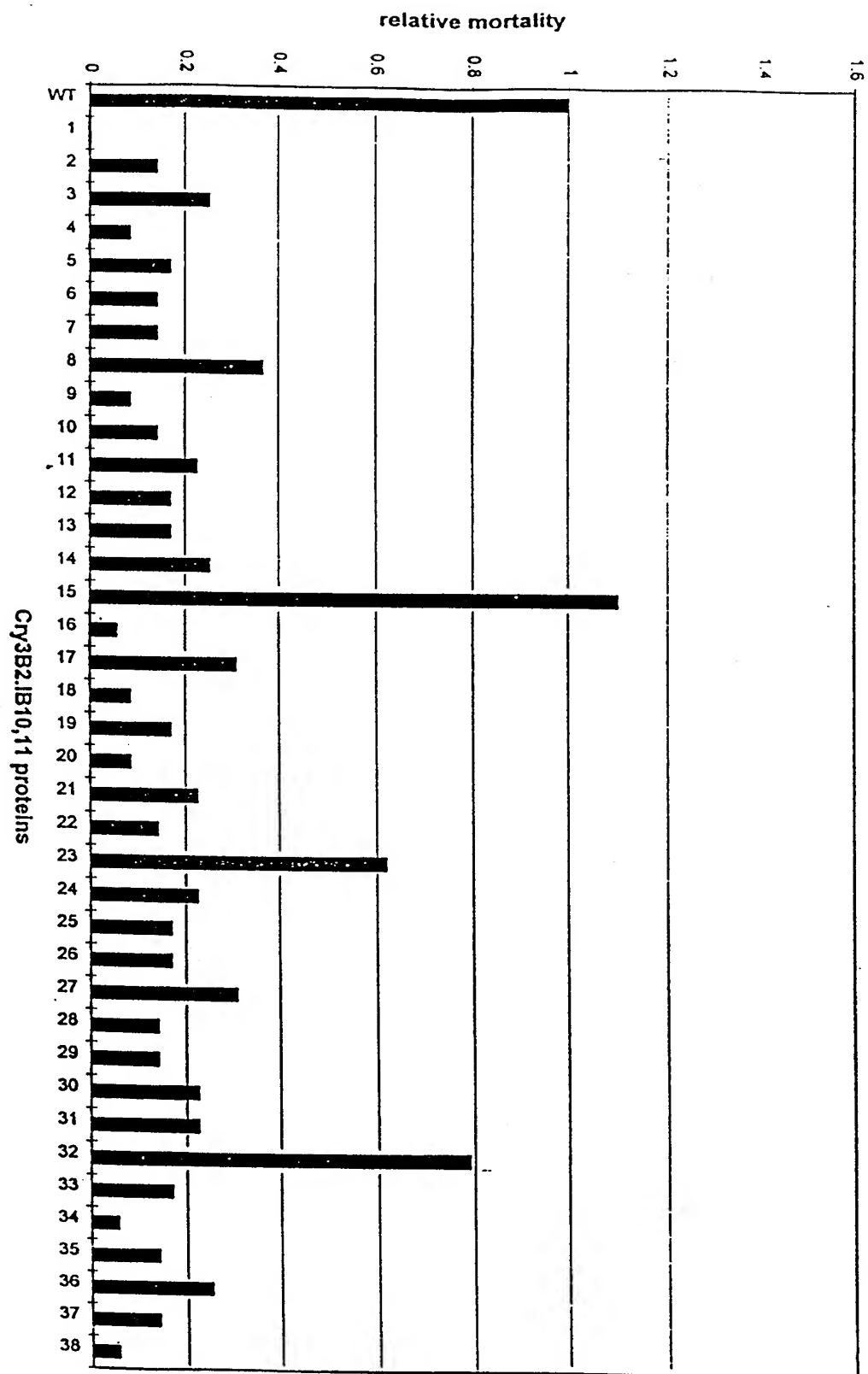


FIG. 14

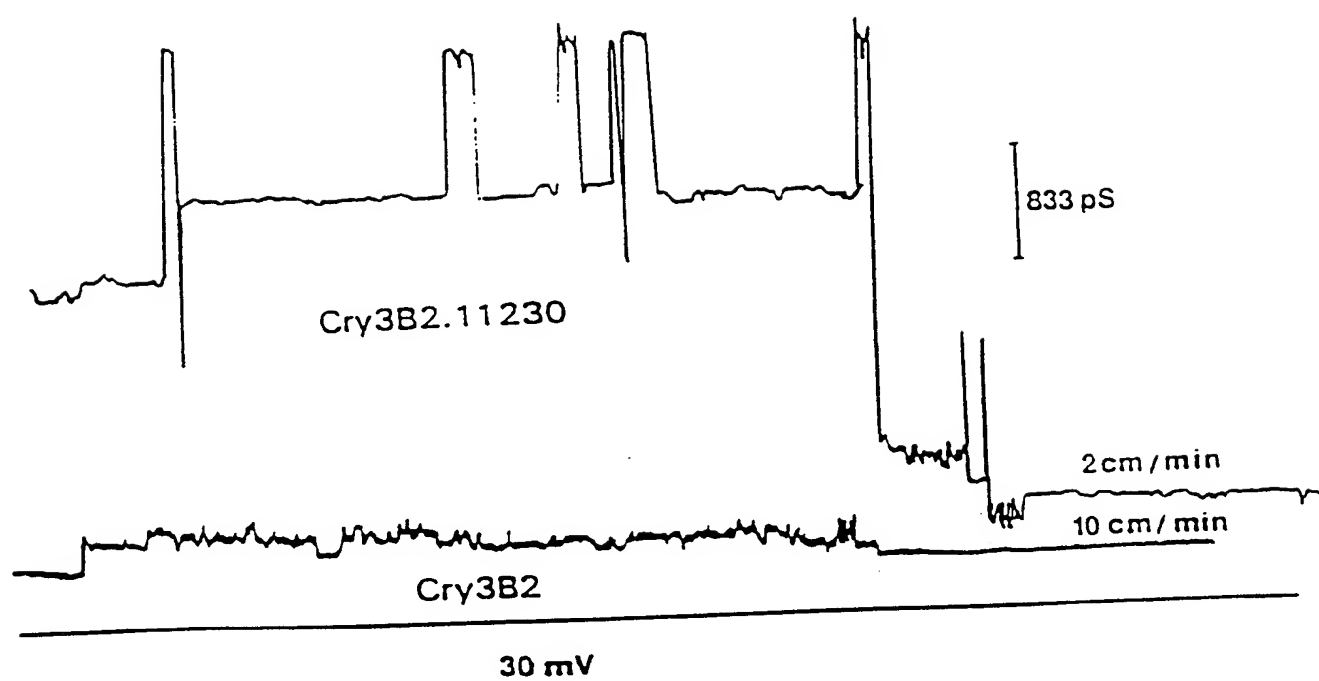


FIG. 15

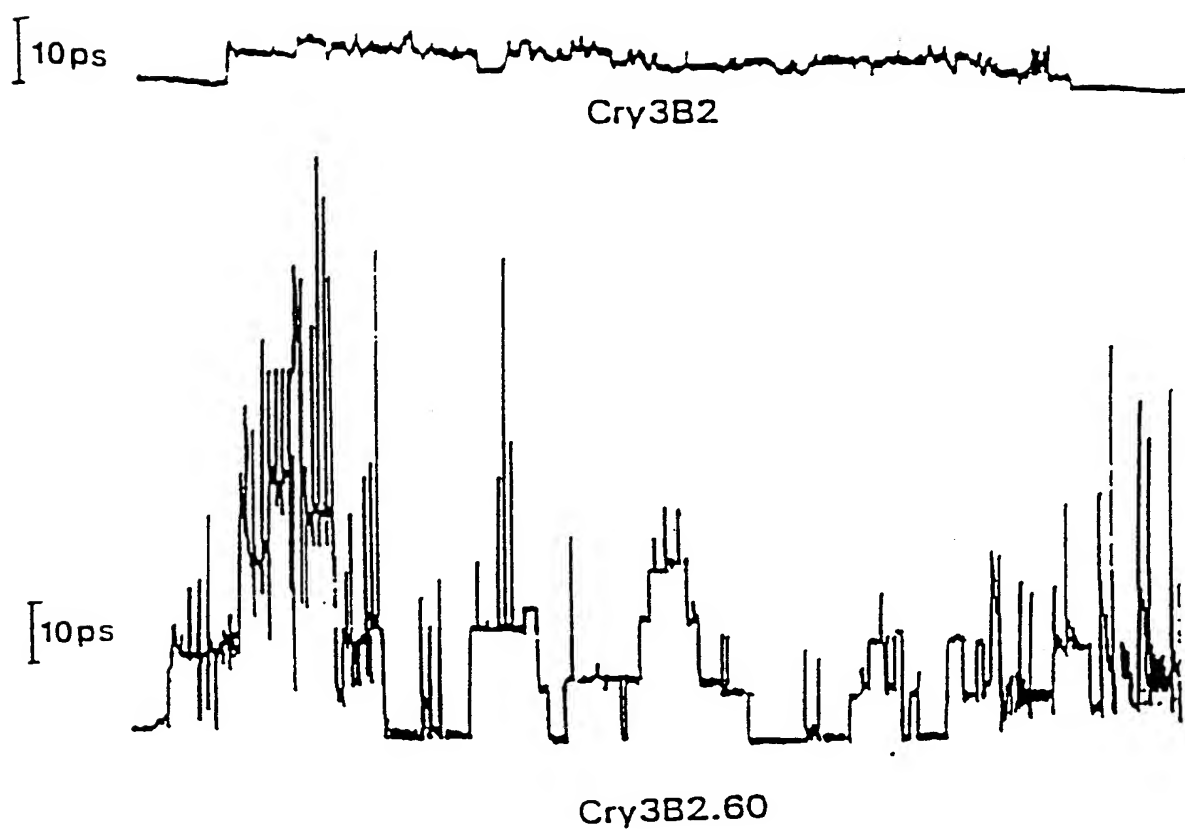


FIG. 16

ALIGNMENT OF CRY3 SEQUENCES

(Numbered according to Cry3BB)
(alpha helices underlined, beta sheets marked with +++'s)

	1	10	20	30	40
CRY3C:	MNPNNRSEHDTIKATENNEVSNNHAQYPIADTP--TLEELNY				
CRY3BB2:	MNPNNRSEHDTIKVTPNSELPTNHNQYPLADNPNSTLEELNY				
CRY3BB:	MNPNNRSEHDTIKVTPNSELQTNHNQYPLADNPNSTLEELNY				
CRY3BA:	MIRMGGRKMNPNNRSEYDTIKVTPNSELPTNHNQYPLADNPNSTLEELNY				
CRY3A:	MIRKGRKMNPNNRSEHDTIKTTENNEVPTNHHVQYPLAETPNPTLEDLNY				

	50	60	70	80	90
CRY3C:	KEFLRRTTDNNVEALDSSTTKDAIQKGISIIGDLLGVVGFPPYGGALVSFY				
CRY3BB2:	KEFLRMTEDSSTEVLNDSTVKDAVGTGISVVGQILGVVGVPFAGALTSFY				
CRY3BB:	KEFLRMTEDSSTEVLNDSTVKDAVGTGISVVGQILGVVGVPFAGALTSFY				
CRY3BA:	KEFLRMTADNSTEVLDSSTVKDAVGTGISVVGQILGVVGVPFAGALTSFY				
CRY3A:	KEFLRMTADNNTEALDSSTTKDVIQKGISVVGDLLGVVGFPPFGGALVSFY				

	100	110	120	130	140
CRY3C:	TNLLNTIWPGE-DPLKAFMQQVEALIDQKIADYAKDKATAELQGLKNVFK				
CRY3BB2:	QSFLDTIWPSDADPWKAFMAQVEVLIDKKIEEYAKSKALAELOGLQNNFE				
CRY3BB:	QSFLNTIWPSDADPWKAFMAQVEVLIDKKIEEYAKSKALAELOGLQNNFE				
CRY3BA:	QSFLNAIWPSDADPWKAFMAQVEVLIDKKIEEYAKSKALAELOGLQNNFE				
CRY3A:	<u>TNFLNTIWPSE-DPWKAFMEQVEALMDQKIADYAKNKALAELOGLQNNVE</u>				

	150	160	170	180	190
CRY3C:	DYVSALDSWDKTPLTLRDGRSQGRIRELFSQAESHFRNSMPSPFAVSGYEV				
CRY3BB2:	DYVNALNSWKKTPLSLRSKRSQDRIRELFSQAESHFRNSMPSPFAVSKFEV				
CRY3BB:	DYVNALNSWKKTPLSLRSKRSQDRIRELFSQAESHFRNSMPSPFAVSKFEV				
CRY3BA:	DYVNALDSWKKAPVNLRSRRSQDRIRELFSQAESHFRNSMPSPFAVSKFEV				
CRY3A:	<u>DYVSALSSWOKNPVSSRNPHSQGRIRELFSQAESHFRNSMPSPFAISGYEV</u>				

FIG. 17A

	200	210	220	230	240
CRY3C:	LFLPTYAQAANTHLL	LLKDAQIYGT	DWGYSTDDL	NEFHTKQKDL	TIEYTN
CRY3BB2:	LFLPTYAQAANTHLL	LLKDAQVFGE	EWGYSS	EDVAEFYHRQL	KLTQQYTD
CRY3BB:	LFLPTYAQAANTHLL	LLKDAQVFGE	EWGYSS	EDVAEFYHRQL	KLTQQYTD
CRY3BA:	LFLPTYAQAANTHLL	LLKDAQVFGE	EWGYSS	EDIAEFYQRQL	KLTQQYTD
CRY3A:	<u>LFLTTYAQAANTH</u>	<u>LFLLKDAQIY</u>	<u>GE</u>	<u>EWGYEKED</u>	<u>IAEFYKROLKLT</u>

	250	260	270	280	290
CRY3C:	HCAKWKAGLDKLRG	STYEEWVKFN	RNRREMTL	TVLDLITL	FPLYDVRTY
CRY3BB2:	HCVNWYNVGLNGLR	GSTYDAWVKFN	RFRREMTL	TVLDLIVL	FPPFYDVRLY
CRY3BB:	HCVNWYNVGLNGLR	GSTYDAWVKFN	RFRREMTL	TVLDLIVL	FPPFYDIRLY
CRY3BA:	HCVNWYNVGLNSLR	GSTYDAWVKFN	RFRREMTL	TVLDLIVL	FPPFYDVRLY
CRY3A:	<u>HCVKWYNVGL</u>	<u>DKLRGSSYES</u>	<u>WVN</u>	<u>FN</u>	<u>RNRREMTLTVLDL</u>

	300	310	320	330	340
CRY3C:	TKGVKTELTRDVL	TDPIVAVNNM	NGYGTTF	SNIENYIRK	PHLFDYLHAIQ
CRY3BB2:	SKGVKTELTRDIF	TDPIFSLN	TLQEYGPT	FLSIENSIRK	PHLFDYLGIE
CRY3BB:	SKGVKTELTRDIF	TDPIFSLN	TLQEYGPT	FLSIENSIRK	PHLFDYLGIE
CRY3BA:	SKGVKTELTRDIF	TDPIFTL	NALQEYGPT	FSSIENSIRK	PHLFDYLRGIE
CRY3A:	<u>PKEVKTELTRDVL</u>	<u>TDPIVGVNN</u>	<u>LRGYGTTF</u>	<u>SNIENYIRK</u>	<u>PHLFDYLHRIQ</u>
	+++++				++++

	350	360	370	380	390
CRY3C:	FHSRLQPGYFGT	DSFNWWSGNY	VSTRSSIGS	DEIIRSPFY	GKNSTLDVQN
CRY3BB2:	FHTRLQPGYSGK	DSFNWWSGNY	VETRPSIGS	SKTITSPFY	GDKSTEPVQK
CRY3BB:	FHTRLQPGYFGK	DSFNWWSGNY	VETRPSIGS	SKTITSPFY	GDKSTEPVQK
CRY3BA:	FHTRLRPGYSGK	DSFNWWSGNY	VETRPSIGS	NDTITSPFY	GDKSIEPIQK
CRY3A:	<u>FHTRFQPGY</u>	<u>GNDSFNWWSGNY</u>	<u>VSTRPSIGS</u>	<u>NDIITSPFY</u>	<u>GKNKSSEPVQN</u>
	+++++	+++++	+++++	+++++	+++

	400	410	420	430
CRY3C:	LEFNGEKVFR	AVANGNLAV	WPVGTGGT	KIHSGVTKVQFSQY
CRY3BB2:	LSFDGQKVYRT	IAN	TDVA	AWPNG----
CRY3BB:	LSFDGQKVYRT	IAN	TDVA	AWPNG----
CRY3BA:	LSFDGQKVYRT	IAN	TDIA	AFPDG----
CRY3A:	<u>LEFNGEKVYRA</u>	<u>VANTNLAV</u>	<u>WPSA----</u>	<u>VYSGVTKVEFSQY</u>
	+++	+++++	++++	+++++

FIG. 17B

```

      440      450      460      470      480
CRY3C:  QTYDSKRNVGGIV-FDSIDQLPPITTTDESLEKAYSHQLNYVRCFLLQGGR
CRY3BB2: QTYDSKRNNGHVGAQDSIDQLPPETTDEPLEKAYSHQLNYAECFLMQDRR
CRY3BB:  QTYDSKRNNGHVSAQDSIDQLPPETTDEPLEKAYSHQLNYAECFLMQDRR
CRY3BA:  QTYDSKRYNGYLGAQDSIDQLPPETTDEPLEKAYSHQLNYAECFLMQDRR
CRY3A:   QTYDSKRNVGAVS-WDSIDQLPPETTDEPLEKGYSHQLNYVMCFLMQGSR
      +++++      +++++      ++++++

      490      500      510      520      530
CRY3C:  GIIPVFTWTHKSVDFYNTLDSEKITQIPFVKAFILVNSTSVVAGPGFTGG
CRY3BB2: GTIPFFTWTTHRSVDFNTIDAELITQLPVVKAYALSSGASIIIEGPGFTGG
CRY3BB:  GTIPFFTWTTHRSVDFNTIDAELITQLPVVKAYALSSGASIIIEGPGFTGG
CRY3BA:  GTIPFFTWTTHRSVDFNTIDAELITQLPVVKAYALSSGASIIIEGPGFTGG
CRY3A:   GTIPVLTWTHKSVDFNMDISKKITQLPLVKAYKLQSGASVVAGPRFTGG
      ++++++      +++++      +++++      ++++++

      540      550      560      570      580
CRY3C:  DII-KCT-NGSGLTLYVTPAPDLTYSKTYKIRIRYASTSQVRFGIDLGSY
CRY3BB2: NLLFLKESSNSIAKFKVTL-NSAALLQRYRVIRIRYASTTNLRLFVQNSNN
CRY3BB:  NLLFLKESSNSIAKFKVTL-NSAALLQRYRVIRIRYASTTNLRLFVQNSNN
CRY3BA:  NLLFLKESSNSIAKFKVTL-NSAALLQRYRVIRIRYASTTNLRLFVQNSNN
CRY3A:   DII-QCTENGSAATIIYVTPD--VSYSQKYRARIHYASTSQITFTLSLDGA
      ++++++      ++++++      ++++++      ++++++

      590      600      610      620      630
CRY3C:  THSISYFDKTMKGNLTLYNSFNLSSVSRPIEISG-GNKIGVSVGGIGSG
CRY3BB2: DFIVIIYINKTMNIDDDLTQTFDLATTNSNMGFSGDTNELIIGAESFVSN
CRY3BB:  DFLVIYINKTMNKDDDLTYQTFDLATTNSNMGFSGDKNELIIGAESFVSN
CRY3BA:  DFLVIYINKTMNIDGDLTYQTFDFATSNSNMGFSGDTNDFIIGAESFVSN
CRY3A:   PFNQYYFDKTINKGDTLTLYNSFNLASFSTPFELSG--NNLQIGVTGLSAG
      ++++++      ++++++      ++++++      ++++++

      640      650
CRY3C:  DEVYIDKIEFIPMD
CRY3BB2: EKIIYIDKIEFIPVQL
CRY3BB:  EKIIYIDKIEFIPVQL
CRY3BA:  EKIIYIDKIEFIPVQ
CRY3A:   DKVYIDKIEFIPVN
      ++++++

```

FIG. 17C

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

(A) NAME: ECOGEN, INC./MONSANTO COMPANY
(B) STREET: 2005 CABOT BLVD W/700 CHESTERFIELD VILLAGE
PKY N
(C) CITY: LANGHORNE/ST. LOUIS
(D) STATE: PA/MO
(E) COUNTRY: USA
(F) POSTAL CODE (ZIP): 819047/63198

(A) NAME: LEIGH H. ENGLISH
(B) STREET: 120 CHAPEL DR
(C) CITY: CHRUCHVILLE
(D) STATE: PA
(E) COUNTRY: USA
(F) POSTAL CODE (ZIP): 18966

(A) NAME: SUSAN M. BRUSSOCK
(B) STREET: 7 HILLSIDE LN
(C) CITY: NEW HOPE
(D) STATE: PA
(E) COUNTRY: USA
(F) POSTAL CODE (ZIP): 18938

(A) NAME: THOMAS M. MALVAR
(B) STREET: 12046 CHARTER HOUSE LN
(C) CITY: ST. LOUIS
(D) STATE: MO
(E) COUNTRY: USA
(F) POSTAL CODE (ZIP): 63146

(A) NAME: JAMES W. BRYSON
(B) STREET: 87 WOOD STREAM DR
(C) CITY: LANGHORNE
(D) STATE: PA
(E) COUNTRY: USA
(F) POSTAL CODE (ZIP): 19053

(A) NAME: CAROLINE A. KULESZA
(B) STREET: 301 OLD LYNCHBURG RD
(C) CITY: CHARLOTTESVILLE
(D) STATE: VA
(E) COUNTRY: USA
(F) POSTAL CODE (ZIP): 22903

(A) NAME: FREDERICK S. WALTERS
(B) STREET: 3413 6TH AVE
(C) CITY: BEAVER FALLS
(D) STATE: PA
(E) COUNTRY: USA
(F) POSTAL CODE (ZIP): 15010

(A) NAME: STEPHEN L. SLATIN
(B) STREET: 3823 LESLIE PL
(C) CITY: FAIR LAWN
(D) STATE: NJ
(E) COUNTRY: USA
(F) POSTAL CODE (ZIP): 07410

(A) NAME: MICHAEL A. VON TERSCH
(B) STREET: 14 RUTLEDGE AVE
(C) CITY: TRENTON
(D) STATE: NJ
(E) COUNTRY: USA
(F) POSTAL CODE (ZIP): 08618

(A) NAME: CHARLES ROMANO
(B) STREET: 2402 MAPLE CROSSING DR
(C) CITY: WILDWOOD
(D) STATE: MO
(E) COUNTRY: USA
(F) POSTAL CODE (ZIP): 63011

(ii) TITLE OF INVENTION: INSECT-RESISTANT TRANSGENIC PLANTS AND
METHODS FOR IMPROVING DELTA-ENDOTOXIN ACTIVITY AGAINST
TARGET INSECTS

(iii) NUMBER OF SEQUENCES: 113

(iv) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

(v) CURRENT APPLICATION DATA:
APPLICATION NUMBER: UNKNOWN

(vi) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: US 08/993,170
(B) FILING DATE: 18-DEC-1997

(vi) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: US 08/993,722
(B) FILING DATE: 18-DEC-1997

(vi) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: US 08/993,775
(B) FILING DATE: 18-DEC-1997

(vi) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: US 08/996,441
(B) FILING DATE: 18-DEC-1997

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1959 base pairs

(B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA TTT CAC CAT TCT CGT CGT TCT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Phe His His Ser Arg Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	

CAT	TTT	CGT	AAT	TCC	ATG	CCG	TCA	TTT	GCA	GTT	TCC	AAA	TTC	GAA	GTG	576
His	Phe	Arg	Asn	Ser	Met	Pro	Ser	Phe	Ala	Val	Ser	Lys	Phe	Glu	Val	
			180					185					190			
CTG	TTT	CTA	CCA	ACA	TAT	GCA	CAA	GCT	GCA	AAT	ACA	CAT	TTA	TTG	CTA	624
Leu	Phe	Leu	Pro	Thr	Tyr	Ala	Gln	Ala	Ala	Asn	Thr	His	Leu	Leu	Leu	
		195					200					205				
TTA	AAA	GAT	GCT	CAA	GTT	TTT	GGA	GAA	GAA	TGG	GGA	TAT	TCT	TCA	GAA	672
Leu	Lys	Asp	Ala	Gln	Val	Phe	Gly	Glu	Glu	Trp	Gly	Tyr	Ser	Ser	Glu	
	210						215				220					
GAT	GTT	GCT	GAA	TTT	TAT	CAT	AGA	CAA	TTA	AAA	CTT	ACA	CAA	CAA	TAC	720
Asp	Val	Ala	Glu	Phe	Tyr	His	Arg	Gln	Leu	Lys	Leu	Thr	Gln	Gln	Tyr	
225					230					235					240	
ACT	GAC	CAT	TGT	GTT	AAT	TGG	TAT	AAT	GTT	GGA	TTA	AAT	GGT	TTA	AGA	768
Thr	Asp	His	Cys	Val	Asn	Trp	Tyr	Asn	Val	Gly	Leu	Asn	Gly	Leu	Arg	
			245					250						255		
GGT	TCA	ACT	TAT	GAT	GCA	TGG	GTC	AAA	TTT	AAC	CGT	TTT	CGC	AGA	GAA	816
Gly	Ser	Thr	Tyr	Asp	Ala	Trp	Val	Lys	Phe	Asn	Arg	Phe	Arg	Arg	Glu	
			260					265					270			
ATG	ACT	TTA	ACT	GTA	TTA	GAT	CTA	ATT	GTA	CTT	TTC	CCA	TTT	TAT	GAT	864
Met	Thr	Leu	Thr	Val	Leu	Asp	Leu	Ile	Val	Leu	Phe	Pro	Phe	Tyr	Asp	
		275					280					285				
ATT	CGG	TTA	TAC	TCA	AAA	GGG	GTT	AAA	ACA	GAA	CTA	ACA	AGA	GAC	ATT	912
Ile	Arg	Leu	Tyr	Ser	Lys	Gly	Val	Lys	Thr	Glu	Leu	Thr	Arg	Asp	Ile	
	290					295					300					
TTT	ACG	GAT	CCA	ATT	TTT	TCA	CTT	AAT	ACT	CTT	CAG	GAG	TAT	GGA	CCA	960
Phe	Thr	Asp	Pro	Ile	Phe	Ser	Leu	Asn	Thr	Leu	Gln	Glu	Tyr	Gly	Pro	
305				310					315					320		
ACT	TTT	TTG	AGT	ATA	GAA	AAC	TCT	ATT	CGA	AAA	CCT	CAT	TTA	TTT	GAT	1008
Thr	Phe	Leu	Ser	Ile	Glu	Asn	Ser	Ile	Arg	Lys	Pro	His	Leu	Phe	Asp	
			325					330					335			
TAT	TTA	CAG	GGG	ATT	GAA	TTT	CAT	ACG	CGT	CTT	CAA	CCT	GGT	TAC	TTT	1056
Tyr	Leu	Gln	Gly	Ile	Glu	Phe	His	Thr	Arg	Leu	Gln	Pro	Gly	Tyr	Phe	
			340					345					350			
GGG	AAA	GAT	TCT	TTC	AAT	TAT	TGG	TCT	GGT	AAT	TAT	GTA	GAA	ACT	AGA	1104
Gly	Lys	Asp	Ser	Phe	Asn	Tyr	Trp	Ser	Gly	Asn	Tyr	Val	Glu	Thr	Arg	
		355					360					365				
CCT	AGT	ATA	GGA	TCT	AGT	AAG	ACA	ATT	ACT	TCC	CCA	TTT	TAT	GGA	GAT	1152
Pro	Ser	Ile	Gly	Ser	Ser	Lys	Thr	Ile	Thr	Ser	Pro	Phe	Tyr	Gly	Asp	
	370					375										

TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG	1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys	
405 410 415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA	1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln	
420 425 430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC	1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly	
435 440 445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA	1392
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr	
450 455 460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA	1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu	
465 470 475 480	
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG	1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	
485 490 495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT	1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	
500 505 510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	

AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC 1920
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640

TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA 1959
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
 1 5 10 15
 Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
 20 25 30
 Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
 35 40 45
 Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50 55 60
 Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65 70 75 80
 Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95
 Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110
 Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125
 Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140
 Val Asn Ala Leu Asn Ser Trp Lys Lys Phe His His Ser Arg Arg Ser
 145 150 155 160
 Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205

Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220

Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240

Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255

Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270

Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285

Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300

Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320

Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335

Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350

Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365

Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380

Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400

Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415

Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430

Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445

His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460

Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480

Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495

Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575
 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590
 Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605
 Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	

CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met 35 40 45	144
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp 50 55 60	192
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val 65 70 75 80	240
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu 85 90 95	288
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala 100 105 110	336
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser 115 120 125	384
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr 130 135 140	432
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser 145 150 155 160	480
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser 165 170 175	528
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val 180 185 190	576
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu 195 200 205	624
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220	672
GAT GTT GCT GAA TTC CTT AGT AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Leu Ser Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240	720
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255	768

GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 265 270	816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 275 280 285	864
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300	912
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro 305 310 315 320	960
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp 325 330 335	1008
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe 340 345 350	1056
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg 355 360 365	1104
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp 370 375 380	1152
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val 385 390 395 400	1200
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys 405 410 415	1248
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln 420 425 430	1296
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly 435 440 445	1344
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr 450 455 460	1392
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu 465 470 475 480	1440

TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG	1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	
485 490 495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT	1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	
500 505 510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC	1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile	
625 630 635 640	
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro

12

1	5	10	15
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	20	25	30
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	35	40	45
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	50	55	60
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	65	70	75
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	85	90	95
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	100	105	110
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	115	120	125
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	130	135	140
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	145	150	155
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	165	170	175
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	180	185	190
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	195	200	205
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	210	215	220
Asp Val Ala Glu Phe Leu Ser Arg Gln Leu Lys Leu Thr Gln Gln Tyr	225	230	235
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	245	250	255
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	260	265	270
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	275	280	285
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	290	295	300

Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575
 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590
 Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605

Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
610 615 620

Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
625 630 635 640

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
645 650

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384

Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	
195 200 205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT CCA GAA	672
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Pro Glu	
210 215 220	
GAT GTT GCT GAA TTC AGT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC	720
Asp Val Ala Glu Phe Ser His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	
225 230 235 240	
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA	768
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	
245 250 255	
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA	816
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	
260 265 270	
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT	864
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	
275 280 285	
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT	912
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	
290 295 300	
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA	960
Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro	
305 310 315 320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT	1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	
325 330 335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT	1056

Tyr	Leu	Gln	Gly	Ile	Glu	Phe	His	Thr	Arg	Leu	Gln	Pro	Gly	Tyr	Phe	
			340					345					350			
GGG	AAA	GAT	TCT	TTC	AAT	TAT	TGG	TCT	GGT	AAT	TAT	GTA	GAA	ACT	AGA	1104
Gly	Lys	Asp	Ser	Phe	Asn	Tyr	Trp	Ser	Gly	Asn	Tyr	Val	Glu	Thr	Arg	
		355					360					365				
CCT	AGT	ATA	GGA	TCT	AGT	AAG	ACA	ATT	ACT	TCC	CCA	TTT	TAT	GGA	GAT	1152
Pro	Ser	Ile	Gly	Ser	Ser	Lys	Thr	Ile	Thr	Ser	Pro	Phe	Tyr	Gly	Asp	
		370				375					380					
AAA	TCT	ACT	GAA	CCT	GTA	CAA	AAG	CTA	AGC	TTT	GAT	GGA	CAA	AAA	GTT	1200
Lys	Ser	Thr	Glu	Pro	Val	Gln	Lys	Leu	Ser	Phe	Asp	Gly	Gln	Lys	Val	
385					390				395						400	
TAT	CGA	ACT	ATA	GCT	AAT	ACA	GAC	GTA	GCG	GCT	TGG	CCG	AAT	GGT	AAG	1248
Tyr	Arg	Thr	Ile	Ala	Asn	Thr	Asp	Val	Ala	Ala	Trp	Pro	Asn	Gly	Lys	
			405					410						415		
GTA	TAT	TTA	GGT	GTT	ACG	AAA	GTT	GAT	TTT	AGT	CAA	TAT	GAT	GAT	CAA	1296
Val	Tyr	Leu	Gly	Val	Thr	Lys	Val	Asp	Phe	Ser	Gln	Tyr	Asp	Asp	Gln	
		420					425					430				
AAA	AAT	GAA	ACT	AGT	ACA	CAA	ACA	TAT	GAT	TCA	AAA	AGA	AAC	AAT	GGC	1344
Lys	Asn	Glu	Thr	Ser	Thr	Gln	Thr	Tyr	Asp	Ser	Lys	Arg	Asn	Asn	Gly	
		435				440					445					
CAT	GTA	AGT	GCA	CAG	GAT	TCT	ATT	GAC	CAA	TTA	CCG	CCA	GAA	ACA	ACA	1392
His	Val	Ser	Ala	Gln	Asp	Ser	Ile	Asp	Gln	Leu	Pro	Pro	Glu	Thr	Thr	
	450				455					460						
GAT	GAA	CCA	CTT	GAA	AAA	GCA	TAT	AGT	CAT	CAG	CTT	AAT	TAC	GCG	GAA	1440
Asp	Glu	Pro	Leu	Glu	Lys	Ala	Tyr	Ser	His	Gln	Leu	Asn	Tyr	Ala	Glu	
465				470				475						480		
TGT	TTC	TTA	ATG	CAG	GAC	CGT	CGT	GGA	ACA	ATT	CCA	TTT	TTT	ACT	TGG	1488
Cys	Phe	Leu	Met	Gln	Asp	Arg	Arg	Gly	Thr	Ile	Pro	Phe	Phe	Thr	Trp	
		485					490						495			
ACA	CAT	AGA	AGT	GTA	GAC	TTT	TTT	AAT	ACA	ATT	GAT	GCT	GAA	AAG	ATT	1536
Thr	His	Arg	Ser	Val	Asp	Phe	Phe	Asn	Thr	Ile	Asp	Ala	Glu	Lys	Ile	
		500				505					510					
ACT	CAA	CTT	CCA	GTA	GTG	AAA	GCA	TAT	GCC	TTG	TCT	TCA	GGT	GCT	TCC	1584
Thr	Gln	Leu	Pro	Val	Val	Lys	Ala	Tyr	Ala	Leu	Ser	Ser	Gly	Ala	Ser	
		515				520					525					
ATT	ATT	GAA	GGT	CCA	GGA	TTC	ACA	GGA	GGA	AAT	TTA	CTA	TTC	CTA	AAA	1632
Ile	Ile	Glu	Gly	Pro	Gly	Phe	Thr	Gly	Gly	Asn	Leu	Leu	Phe	Leu	Lys	
	530					535				540						
GAA	TCT	AGT	AAT	TCA	ATT	GCT	AAA	TTT	AAA	GTT	ACA	TTA	AAT	TCA	GCA	1680
Glu	Ser	Ser	Asn	Ser	Ile	Ala	Lys	Phe	Lys	Val	Thr	Leu	Asn	Ser	Ala	
545				550				555						560		
GCC	TTG	TTA	CAA	CGA	TAT	CGT	GTA	AGA	ATA	CGC	TAT	GCT	TCT	ACC	ACT	1728

Ala	Leu	Leu	Gln	Arg	Tyr	Arg	Val	Arg	Ile	Arg	Tyr	Ala	Ser	Thr	Thr		
				565					570					575			
AAC	TTA	CGA	CTT	TTT	GTG	CAA	AAT	TCA	AAC	AAT	GAT	TTT	CTT	GTC	ATC	1776	
Asn	Leu	Arg	Leu	Phe	Val	Gln	Asn	Ser	Asn	Asn	Asp	Phe	Leu	Val	Ile		
			580					585					590				
TAC	ATT	AAT	AAA	ACT	ATG	AAT	AAA	GAT	GAT	GAT	TTA	ACA	TAT	CAA	ACA	1824	
Tyr	Ile	Asn	Lys	Thr	Met	Asn	Lys	Asp	Asp	Asp	Leu	Thr	Tyr	Gln	Thr		
			595				600					605					
TTT	GAT	CTC	GCA	ACT	ACT	AAT	TCT	AAT	ATG	GGG	TTC	TCG	GGT	GAT	AAG	1872	
Phe	Asp	Leu	Ala	Thr	Thr	Asn	Ser	Asn	Met	Gly	Phe	Ser	Gly	Asp	Lys		
	610						615					620					
AAT	GAA	CTT	ATA	ATA	GGA	GCA	GAA	TCT	TTC	GTT	TCT	AAT	GAA	AAA	ATC	1920	
Asn	Glu	Leu	Ile	Ile	Gly	Ala	Glu	Ser	Phe	Val	Ser	Asn	Glu	Lys	Ile		
625					630					635				640			
TAT	ATA	GAT	AAG	ATA	GAA	TTT	ATC	CCA	GTA	CAA	TTG	TAA				1959	
Tyr	Ile	Asp	Lys	Ile	Glu	Phe	Ile	Pro	Val	Gln	Leu						
			645					650									

(2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu	His	Asp	Thr	Ile	Lys	Val	Thr	Pro		
1				5					10					15			
Asn	Ser	Glu	Leu	Gln	Thr	Asn	His	Asn	Gln	Tyr	Pro	Leu	Ala	Asp	Asn		
			20					25					30				
Pro	Asn	Ser	Thr	Leu	Glu	Glu	Leu	Asn	Tyr	Lys	Glu	Phe	Leu	Arg	Met		
			35				40					45					
Thr	Glu	Asp	Ser	Ser	Thr	Glu	Val	Leu	Asp	Asn	Ser	Thr	Val	Lys	Asp		
		50				55					60						
Ala	Val	Gly	Thr	Gly	Ile	Ser	Val	Val	Gly	Gln	Ile	Leu	Gly	Val	Val		
65					70					75					80		
Gly	Val	Pro	Phe	Ala	Gly	Ala	Leu	Thr	Ser	Phe	Tyr	Gln	Ser	Phe	Leu		
				85					90					95			
Asn	Thr	Ile	Trp	Pro	Ser	Asp	Ala	Asp	Pro	Trp	Lys	Ala	Phe	Met	Ala		
			100					105						110			
Gln	Val	Glu	Val	Leu	Ile	Asp	Lys	Lys	Ile	Glu	Glu	Tyr	Ala	Lys	Ser		

115	120	125
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr		
130	135	140
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser		
145	150	155
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser		
	165	170
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val		
	180	185
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu		
	195	200
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Pro Glu		
	210	215
Asp Val Ala Glu Phe Ser His Arg Gln Leu Lys Leu Thr Gln Gln Tyr		
	225	230
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg		
	245	250
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu		
	260	265
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp		
	275	280
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile		
	290	295
Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro		
	305	310
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp		
	325	330
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe		
	340	345
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg		
	355	360
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp		
	370	375
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val		
	385	390
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys		
	405	410
		415

Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575
 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590
 Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605
 Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Cln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	

195	200	205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220			672
GAT GTT GCT GAA TTC TAT CGT AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Tyr Arg Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240			720
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255			768
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 265 270			816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 275 280 285			864
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300			912
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro 305 310 315 320			960
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp 325 330 335			1008
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe 340 345 350			1056
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg 355 360 365			1104
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp 370 375 380			1152
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val 385 390 395 400			1200
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys 405 410 415			1248
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln			1296

420	425	430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly 435 440 445			1344
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr 450 455 460			1392
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu 465 470 475 480			1440
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp 485 490 495			1488
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile 500 505 510			1536
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser 515 520 525			1584
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys 530 535 540			1632
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala 545 550 555 560			1680
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr 565 570 575			1728
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile 580 585 590			1776
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr 595 600 605			1824
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys 610 615 620			1872
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile 625 630 635 640			1920
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu			1959

(2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

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Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
 1             5             10             15

Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
      20             25             30

Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
      35             40             45

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50             55             60

Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65             70             75             80

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
      85             90             95

Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
      100            105            110

Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
      115            120            125

Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
      130            135            140

Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
      145            150            155            160

Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
      165            170            175

His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
      180            185            190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
      195            200            205

Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
      210            215            220

Asp Val Ala Glu Phe Tyr Arg Arg Gln Leu Lys Leu Thr Gln Gln Tyr

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225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525

Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540

Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560

Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575

Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590

Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605

Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620

Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	

GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val 65 70 75 80	240
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu 85 90 95	288
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala 100 105 110	336
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser 115 120 125	384
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr 130 135 140	432
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser 145 150 155 160	480
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser 165 170 175	528
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val 180 185 190	576
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu 195 200 205	624
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220	672
GAT GTT GCT GAA TTC TAT AAT AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Tyr Asn Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240	720
TCT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Ser Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255	768
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 265 270	816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 275 280 285	864

ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300	912
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro 305 310 315 320	960
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp 325 330 335	1008
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe 340 345 350	1056
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg 355 360 365	1104
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp 370 375 380	1152
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val 385 390 395 400	1200
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys 405 410 415	1248
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln 420 425 430	1296
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly 435 440 445	1344
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr 450 455 460	1392
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu 465 470 475 480	1440
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp 485 490 495	1488
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile 500 505 510	1536

ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC	1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile	
625 630 635 640	
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
1 5 10 15
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
20 25 30
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
35 40 45

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50 55 60
 Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65 70 75 80
 Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95
 Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110
 Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125
 Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140
 Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
 145 150 155 160
 Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190
 Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr Asn Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Ser Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe

340	345	350
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg		
355	360	365
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp		
370	375	380
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val		
385	390	395
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys		
405	410	415
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln		
420	425	430
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly		
435	440	445
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr		
450	455	460
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu		
465	470	475
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp		
485	490	495
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile		
500	505	510
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser		
515	520	525
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys		
530	535	540
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala		
545	550	555
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr		
565	570	575
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile		
580	585	590
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr		
595	600	605
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys		
610	615	620
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile		
625	630	635
		640

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
645 650

(2) INFORMATION FOR SEQ ID NO: 11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	

GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser 145 150 155 160	480
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser 165 170 175	528
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val 180 185 190	576
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu 195 200 205	624
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220	672
GAT GTT GCT GAA TTC TAT ACC AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Tyr Thr Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240	720
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255	768
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 265 270	816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 275 280 285	864
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300	912
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro 305 310 315 320	960
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp 325 330 335	1008
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe 340 345 350	1056
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg 355 360 365	1104

CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp 370 375 380	1152
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val 385 390 395 400	1200
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys 405 410 415	1248
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln 420 425 430	1296
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly 435 440 445	1344
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr 450 455 460	1392
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu 465 470 475 480	1440
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp 485 490 495	1488
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile 500 505 510	1536
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser 515 520 525	1584
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys 530 535 540	1632
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala 545 550 555 560	1680
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr 565 570 575	1728
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile 580 585 590	1776

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TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA      1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
      595                      600                      605

TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG      1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
      610                      615                      620

AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC      1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
      625                      630                      635                      640

TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA      1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
      645                      650

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(2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

```

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
  1                      5                      10                      15

Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
      20                      25                      30

Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
      35                      40                      45

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
      50                      55                      60

Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
      65                      70                      75                      80

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
      85                      90                      95

Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
      100                      105                      110

Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
      115                      120                      125

Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
      130                      135                      140

Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
      145                      150                      155                      160

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Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190
 Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr Thr Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr

36

450	455	460
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu		
465	470	475 480
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp		
	485	490 495
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile		
	500	505 510
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser		
	515	520 525
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys		
	530	535 540
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala		
545	550	555 560
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr		
	565	570 575
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile		
	580	585 590
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr		
	595	600 605
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys		
610	615	620
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile		
625	630	635 640
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu		
	645	650

(2) INFORMATION FOR SEQ ID NO: 13:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1959 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION:1..1956

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT

48

Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu	His	Asp	Thr	Ile	Lys	Val	Thr	Pro	
1				5					10					15		
AAC	AGT	GAA	TTG	CAA	ACT	AAC	CAT	AAT	CAA	TAT	CCT	TTA	GCT	GAC	AAT	96
Asn	Ser	Glu	Leu	Gln	Thr	Asn	His	Asn	Gln	Tyr	Pro	Leu	Ala	Asp	Asn	
			20					25					30			
CCA	AAT	TCA	ACA	CTA	GAA	GAA	TTA	AAT	TAT	AAA	GAA	TTT	TTA	AGA	ATG	144
Pro	Asn	Ser	Thr	Leu	Glu	Glu	Leu	Asn	Tyr	Lys	Glu	Phe	Leu	Arg	Met	
			35					40					45			
ACT	GAA	GAC	AGT	TCT	ACG	GAA	GTG	CTA	GAC	AAC	TCT	ACA	GTA	AAA	GAT	192
Thr	Glu	Asp	Ser	Ser	Thr	Glu	Val	Leu	Asp	Asn	Ser	Thr	Val	Lys	Asp	
	50						55				60					
GCA	GTT	GGG	ACA	GGA	ATT	TCT	GTT	GTA	GGG	CAG	ATT	TTA	GGT	GTT	GTA	240
Ala	Val	Gly	Thr	Gly	Ile	Ser	Val	Val	Gly	Gln	Ile	Leu	Gly	Val	Val	
65					70					75					80	
GGA	GTT	CCA	TTT	GCT	GGG	GCA	CTC	ACT	TCA	TTT	TAT	CAA	TCA	TTT	CTT	288
Gly	Val	Pro	Phe	Ala	Gly	Ala	Leu	Thr	Ser	Phe	Tyr	Gln	Ser	Phe	Leu	
				85					90					95		
AAC	ACT	ATA	TGG	CCA	AGT	GAT	GCT	GAC	CCA	TGG	AAG	GCT	TTT	ATG	GCA	336
Asn	Thr	Ile	Trp	Pro	Ser	Asp	Ala	Asp	Pro	Trp	Lys	Ala	Phe	Met	Ala	
			100					105					110			
CAA	GTT	GAA	GTA	CTG	ATA	GAT	AAG	AAA	ATA	GAG	GAG	TAT	GCT	AAA	AGT	384
Gln	Val	Glu	Val	Leu	Ile	Asp	Lys	Lys	Ile	Glu	Glu	Tyr	Ala	Lys	Ser	
		115					120					125				
AAA	GCT	CTT	GCA	GAG	TTA	CAG	GGT	CTT	CAA	AAT	AAT	TTC	GAA	GAT	TAT	432
Lys	Ala	Leu	Ala	Glu	Leu	Gln	Gly	Leu	Gln	Asn	Asn	Phe	Glu	Asp	Tyr	
	130					135					140					
GTT	AAT	GCG	TTA	AAT	TCC	TGG	AAG	AAA	ACA	CCT	TTA	AGT	TTG	CGA	AGT	480
Val	Asn	Ala	Leu	Asn	Ser	Trp	Lys	Lys	Thr	Pro	Leu	Ser	Leu	Arg	Ser	
145					150					155					160	
AAA	AGA	AGC	CAA	GAT	CGA	ATA	AGG	GAA	CTT	TTT	TCT	CAA	GCA	GAA	AGT	528
Lys	Arg	Ser	Gln	Asp	Arg	Ile	Arg	Glu	Leu	Phe	Ser	Gln	Ala	Glu	Ser	
				165					170					175		
CAT	TTT	CGT	AAT	TCC	ATG	CCG	TCA	TTT	GCA	GTT	TCC	AAA	TTC	GAA	GTG	576
His	Phe	Arg	Asn	Ser	Met	Pro	Ser	Phe	Ala	Val	Ser	Lys	Phe	Glu	Val	
			180					185					190			
CTG	TTT	CTA	CCA	ACA	TAT	GCA	CAA	GCT	GCA	AAT	ACA	CAT	TTA	TTG	CTA	624
Leu	Phe	Leu	Pro	Thr	Tyr	Ala	Gln	Ala	Ala	Asn	Thr	His	Leu	Leu	Leu	
		195					200					205				
TTA	AAA	GAT	GCT	CAA	GTT	TTT	GGA	GAA	GAA	TGG	GGA	TAT	TCT	TCA	GAA	672
Leu	Lys	Asp	Ala	Gln	Val	Phe	Gly	Glu	Glu	Trp	Gly	Tyr	Ser	Ser	Glu	
	210					215				220						
GAT	GTT	GCT	GAA	TTT	TAT	CAT	AGA	CAA	TTA	AAA	CTT	ACA	CAA	CAA	TAC	720

Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	
225 230 235 240	
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA	768
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	
245 250 255	
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA	816
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	
260 265 270	
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT	864
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	
275 280 285	
ATT AAT TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT	912
Ile Asn Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	
290 295 300	
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA	960
Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro	
305 310 315 320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT	1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	
325 330 335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT	1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe	
340 345 350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA	1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg	
355 360 365	
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT	1152
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp	
370 375 380	
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT	1200
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val	
385 390 395 400	
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG	1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys	
405 410 415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA	1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln	
420 425 430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC	1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly	
435 440 445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA	1392

His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr	
450 455 460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA	1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu	
465 470 475 480	
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG	1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	
485 490 495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT	1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	
500 505 510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC	1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile	
625 630 635 640	
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 14:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

40

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

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Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
 1             5             10             15

Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
 20             25             30

Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
 35             40             45

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50             55             60

Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65             70             75             80

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85             90             95

Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
100            105            110

Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
115            120            125

Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
130            135            140

Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
145            150            155            160

Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
165            170            175

His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
180            185            190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
195            200            205

Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
210            215            220

Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
225            230            235            240

Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
245            250            255

Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
260            265            270

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Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Asn Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr

42

	565		570		575
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile					
	580		585		590
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr					
	595		600		605
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys					
	610		615		620
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile					
	625		630		635
					640
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu					
	645		650		

(2) INFORMATION FOR SEQ ID NO: 15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	

85	90	95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala 100 105 110			336
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser 115 120 125			384
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr 130 135 140			432
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser 145 150 155 160			480
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser 165 170 175			528
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val 180 185 190			576
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu 195 200 205			624
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220			672
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240			720
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255			768
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 265 270			816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 275 280 285			864
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300			912
TTT ACG GAT CCA ATT TTT TTA CTT ACT ACG CTT CAG AAG TAC GGA CCA Phe Thr Asp Pro Ile Phe Leu Leu Thr Thr Leu Gln Lys Tyr Gly Pro			960

305	310	315	320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT				1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp				
325		330	335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT				1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe				
340	345		350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA				1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg				
355	360		365	
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT				1152
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp				
370	375		380	
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT				1200
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val				
385	390		395	400
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG				1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys				
405	410		415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA				1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln				
420	425		430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC				1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly				
435	440		445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA				1392
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr				
450	455		460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA				1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu				
465	470		475	480
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG				1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp				
485	490		495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT				1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile				
500	505		510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC				1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser				
515	520		525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA				1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys				

530	535	540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA			1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala			
545	550	555	560
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT			1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr			
	565	570	575
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC			1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile			
	580	585	590
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA			1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr			
	595	600	605
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG			1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys			
	610	615	620
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC			1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile			
	625	630	635
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA			1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu			
	645	650	

(2) INFORMATION FOR SEQ ID NO: 16:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro			
1	5	10	15
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn			
	20	25	30
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met			
	35	40	45
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp			
	50	55	60
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val			
	65	70	75
			80

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95

Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110

Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125

Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140

Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
 145 150 155 160

Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175

His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205

Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220

Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240

Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255

Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270

Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285

Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300

Phe Thr Asp Pro Ile Phe Leu Leu Thr Thr Leu Gln Lys Tyr Gly Pro
 305 310 315 320

Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335

Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350

Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365

Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380

Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575
 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590
 Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605
 Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 17:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1959 base pairs

(B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	

CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	
195 200 205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA	672
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	
210 215 220	
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC	720
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	
225 230 235 240	
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA	768
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	
245 250 255	
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA	816
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	
260 265 270	
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT	864
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	
275 280 285	
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT	912
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	
290 295 300	
TTT ACG GAT CCA ATT TTT ACC CTT AAT ACA CTA CAG AAG TGC GGA CCA	960
Phe Thr Asp Pro Ile Phe Thr Leu Asn Thr Leu Gln Lys Cys Gly Pro	
305 310 315 320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT	1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	
325 330 335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT	1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe	
340 345 350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA	1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg	
355 360 365	
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT	1152
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp	
370 375 380	
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT	1200
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val	
385 390 395 400	

TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys 405 410 415	1248
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln 420 425 430	1296
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly 435 440 445	1344
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr 450 455 460	1392
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu 465 470 475 480	1440
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp 485 490 495	1488
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile 500 505 510	1536
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser 515 520 525	1584
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys 530 535 540	1632
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala 545 550 555 560	1680
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr 565 570 575	1728
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile 580 585 590	1776
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr 595 600 605	1824
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys 610 615 620	1872

AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC 1920
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640

TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA 1959
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 18:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
 1 5 10 15
 Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
 20 25 30
 Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
 35 40 45
 Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50 55 60
 Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65 70 75 80
 Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95
 Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110
 Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125
 Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140
 Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
 145 150 155 160
 Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205

Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220

Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240

Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255

Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270

Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285

Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300

Phe Thr Asp Pro Ile Phe Thr Leu Asn Thr Leu Gln Lys Cys Gly Pro
 305 310 315 320

Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335

Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350

Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365

Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380

Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400

Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415

Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430

Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445

His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460

Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480

Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495

Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575
 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590
 Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605
 Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 19:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	

CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	
195 200 205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA	672
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	
210 215 220	
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC	720
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	
225 230 235 240	
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA	768
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	
245 250 255	

GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA	816
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	
260 265 270	
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT	864
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	
275 280 285	
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT	912
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	
290 295 300	
TTT ACG GAT CCA ATT TTT GCC GTT AAT ACT CTG TGG GAA TAC GGA CCA	960
Phe Thr Asp Pro Ile Phe Ala Val Asn Thr Leu Trp Glu Tyr Gly Pro	
305 310 315 320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT	1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	
325 330 335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT	1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe	
340 345 350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA	1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg	
355 360 365	
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT	1152
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp	
370 375 380	
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT	1200
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val	
385 390 395 400	
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG	1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys	
405 410 415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA	1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln	
420 425 430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC	1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly	
435 440 445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA	1392
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr	
450 455 460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA	1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu	
465 470 475 480	

56

TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG	1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	
485 490 495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT	1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	
500 505 510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC	1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile	
625 630 635 640	
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 20:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro

57

1	5	10	15
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	20	25	30
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	35	40	45
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	50	55	60
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	65	70	75
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	85	90	95
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	100	105	110
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	115	120	125
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	130	135	140
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	145	150	155
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	165	170	175
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	180	185	190
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	195	200	205
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	210	215	220
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	225	230	235
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	245	250	255
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	260	265	270
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	275	280	285
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	290	295	300

Phe Thr Asp Pro Ile Phe Ala Val Asn Thr Leu Trp Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575
 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590
 Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605

Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620

Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 21:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384

Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	
195 200 205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA	672
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	
210 215 220	
GAT GTT GCT GAA TTC TAT CGT AGA CAA TTA AAA CTT ACA CAA CAA TAC	720
Asp Val Ala Glu Phe Tyr Arg Arg Gln Leu Lys Leu Thr Gln Gln Tyr	
225 230 235 240	
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA	768
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	
245 250 255	
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA	816
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	
260 265 270	
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT	864
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	
275 280 285	
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT	912
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	
290 295 300	
TTT ACG GAT CCA ATT TTT TTA CTT ACT ACG CTT CAG AAG TAC GGA CCA	960
Phe Thr Asp Pro Ile Phe Leu Leu Thr Thr Leu Gln Lys Tyr Gly Pro	
305 310 315 320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT	1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	
325 330 335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT	1056

Tyr	Leu	Gln	Gly	Ile	Glu	Phe	His	Thr	Arg	Leu	Gln	Pro	Gly	Tyr	Phe	
			340					345					350			
GGG	AAA	GAT	TCT	TTC	AAT	TAT	TGG	TCT	GGT	AAT	TAT	GTA	GAA	ACT	AGA	1104
Gly	Lys	Asp	Ser	Phe	Asn	Tyr	Trp	Ser	Gly	Asn	Tyr	Val	Glu	Thr	Arg	
		355					360					365				
CCT	AGT	ATA	GGA	TCT	AGT	AAG	ACA	ATT	ACT	TCC	CCA	TTT	TAT	GGA	GAT	1152
Pro	Ser	Ile	Gly	Ser	Ser	Lys	Thr	Ile	Thr	Ser	Pro	Phe	Tyr	Gly	Asp	
	370					375					380					
AAA	TCT	ACT	GAA	CCT	GTA	CAA	AAG	CTA	AGC	TTT	GAT	GGA	CAA	AAA	GTT	1200
Lys	Ser	Thr	Glu	Pro	Val	Gln	Lys	Leu	Ser	Phe	Asp	Gly	Gln	Lys	Val	
385					390				395						400	
TAT	CGA	ACT	ATA	GCT	AAT	ACA	GAC	GTA	GCG	GCT	TGG	CCG	AAT	GGT	AAG	1248
Tyr	Arg	Thr	Ile	Ala	Asn	Thr	Asp	Val	Ala	Ala	Trp	Pro	Asn	Gly	Lys	
			405					410						415		
GTA	TAT	TTA	GGT	GTT	ACG	AAA	GTT	GAT	TTT	AGT	CAA	TAT	GAT	GAT	CAA	1296
Val	Tyr	Leu	Gly	Val	Thr	Lys	Val	Asp	Phe	Ser	Gln	Tyr	Asp	Asp	Gln	
		420					425						430			
AAA	AAT	GAA	ACT	AGT	ACA	CAA	ACA	TAT	GAT	TCA	AAA	AGA	AAC	AAT	GGC	1344
Lys	Asn	Glu	Thr	Ser	Thr	Gln	Thr	Tyr	Asp	Ser	Lys	Arg	Asn	Asn	Gly	
	435					440						445				
CAT	GTA	AGT	GCA	CAG	GAT	TCT	ATT	GAC	CAA	TTA	CCG	CCA	GAA	ACA	ACA	1392
His	Val	Ser	Ala	Gln	Asp	Ser	Ile	Asp	Gln	Leu	Pro	Pro	Glu	Thr	Thr	
	450					455					460					
GAT	GAA	CCA	CTT	GAA	AAA	GCA	TAT	AGT	CAT	CAG	CTT	AAT	TAC	GCG	GAA	1440
Asp	Glu	Pro	Leu	Glu	Lys	Ala	Tyr	Ser	His	Gln	Leu	Asn	Tyr	Ala	Glu	
465					470					475					480	
TGT	TTC	TTA	ATG	CAG	GAC	CGT	CGT	GGA	ACA	ATT	CCA	TTT	TTT	ACT	TGG	1488
Cys	Phe	Leu	Met	Gln	Asp	Arg	Arg	Gly	Thr	Ile	Pro	Phe	Phe	Thr	Trp	
			485					490						495		
ACA	CAT	AGA	AGT	GTA	GAC	TTT	TTT	AAT	ACA	ATT	GAT	GCT	GAA	AAG	ATT	1536
Thr	His	Arg	Ser	Val	Asp	Phe	Phe	Asn	Thr	Ile	Asp	Ala	Glu	Lys	Ile	
		500						505					510			
ACT	CAA	CTT	CCA	GTA	GTG	AAA	GCA	TAT	GCC	TTG	TCT	TCA	GGT	GCT	TCC	1584
Thr	Gln	Leu	Pro	Val	Val	Lys	Ala	Tyr	Ala	Leu	Ser	Ser	Gly	Ala	Ser	
		515					520					525				
ATT	ATT	GAA	GGT	CCA	GGA	TTC	ACA	GGA	GGA	AAT	TTA	CTA	TTC	CTA	AAA	1632
Ile	Ile	Glu	Gly	Pro	Gly	Phe	Thr	Gly	Gly	Asn	Leu	Leu	Phe	Leu	Lys	
	530					535					540					
GAA	TCT	AGT	AAT	TCA	ATT	GCT	AAA	TTT	AAA	GTT	ACA	TTA	AAT	TCA	GCA	1680
Glu	Ser	Ser	Asn	Ser	Ile	Ala	Lys	Phe	Lys	Val	Thr	Leu	Asn	Ser	Ala	
545					550					555					560	
GCC	TTG	TTA	CAA	CGA	TAT	CGT	GTA	AGA	ATA	CGC	TAT	GCT	TCT	ACC	ACT	1728

62

Ala	Leu	Leu	Gln	Arg	Tyr	Arg	Val	Arg	Ile	Arg	Tyr	Ala	Ser	Thr	Thr		
				565					570					575			
AAC	TTA	CGA	CTT	TTT	GTG	CAA	AAT	TCA	AAC	AAT	GAT	TTT	CTT	GTC	ATC	1776	
Asn	Leu	Arg	Leu	Phe	Val	Gln	Asn	Ser	Asn	Asn	Asp	Phe	Leu	Val	Ile		
			580					585					590				
TAC	ATT	AAT	AAA	ACT	ATG	AAT	AAA	GAT	GAT	GAT	TTA	ACA	TAT	CAA	ACA	1824	
Tyr	Ile	Asn	Lys	Thr	Met	Asn	Lys	Asp	Asp	Asp	Leu	Thr	Tyr	Gln	Thr		
			595				600					605					
TTT	GAT	CTC	GCA	ACT	ACT	AAT	TCT	AAT	ATG	GGG	TTC	TCG	GGT	GAT	AAG	1872	
Phe	Asp	Leu	Ala	Thr	Thr	Asn	Ser	Asn	Met	Gly	Phe	Ser	Gly	Asp	Lys		
	610						615				620						
AAT	GAA	CTT	ATA	ATA	GGA	GCA	GAA	TCT	TTC	GTT	TCT	AAT	GAA	AAA	ATC	1920	
Asn	Glu	Leu	Ile	Ile	Gly	Ala	Glu	Ser	Phe	Val	Ser	Asn	Glu	Lys	Ile		
	625				630					635					640		
TAT	ATA	GAT	AAG	ATA	GAA	TTT	ATC	CCA	GTA	CAA	TTG	TAA				1959	
Tyr	Ile	Asp	Lys	Ile	Glu	Phe	Ile	Pro	Val	Gln	Leu						
				645					650								

(2) INFORMATION FOR SEQ ID NO: 22:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22:

Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu	His	Asp	Thr	Ile	Lys	Val	Thr	Pro		
1				5					10					15			
Asn	Ser	Glu	Leu	Gln	Thr	Asn	His	Asn	Gln	Tyr	Pro	Leu	Ala	Asp	Asn		
			20					25					30				
Pro	Asn	Ser	Thr	Leu	Glu	Glu	Leu	Asn	Tyr	Lys	Glu	Phe	Leu	Arg	Met		
		35					40					45					
Thr	Glu	Asp	Ser	Ser	Thr	Glu	Val	Leu	Asp	Asn	Ser	Thr	Val	Lys	Asp		
		50				55					60						
Ala	Val	Gly	Thr	Gly	Ile	Ser	Val	Val	Gly	Gln	Ile	Leu	Gly	Val	Val		
	65				70					75				80			
Gly	Val	Pro	Phe	Ala	Gly	Ala	Leu	Thr	Ser	Phe	Tyr	Gln	Ser	Phe	Leu		
				85				90						95			
Asn	Thr	Ile	Trp	Pro	Ser	Asp	Ala	Asp	Pro	Trp	Lys	Ala	Phe	Met	Ala		
		100					105						110				
Gln	Val	Glu	Val	Leu	Ile	Asp	Lys	Lys	Ile	Glu	Glu	Tyr	Ala	Lys	Ser		

115	120	125
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr		
130	135	140
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser		
145	150	155
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser		
	165	170
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val		
	180	185
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu		
	195	200
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu		
	210	215
Asp Val Ala Glu Phe Tyr Arg Arg Gln Leu Lys Leu Thr Gln Gln Tyr		
225	230	235
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg		
	245	250
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu		
	260	265
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp		
	275	280
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile		
	290	295
Phe Thr Asp Pro Ile Phe Leu Leu Thr Thr Leu Gln Lys Tyr Gly Pro		
305	310	315
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp		
	325	330
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe		
	340	345
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg		
	355	360
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp		
	370	375
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val		
385	390	395
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys		
	405	410
		415

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Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
      420                      425                      430

Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
      435                      440                      445

His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
      450                      455                      460

Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
465                      470                      475                      480

Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
      485                      490                      495

Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
      500                      505                      510

Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
      515                      520                      525

Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
530                      535                      540

Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
545                      550                      555                      560

Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
      565                      570                      575

Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
580                      585                      590

Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
595                      600                      605

Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
610                      615                      620

Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
625                      630                      635                      640

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
      645                      650

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(2) INFORMATION FOR SEQ ID NO: 23:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 23:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	

195	200	205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220			672
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240			720
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255			768
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 265 270			816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 275 280 285			864
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300			912
TTT ACG GAT CCA ATT TTT ACG CCA ACC ACC CTA CAG GAT TAC GGA CCA Phe Thr Asp Pro Ile Phe Thr Pro Thr Thr Leu Gln Asp Tyr Gly Pro 305 310 315 320			960
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp 325 330 335			1008
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe 340 345 350			1056
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg 355 360 365			1104
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp 370 375 380			1152
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val 385 390 395 400			1200
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys 405 410 415			1248
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln			1296

420	425	430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly 435 440 445			1344
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr 450 455 460			1392
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu 465 470 475 480			1440
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp 485 490 495			1488
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile 500 505 510			1536
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser 515 520 525			1584
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys 530 535 540			1632
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala 545 550 555 560			1680
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr 565 570 575			1728
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile 580 585 590			1776
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr 595 600 605			1824
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys 610 615 620			1872
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile 625 630 635 640			1920
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu			1959

68

645

650

(2) INFORMATION FOR SEQ ID NO: 24:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 24:

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Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
 1             5             10             15

Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
      20             25             30

Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
      35             40             45

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50             55             60

Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65             70             75             80

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
      85             90             95

Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
      100            105            110

Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
      115            120            125

Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
      130            135            140

Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
      145            150            155            160

Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
      165            170            175

His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
      180            185            190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
      195            200            205

Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
      210            215            220

Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr

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225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Thr Pro Thr Thr Leu Gln Asp Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525

Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540

Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560

Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575

Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590

Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605

Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620

Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 25:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 25:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	

GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	
195 200 205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA	672
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	
210 215 220	
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC	720
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	
225 230 235 240	
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA	768
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	
245 250 255	
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA	816
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	
260 265 270	
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT	864
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	
275 280 285	

ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT	912
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	
290 295 300	
TTT ACG GAT CCA ATT TTT GCC CTG AAT ACC TTA GAC GAG TAC GGA CCA	960
Phe Thr Asp Pro Ile Phe Ala Leu Asn Thr Leu Asp Glu Tyr Gly Pro	
305 310 315 320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT	1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	
325 330 335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT	1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe	
340 345 350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA	1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg	
355 360 365	
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT	1152
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp	
370 375 380	
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT	1200
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val	
385 390 395 400	
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG	1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys	
405 410 415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA	1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln	
420 425 430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC	1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly	
435 440 445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA	1392
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr	
450 455 460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA	1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu	
465 470 475 480	
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG	1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	
485 490 495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT	1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	
500 505 510	

ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC	1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile	
625 630 635 640	
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 26:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 26:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50 55 60
 Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65 70 75 80
 Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95
 Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110
 Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125
 Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140
 Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
 145 150 155 160
 Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190
 Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ala Leu Asn Thr Leu Asp Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe

340	345	350
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg		
355	360	365
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp		
370	375	380
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val		
385	390	395
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys		
405	410	415
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln		
420	425	430
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly		
435	440	445
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr		
450	455	460
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu		
465	470	475
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp		
485	490	495
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile		
500	505	510
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser		
515	520	525
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys		
530	535	540
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala		
545	550	555
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr		
565	570	575
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile		
580	585	590
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr		
595	600	605
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys		
610	615	620
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile		
625	630	635
		640

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
645 650

(2) INFORMATION FOR SEQ ID NO: 27:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 27:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	

GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	
195 200 205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA	672
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	
210 215 220	
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC	720
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	
225 230 235 240	
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA	768
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	
245 250 255	
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA	816
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	
260 265 270	
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAC GAT	864
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	
275 280 285	
ACT AGG CGA TTC AGA AAG GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT	912
Thr Arg Arg Phe Arg Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	
290 295 300	
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA	960
Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro	
305 310 315 320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT	1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	
325 330 335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT	1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe	
340 345 350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA	1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg	
355 360 365	

CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp 370 375 380	1152
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val 385 390 395 400	1200
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys 405 410 415	1248
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln 420 425 430	1296
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly 435 440 445	1344
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr 450 455 460	1392
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu 465 470 475 480	1440
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp 485 490 495	1488
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile 500 505 510	1536
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser 515 520 525	1584
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys 530 535 540	1632
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala 545 550 555 560	1680
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr 565 570 575	1728
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile 580 585 590	1776

TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC	1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile	
625 630 635 640	
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 28:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 28:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	

Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190
 Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Thr Arg Arg Phe Arg Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr

81

450		455		460
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu				
465		470		475 480
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp				
	485		490	495
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile				
	500		505	510
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser				
	515		520	525
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys				
	530		535	540
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala				
545		550		555 560
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr				
	565		570	575
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile				
	580		585	590
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr				
	595		600	605
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys				
	610		615	620
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile				
625		630		635 640
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu				
	645		650	

(2) INFORMATION FOR SEQ ID NO: 29:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 29:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT

48

Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu	His	Asp	Thr	Ile	Lys	Val	Thr	Pro	
1				5					10					15		
AAC	AGT	GAA	TTG	CAA	ACT	AAC	CAT	AAT	CAA	TAT	CCT	TTA	GCT	GAC	AAT	96
Asn	Ser	Glu	Leu	Gln	Thr	Asn	His	Asn	Gln	Tyr	Pro	Leu	Ala	Asp	Asn	
			20					25					30			
CCA	AAT	TCA	ACA	CTA	GAA	GAA	TTA	AAT	TAT	AAA	GAA	TTT	TTA	AGA	ATG	144
Pro	Asn	Ser	Thr	Leu	Glu	Glu	Leu	Asn	Tyr	Lys	Glu	Phe	Leu	Arg	Met	
			35				40					45				
ACT	GAA	GAC	AGT	TCT	ACG	GAA	GTG	CTA	GAC	AAC	TCT	ACA	GTA	AAA	GAT	192
Thr	Glu	Asp	Ser	Ser	Thr	Glu	Val	Leu	Asp	Asn	Ser	Thr	Val	Lys	Asp	
	50					55					60					
GCA	GTT	GGG	ACA	GGA	ATT	TCT	GTT	GTA	GGG	CAG	ATT	TTA	GGT	GTT	GTA	240
Ala	Val	Gly	Thr	Gly	Ile	Ser	Val	Val	Gly	Gln	Ile	Leu	Gly	Val	Val	
	65				70				75						80	
GGA	GTT	CCA	TTT	GCT	GGG	GCA	CTC	ACT	TCA	TTT	TAT	CAA	TCA	TTT	CTT	288
Gly	Val	Pro	Phe	Ala	Gly	Ala	Leu	Thr	Ser	Phe	Tyr	Gln	Ser	Phe	Leu	
				85				90						95		
AAC	ACT	ATA	TGG	CCA	AGT	GAT	GCT	GAC	CCA	TGG	AAG	GCT	TTT	ATG	GCA	336
Asn	Thr	Ile	Trp	Pro	Ser	Asp	Ala	Asp	Pro	Trp	Lys	Ala	Phe	Met	Ala	
			100					105					110			
CAA	GTT	GAA	GTA	CTG	ATA	GAT	AAG	AAA	ATA	GAG	GAG	TAT	GCT	AAA	AGT	384
Gln	Val	Glu	Val	Leu	Ile	Asp	Lys	Lys	Ile	Glu	Glu	Tyr	Ala	Lys	Ser	
		115					120					125				
AAA	GCT	CTT	GCA	GAG	TTA	CAG	GGT	CTT	CAA	AAT	AAT	TTC	GAA	GAT	TAT	432
Lys	Ala	Leu	Ala	Glu	Leu	Gln	Gly	Leu	Gln	Asn	Asn	Phe	Glu	Asp	Tyr	
	130					135				140						
GTT	AAT	GCG	TTA	AAT	TCC	TGG	AAG	AAA	ACA	CCT	TTA	AGT	TTG	CGA	AGT	480
Val	Asn	Ala	Leu	Asn	Ser	Trp	Lys	Lys	Thr	Pro	Leu	Ser	Leu	Arg	Ser	
145					150					155					160	
AAA	AGA	AGC	CAA	GAT	CGA	ATA	AGG	GAA	CTT	TTT	TCT	CAA	GCA	GAA	AGT	528
Lys	Arg	Ser	Gln	Asp	Arg	Ile	Arg	Glu	Leu	Phe	Ser	Gln	Ala	Glu	Ser	
				165				170					175			
CAT	TTT	CGT	AAT	TCC	ATG	CCG	TCA	TTT	GCA	GTT	TCC	AAA	TTC	GAA	GTG	576
His	Phe	Arg	Asn	Ser	Met	Pro	Ser	Phe	Ala	Val	Ser	Lys	Phe	Glu	Val	
			180					185					190			
CTG	TTT	CTA	CCA	ACA	TAT	GCA	CAA	GCT	GCA	AAT	ACA	CAT	TTA	TTG	CTA	624
Leu	Phe	Leu	Pro	Thr	Tyr	Ala	Gln	Ala	Ala	Asn	Thr	His	Leu	Leu	Leu	
		195					200					205				
TTA	AAA	GAT	GCT	CAA	GTT	TTT	GGA	GAA	GAA	TGG	GGA	TAT	TCT	TCA	GAA	672
Leu	Lys	Asp	Ala	Gln	Val	Phe	Gly	Glu	Glu	Trp	Gly	Tyr	Ser	Ser	Glu	
	210					215				220						
GAT	GTT	GCT	GAA	TTC	TAT	CGT	AGA	CAA	TTA	AAA	CTT	ACA	CAA	CAA	TAC	720

Asp	Val	Ala	Glu	Phe	Tyr	Arg	Arg	Gln	Leu	Lys	Leu	Thr	Gln	Gln	Tyr	
225					230					235					240	
ACT	GAC	CAT	TGT	GTT	AAT	TGG	TAT	AAT	GTT	GGA	TTA	AAT	GGT	TTA	AGA	768
Thr	Asp	His	Cys	Val	Asn	Trp	Tyr	Asn	Val	Gly	Leu	Asn	Gly	Leu	Arg	
				245					250					255		
GGT	TCA	ACT	TAT	GAT	GCA	TGG	GTC	AAA	TTT	AAC	CGT	TTT	CGC	AGA	GAA	816
Gly	Ser	Thr	Tyr	Asp	Ala	Trp	Val	Lys	Phe	Asn	Arg	Phe	Arg	Arg	Glu	
			260					265					270			
ATG	ACT	TTA	ACT	GTA	TTA	GAT	CTA	ATT	GTA	CTT	TTC	CCA	TTT	TAT	GAT	864
Met	Thr	Leu	Thr	Val	Leu	Asp	Leu	Ile	Val	Leu	Phe	Pro	Phe	Tyr	Asp	
		275					280					285				
ATT	CGG	TTA	TAC	TCA	AAA	GGG	GTT	AAA	ACA	GAA	CTA	ACA	AGA	GAC	ATT	912
Ile	Arg	Leu	Tyr	Ser	Lys	Gly	Val	Lys	Thr	Glu	Leu	Thr	Arg	Asp	Ile	
	290					295					300					
TTT	ACG	GAT	CCA	ATT	TTT	TTA	CTT	AAT	ACT	CTT	CAG	GAG	TAT	GGA	CCA	960
Phe	Thr	Asp	Pro	Ile	Phe	Leu	Leu	Asn	Thr	Leu	Gln	Glu	Tyr	Gly	Pro	
305					310					315					320	
ACT	TTT	TTG	AGT	ATA	GAA	AAC	TCT	ATT	CGA	AAA	CCT	CAT	TTA	TTT	GAT	1008
Thr	Phe	Leu	Ser	Ile	Glu	Asn	Ser	Ile	Arg	Lys	Pro	His	Leu	Phe	Asp	
				325					330					335		
TAT	TTA	CAG	GGG	ATT	GAA	TTT	CAT	ACG	CGT	CTT	CAA	CCT	GGT	TAC	TTT	1056
Tyr	Leu	Gln	Gly	Ile	Glu	Phe	His	Thr	Arg	Leu	Gln	Pro	Gly	Tyr	Phe	
		340						345					350			
GGG	AAA	GAT	TCT	TTC	AAT	TAT	TGG	TCT	GGT	AAT	TAT	GTA	GAA	ACT	AGA	1104
Gly	Lys	Asp	Ser	Phe	Asn	Tyr	Trp	Ser	Gly	Asn	Tyr	Val	Glu	Thr	Arg	
		355					360					365				
CCT	AGT	ATA	GGA	TCT	AGT	AAG	ACA	ATT	ACT	TCC	CCA	TTT	TAT	GGA	GAT	1152
Pro	Ser	Ile	Gly	Ser	Ser	Lys	Thr	Ile	Thr	Ser	Pro	Phe	Tyr	Gly	Asp	
	370					375					380					
AAA	TCT	ACT	GAA	CCT	GTA	CAA	AAG	CTA	AGC	TTT	GAT	GGA	CAA	AAA	GTT	1200
Lys	Ser	Thr	Glu	Pro	Val	Gln	Lys	Leu	Ser	Phe	Asp	Gly	Gln	Lys	Val	
385					390					395					400	
TAT	CGA	ACT	ATA	GCT	AAT	ACA	GAC	GTA	GCG	GCT	TGG	CCG	AAT	GGT	AAG	1248
Tyr	Arg	Thr	Ile	Ala	Asn	Thr	Asp	Val	Ala	Ala	Trp	Pro	Asn	Gly	Lys	
				405					410					415		
GTA	TAT	TTA	GGT	GTT	ACG	AAA	GTT	GAT	TTT	AGT	CAA	TAT	GAT	GAT	CAA	1296
Val	Tyr	Leu	Gly	Val	Thr	Lys	Val	Asp	Phe	Ser	Gln	Tyr	Asp	Asp	Gln	
			420					425					430			
AAA	AAT	GAA	ACT	AGT	ACA	CAA	ACA	TAT	GAT	TCA	AAA	AGA	AAC	AAT	GGC	1344
Lys	Asn	Glu	Thr	Ser	Thr	Gln	Thr	Tyr	Asp	Ser	Lys	Arg	Asn	Asn	Gly	
		435					440					445				
CAT	GTA	AGT	GCA	CAG	GAT	TCT	ATT	GAC	CAA	TTA	CCG	CCA	GAA	ACA	ACA	1392

His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr	
450 455 460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA	1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu	
465 470 475 480	
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG	1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	
485 490 495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT	1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	
500 505 510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC	1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile	
625 630 635 640	
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 30:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

85

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 30:

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Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
 1             5             10             15

Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
      20             25             30

Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
      35             40             45

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
      50             55             60

Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
      65             70             75             80

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
      85             90             95

Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
      100            105            110

Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
      115            120            125

Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
      130            135            140

Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
      145            150            155            160

Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
      165            170            175

His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
      180            185            190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
      195            200            205

Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
      210            215            220

Asp Val Ala Glu Phe Tyr Arg Arg Gln Leu Lys Leu Thr Gln Gln Tyr
      225            230            235            240

Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
      245            250            255

Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
      260            265            270

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Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Leu Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr

87

	565		570		575
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile					
	580		585		590
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr					
	595		600		605
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys					
	610		615		620
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile					
	625		630		635
					640
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu					
	645		650		

(2) INFORMATION FOR SEQ ID NO: 31:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 31:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	

88

85	90	95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala 100 105 110			336
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser 115 120 125			384
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr 130 135 140			432
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser 145 150 155 160			480
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser 165 170 175			528
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val 180 185 190			576
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu 195 200 205			624
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220			672
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240			720
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255			768
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 265 270			816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 275 280 285			864
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300			912
TTT ACG GAT CCA ATT TTT ATC CTC AAT ACG CTA CAG GAG TAC GGA CCA Phe Thr Asp Pro Ile Phe Ile Leu Asn Thr Leu Gln Glu Tyr Gly Pro			960

305	310	315	320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT				1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp				
	325	330	335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT				1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe				
	340	345	350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA				1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg				
	355	360	365	
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT				1152
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp				
	370	375	380	
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT				1200
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val				
	385	390	395	400
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG				1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys				
	405	410	415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA				1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln				
	420	425	430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC				1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly				
	435	440	445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA				1392
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr				
	450	455	460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA				1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu				
	465	470	475	480
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG				1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp				
	485	490	495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT				1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile				
	500	505	510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC				1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser				
	515	520	525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA				1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys				

530	535	540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA			1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala			
545	550	555	560
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT			1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr			
	565	570	575
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC			1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile			
	580	585	590
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA			1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr			
	595	600	605
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG			1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys			
	610	615	620
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC			1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile			
	625	630	635
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA			1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu			
	645	650	

(2) INFORMATION FOR SEQ ID NO: 32:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 32:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro			
1	5	10	15
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn			
	20	25	30
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met			
	35	40	45
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp			
	50	55	60
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val			
	65	70	75
			80

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95
 Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110
 Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125
 Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140
 Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
 145 150 155 160
 Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190
 Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ile Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380

Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575
 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590
 Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605
 Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 33:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1959 base pairs

(B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 33:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	

CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	
195 200 205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA	672
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	
210 215 220	
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC	720
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	
225 230 235 240	
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA	768
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	
245 250 255	
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA	816
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	
260 265 270	
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT	864
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	
275 280 285	
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT	912
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	
290 295 300	
TTT ACG GAT CCA ATT TTT ATC CTA CAT ACG CTG CAG GAG TAC GGA CCA	960
Phe Thr Asp Pro Ile Phe Ile Leu His Thr Leu Gln Glu Tyr Gly Pro	
305 310 315 320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT	1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	
325 330 335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT	1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe	
340 345 350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA	1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg	
355 360 365	
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT	1152
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp	
370 375 380	
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT	1200
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val	
385 390 395 400	

TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG	1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys	
405 410 415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA	1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln	
420 425 430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC	1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly	
435 440 445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA	1392
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr	
450 455 460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA	1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu	
465 470 475 480	
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG	1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	
485 490 495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT	1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	
500 505 510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	

AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC 1920
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640

TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA 1959
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 34:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 652 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 34:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
 1 5 10 15
 Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
 20 25 30
 Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
 35 40 45
 Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50 55 60
 Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65 70 75 80
 Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95
 Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110
 Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125
 Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140
 Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
 145 150 155 160
 Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205

Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220

Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240

Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255

Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270

Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285

Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300

Phe Thr Asp Pro Ile Phe Ile Leu His Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320

Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335

Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350

Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365

Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380

Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp, Gly Gln Lys Val
 385 390 395 400

Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415

Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430

Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445

His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460

Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480

Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495

Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575
 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590
 Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605
 Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 35:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 35:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	

CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	
195 200 205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA	672
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	
210 215 220	
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC	720
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	
225 230 235 240	
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA	768
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	
245 250 255	

GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA	816
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	
260 265 270	
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT	864
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	
275 280 285	
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT	912
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	
290 295 300	
TTT ACG GAT CCA ATT TTT TCC CTC GTT AAC CTA ATG GTG TAC GGA CCA	960
Phe Thr Asp Pro Ile Phe Ser Leu Val Asn Leu Met Val Tyr Gly Pro	
305 310 315 320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT	1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	
325 330 335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT	1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe	
340 345 350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA	1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg	
355 360 365	
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT	1152
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp	
370 375 380	
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT	1200
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val	
385 390 395 400	
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG	1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys	
405 410 415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA	1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln	
420 425 430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC	1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly	
435 440 445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA	1392
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr	
450 455 460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA	1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu	
465 470 475 480	

TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG	1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	
485 490 495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT	1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	
500 505 510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC	1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile	
625 630 635 640	
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 36:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 36:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro

102

1	5	10	15
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	20	25	30
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	35	40	45
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	50	55	60
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	65	70	75
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	85	90	95
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	100	105	110
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	115	120	125
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	130	135	140
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	145	150	155
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	165	170	175
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	180	185	190
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	195	200	205
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	210	215	220
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	225	230	235
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	245	250	255
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	260	265	270
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	275	280	285
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	290	295	300

Phe Thr Asp Pro Ile Phe Ser Leu Val Asn Leu Met Val Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575
 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590
 Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605

Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620

Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 37:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 37:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384

Gln	Val	Glu	Val	Leu	Ile	Asp	Lys	Lys	Ile	Glu	Glu	Tyr	Ala	Lys	Ser	
	115						120					125				
AAA	GCT	CTT	GCA	GAG	TTA	CAG	GGT	CTT	CAA	AAT	AAT	TTC	GAA	GAT	TAT	432
Lys	Ala	Leu	Ala	Glu	Leu	Gln	Gly	Leu	Gln	Asn	Asn	Phe	Glu	Asp	Tyr	
	130					135					140					
GTT	AAT	GCG	TTA	AAT	TCC	TGG	AAG	AAA	ACA	CCT	TTA	AGT	TTG	CGA	AGT	480
Val	Asn	Ala	Leu	Asn	Ser	Trp	Lys	Lys	Thr	Pro	Leu	Ser	Leu	Arg	Ser	
145					150					155					160	
AAA	AGA	AGC	CAA	GAT	CGA	ATA	AGG	GAA	CTT	TTT	TCT	CAA	GCA	GAA	AGT	528
Lys	Arg	Ser	Gln	Asp	Arg	Ile	Arg	Glu	Leu	Phe	Ser	Gln	Ala	Glu	Ser	
				165					170					175		
CAT	TTT	CGT	AAT	TCC	ATG	CCG	TCA	TTT	GCA	GTT	TCC	AAA	TTC	GAA	GTG	576
His	Phe	Arg	Asn	Ser	Met	Pro	Ser	Phe	Ala	Val	Ser	Lys	Phe	Glu	Val	
			180					185					190			
CTG	TTT	CTA	CCA	ACA	TAT	GCA	CAA	GCT	GCA	AAT	ACA	CAT	TTA	TTG	CTA	624
Leu	Phe	Leu	Pro	Thr	Tyr	Ala	Gln	Ala	Ala	Asn	Thr	His	Leu	Leu	Leu	
	195						200					205				
TTA	AAA	GAT	GCT	CAA	GTT	TTT	GGA	GAA	GAA	TGG	GGA	TAT	TCT	TCA	GAA	672
Leu	Lys	Asp	Ala	Gln	Val	Phe	Gly	Glu	Glu	Trp	Gly	Tyr	Ser	Ser	Glu	
	210					215					220					
GAT	GTT	GCT	GAA	TTT	TAT	CAT	AGA	CAA	TTA	AAA	CTT	ACA	CAA	CAA	TAC	720
Asp	Val	Ala	Glu	Phe	Tyr	His	Arg	Gln	Leu	Lys	Leu	Thr	Gln	Gln	Tyr	
225					230					235					240	
ACT	GAC	CAT	TGT	GTT	AAT	TGG	TAT	AAT	GTT	GGA	TTA	AAT	GGT	TTA	AGA	768
Thr	Asp	His	Cys	Val	Asn	Trp	Tyr	Asn	Val	Gly	Leu	Asn	Gly	Leu	Arg	
			245					250					255			
GGT	TCA	ACT	TAT	GAT	GCA	TGG	GTC	AAA	TTT	AAC	CGT	TTT	CGC	AGA	GAA	816
Gly	Ser	Thr	Tyr	Asp	Ala	Trp	Val	Lys	Phe	Asn	Arg	Phe	Arg	Arg	Glu	
		260						265					270			
ATG	ACT	TTA	ACT	GTA	TTA	GAT	CTA	ATT	GTA	CTT	TTC	CCA	TTT	TAT	GAT	864
Met	Thr	Leu	Thr	Val	Leu	Asp	Leu	Ile	Val	Leu	Phe	Pro	Phe	Tyr	Asp	
		275					280					285				
ATT	CGG	TTA	TAC	TCA	AAA	GGG	GTT	AAA	ACA	GAA	CTA	ACA	AGA	GAC	ATT	912
Ile	Arg	Leu	Tyr	Ser	Lys	Gly	Val	Lys	Thr	Glu	Leu	Thr	Arg	Asp	Ile	
	290					295					300					
TTT	ACG	GAT	CCA	ATT	TTT	TCT	CTT	AGG	ACA	CCA	CTT	GCG	TAC	GGA	CCA	960
Phe	Thr	Asp	Pro	Ile	Phe	Ser	Leu	Arg	Thr	Pro	Leu	Ala	Tyr	Gly	Pro	
305				310						315					320	
ACT	TTT	TTG	AGT	ATA	GAA	AAC	TCT	ATT	CGA	AAA	CCT	CAT	TTA	TTT	GAT	1008
Thr	Phe	Leu	Ser	Ile	Glu	Asn	Ser	Ile	Arg	Lys	Pro	His	Leu	Phe	Asp	
				325					330				335			
TAT	TTA	CAG	GGG	ATT	GAA	TTT	CAT	ACG	CGT	CTT	CAA	CCT	GGT	TAC	TTT	1056

Tyr	Leu	Gln	Gly	Ile	Glu	Phe	His	Thr	Arg	Leu	Gln	Pro	Gly	Tyr	Phe	
			340					345					350			
GGG	AAA	GAT	TCT	TTC	AAT	TAT	TGG	TCT	GGT	AAT	TAT	GTA	GAA	ACT	AGA	1104
Gly	Lys	Asp	Ser	Phe	Asn	Tyr	Trp	Ser	Gly	Asn	Tyr	Val	Glu	Thr	Arg	
		355					360					365				
CCT	AGT	ATA	GGA	TCT	AGT	AAG	ACA	ATT	ACT	TCC	CCA	TTT	TAT	GGA	GAT	1152
Pro	Ser	Ile	Gly	Ser	Ser	Lys	Thr	Ile	Thr	Ser	Pro	Phe	Tyr	Gly	Asp	
		370				375					380					
AAA	TCT	ACT	GAA	CCT	GTA	CAA	AAG	CTA	AGC	TTT	GAT	GGA	CAA	AAA	GTT	1200
Lys	Ser	Thr	Glu	Pro	Val	Gln	Lys	Leu	Ser	Phe	Asp	Gly	Gln	Lys	Val	
385					390				395					400		
TAT	CGA	ACT	ATA	GCT	AAT	ACA	GAC	GTA	GCG	GCT	TGG	CCG	AAT	GGT	AAG	1248
Tyr	Arg	Thr	Ile	Ala	Asn	Thr	Asp	Val	Ala	Ala	Trp	Pro	Asn	Gly	Lys	
				405				410						415		
GTA	TAT	TTA	GGT	GTT	ACG	AAA	GTT	GAT	TTT	AGT	CAA	TAT	GAT	GAT	CAA	1296
Val	Tyr	Leu	Gly	Val	Thr	Lys	Val	Asp	Phe	Ser	Gln	Tyr	Asp	Asp	Gln	
			420					425					430			
AAA	AAT	GAA	ACT	AGT	ACA	CAA	ACA	TAT	GAT	TCA	AAA	AGA	AAC	AAT	GGC	1344
Lys	Asn	Glu	Thr	Ser	Thr	Gln	Thr	Tyr	Asp	Ser	Lys	Arg	Asn	Asn	Gly	
		435					440					445				
CAT	GTA	AGT	GCA	CAG	GAT	TCT	ATT	GAC	CAA	TTA	CCG	CCA	GAA	ACA	ACA	1392
His	Val	Ser	Ala	Gln	Asp	Ser	Ile	Asp	Gln	Leu	Pro	Pro	Glu	Thr	Thr	
		450				455					460					
GAT	GAA	CCA	CTT	GAA	AAA	GCA	TAT	AGT	CAT	CAG	CTT	AAT	TAC	GCG	GAA	1440
Asp	Glu	Pro	Leu	Glu	Lys	Ala	Tyr	Ser	His	Gln	Leu	Asn	Tyr	Ala	Glu	
465					470					475				480		
TGT	TTC	TTA	ATG	CAG	GAC	CGT	CGT	GGA	ACA	ATT	CCA	TTT	TTT	ACT	TGG	1488
Cys	Phe	Leu	Met	Gln	Asp	Arg	Arg	Gly	Thr	Ile	Pro	Phe	Phe	Thr	Trp	
				485				490						495		
ACA	CAT	AGA	AGT	GTA	GAC	TTT	TTT	AAT	ACA	ATT	GAT	GCT	GAA	AAG	ATT	1536
Thr	His	Arg	Ser	Val	Asp	Phe	Phe	Asn	Thr	Ile	Asp	Ala	Glu	Lys	Ile	
			500					505					510			
ACT	CAA	CTT	CCA	GTA	GTG	AAA	GCA	TAT	GCC	TTG	TCT	TCA	GGT	GCT	TCC	1584
Thr	Gln	Leu	Pro	Val	Val	Lys	Ala	Tyr	Ala	Leu	Ser	Ser	Gly	Ala	Ser	
		515					520					525				
ATT	ATT	GAA	GGT	CCA	GGA	TTC	ACA	GGA	GGA	AAT	TTA	CTA	TTC	CTA	AAA	1632
Ile	Ile	Glu	Gly	Pro	Gly	Phe	Thr	Gly	Gly	Asn	Leu	Leu	Phe	Leu	Lys	
		530				535					540					
GAA	TCT	AGT	AAT	TCA	ATT	GCT	AAA	TTT	AAA	GTT	ACA	TTA	AAT	TCA	GCA	1680
Glu	Ser	Ser	Asn	Ser	Ile	Ala	Lys	Phe	Lys	Val	Thr	Leu	Asn	Ser	Ala	
545					550				555					560		
GCC	TTG	TTA	CAA	CGA	TAT	CGT	GTA	AGA	ATA	CGC	TAT	GCT	TCT	ACC	ACT	1728

107

Ala	Leu	Leu	Gln	Arg	Tyr	Arg	Val	Arg	Ile	Arg	Tyr	Ala	Ser	Thr	Thr		
				565					570					575			
AAC	TTA	CGA	CTT	TTT	GTG	CAA	AAT	TCA	AAC	AAT	GAT	TTT	CTT	GTC	ATC	1776	
Asn	Leu	Arg	Leu	Phe	Val	Gln	Asn	Ser	Asn	Asn	Asp	Phe	Leu	Val	Ile		
			580					585					590				
TAC	ATT	AAT	AAA	ACT	ATG	AAT	AAA	GAT	GAT	GAT	TTA	ACA	TAT	CAA	ACA	1824	
Tyr	Ile	Asn	Lys	Thr	Met	Asn	Lys	Asp	Asp	Asp	Leu	Thr	Tyr	Gln	Thr		
			595				600					605					
TTT	GAT	CTC	GCA	ACT	ACT	AAT	TCT	AAT	ATG	GGG	TTC	TCG	GGT	GAT	AAG	1872	
Phe	Asp	Leu	Ala	Thr	Thr	Asn	Ser	Asn	Met	Gly	Phe	Ser	Gly	Asp	Lys		
			610				615				620						
AAT	GAA	CTT	ATA	ATA	GGA	GCA	GAA	TCT	TTC	GTT	TCT	AAT	GAA	AAA	ATC	1920	
Asn	Glu	Leu	Ile	Ile	Gly	Ala	Glu	Ser	Phe	Val	Ser	Asn	Glu	Lys	Ile		
			625			630				635					640		
TAT	ATA	GAT	AAG	ATA	GAA	TTT	ATC	CCA	GTA	CAA	TTG	TAA				1959	
Tyr	Ile	Asp	Lys	Ile	Glu	Phe	Ile	Pro	Val	Gln	Leu						
				645					650								

(2) INFORMATION FOR SEQ ID NO: 38:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 38:

Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu	His	Asp	Thr	Ile	Lys	Val	Thr	Pro		
1				5					10					15			
Asn	Ser	Glu	Leu	Gln	Thr	Asn	His	Asn	Gln	Tyr	Pro	Leu	Ala	Asp	Asn		
			20					25					30				
Pro	Asn	Ser	Thr	Leu	Glu	Glu	Leu	Asn	Tyr	Lys	Glu	Phe	Leu	Arg	Met		
			35				40					45					
Thr	Glu	Asp	Ser	Ser	Thr	Glu	Val	Leu	Asp	Asn	Ser	Thr	Val	Lys	Asp		
			50			55					60						
Ala	Val	Gly	Thr	Gly	Ile	Ser	Val	Val	Gly	Gln	Ile	Leu	Gly	Val	Val		
			65			70				75				80			
Gly	Val	Pro	Phe	Ala	Gly	Ala	Leu	Thr	Ser	Phe	Tyr	Gln	Ser	Phe	Leu		
				85				90						95			
Asn	Thr	Ile	Trp	Pro	Ser	Asp	Ala	Asp	Pro	Trp	Lys	Ala	Phe	Met	Ala		
			100					105					110				
Gln	Val	Glu	Val	Leu	Ile	Asp	Lys	Lys	Ile	Glu	Glu	Tyr	Ala	Lys	Ser		

115	120	125
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr		
130	135	140
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser		
145	150	155
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser		
165	170	175
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val		
180	185	190
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu		
195	200	205
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu		
210	215	220
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr		
225	230	235
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg		
245	250	255
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu		
260	265	270
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp		
275	280	285
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile		
290	295	300
Phe Thr Asp Pro Ile Phe Ser Leu Arg Thr Pro Leu Ala Tyr Gly Pro		
305	310	315
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp		
325	330	335
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe		
340	345	350
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg		
355	360	365
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp		
370	375	380
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val		
385	390	395
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys		
405	410	415

Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575
 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590
 Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605
 Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 39:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 39:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	

195	200	205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220			672
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240			720
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255			768
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 265 270			816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TTC AAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Phe Asn 275 280 285			864
ATT TTG CTT TAC AGT AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Ile Leu Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300			912
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro 305 310 315 320			960
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp 325 330 335			1008
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe 340 345 350			1056
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg 355 360 365			1104
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp 370 375 380			1152
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val 385 390 395 400			1200
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys 405 410 415			1248
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln			1296

420	425	430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly 435 440 445			1344
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr 450 455 460			1392
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu 465 470 475 480			1440
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp 485 490 495			1488
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile 500 505 510			1536
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser 515 520 525			1584
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys 530 535 540			1632
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala 545 550 555 560			1680
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr 565 570 575			1728
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile 580 585 590			1776
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr 595 600 605			1824
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys 610 615 620			1872
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile 625 630 635 640			1920
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu			1959

645

650

(2) INFORMATION FOR SEQ ID NO: 40:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 40:

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Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
 1             5             10             15

Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
 20             25             30

Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
 35             40             45

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50             55             60

Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65             70             75             80

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85             90             95

Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
100            105            110

Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
115            120            125

Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
130            135            140

Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
145            150            155            160

Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
165            170            175

His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
180            185            190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
195            200            205

Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
210            215            220

Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr

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225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Phe Asn
 275 280 285
 Ile Leu Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525

115

Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540

Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560

Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575

Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590

Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605

Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620

Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 41:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 41:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	

GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val 65 70 75 80	240
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu 85 90 95	288
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala 100 105 110	336
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser 115 120 125	384
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr 130 135 140	432
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser 145 150 155 160	480
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser 165 170 175	528
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val 180 185 190	576
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu 195 200 205	624
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220	672
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240	720
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255	768
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 265 270	816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 275 280 285	864

ATT GTG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Ile Val Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300	912
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro 305 310 315 320	960
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp 325 330 335	1008
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe 340 345 350	1056
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg 355 360 365	1104
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp 370 375 380	1152
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val 385 390 395 400	1200
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys 405 410 415	1248
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln 420 425 430	1296
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly 435 440 445	1344
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr 450 455 460	1392
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu 465 470 475 480	1440
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp 485 490 495	1488
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile 500 505 510	1536

ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC	1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile	
625 630 635 640	
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 42:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 42:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50 55 60
 Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65 70 75 80
 Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95
 Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110
 Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125
 Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140
 Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
 145 150 155 160
 Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190
 Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Val Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe

340	345	350
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg		
355	360	365
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp		
370	375	380
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val		
385	390	395
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys		
405	410	415
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln		
420	425	430
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly		
435	440	445
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr		
450	455	460
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu		
465	470	475
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp		
485	490	495
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile		
500	505	510
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser		
515	520	525
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys		
530	535	540
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala		
545	550	555
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr		
565	570	575
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile		
580	585	590
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr		
595	600	605
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys		
610	615	620
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile		
625	630	635
		640

121

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
645 650

(2) INFORMATION FOR SEQ ID NO: 43:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 43:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	

GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser 145 150 155 160	480
AAA AGA AGC CAA GGT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT Lys Arg Ser Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser 165 170 175	528
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val 180 185 190	576
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu 195 200 205	624
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220	672
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240	720
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255	768
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 265 270	816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 275 280 285	864
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300	912
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro 305 310 315 320	960
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp 325 330 335	1008
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe 340 345 350	1056
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg 355 360 365	1104

CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp 370 375 380	1152
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val 385 390 395 400	1200
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys 405 410 415	1248
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln 420 425 430	1296
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly 435 440 445	1344
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr 450 455 460	1392
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu 465 470 475 480	1440
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp 485 490 495	1488
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile 500 505 510	1536
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser 515 520 525	1584
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys 530 535 540	1632
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala 545 550 555 560	1680
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr 565 570 575	1728
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile 580 585 590	1776

124

TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC	1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile	
625 630 635 640	
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 44:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 44:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	

Lys Arg Ser Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190
 Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr

126

450	455	460
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu		
465	470	475 480
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp		
	485 490	495
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile		
	500 505	510
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser		
	515 520	525
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys		
	530 535	540
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala		
	545 550	555 560
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr		
	565 570	575
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile		
	580 585	590
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr		
	595 600	605
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys		
	610 615	620
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile		
	625 630	635 640
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu		
	645	650

(2) INFORMATION FOR SEQ ID NO: 45:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 1959 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 45:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT

48

127

Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu	His	Asp	Thr	Ile	Lys	Val	Thr	Pro	
1				5					10					15		
AAC	AGT	GAA	TTG	CAA	ACT	AAC	CAT	AAT	CAA	TAT	CCT	TTA	GCT	GAC	AAT	96
Asn	Ser	Glu	Leu	Gln	Thr	Asn	His	Asn	Gln	Tyr	Pro	Leu	Ala	Asp	Asn	
			20					25					30			
CCA	AAT	TCA	ACA	CTA	GAA	GAA	TTA	AAT	TAT	AAA	GAA	TTT	TTA	AGA	ATG	144
Pro	Asn	Ser	Thr	Leu	Glu	Glu	Leu	Asn	Tyr	Lys	Glu	Phe	Leu	Arg	Met	
			35				40					45				
ACT	GAA	GAC	AGT	TCT	ACG	GAA	GTG	CTA	GAC	AAC	TCT	ACA	GTA	AAA	GAT	192
Thr	Glu	Asp	Ser	Ser	Thr	Glu	Val	Leu	Asp	Asn	Ser	Thr	Val	Lys	Asp	
	50					55					60					
GCA	GTT	GGG	ACA	GGA	ATT	TCT	GTT	GTA	GGG	CAG	ATT	TTA	GGT	GTT	GTA	240
Ala	Val	Gly	Thr	Gly	Ile	Ser	Val	Val	Gly	Gln	Ile	Leu	Gly	Val	Val	
65					70				75						80	
GGA	GTT	CCA	TTT	GCT	GGG	GCA	CTC	ACT	TCA	TTT	TAT	CAA	TCA	TTT	CTT	288
Gly	Val	Pro	Phe	Ala	Gly	Ala	Leu	Thr	Ser	Phe	Tyr	Gln	Ser	Phe	Leu	
				85					90					95		
AAC	ACT	ATA	TGG	CCA	AGT	GAT	GCT	GAC	CCA	TGG	AAG	GCT	TTT	ATG	GCA	336
Asn	Thr	Ile	Trp	Pro	Ser	Asp	Ala	Asp	Pro	Trp	Lys	Ala	Phe	Met	Ala	
			100					105					110			
CAA	GTT	GAA	GTA	CTG	ATA	GAT	AAG	AAA	ATA	GAG	GAG	TAT	GCT	AAA	AGT	384
Gln	Val	Glu	Val	Leu	Ile	Asp	Lys	Lys	Ile	Glu	Glu	Tyr	Ala	Lys	Ser	
		115					120					125				
AAA	GCT	CTT	GCA	GAG	TTA	CAG	GGT	CTT	CAA	AAT	AAT	TTC	GAA	GAT	TAT	432
Lys	Ala	Leu	Ala	Glu	Leu	Gln	Gly	Leu	Gln	Asn	Asn	Phe	Glu	Asp	Tyr	
	130					135					140					
GTT	AAT	GCG	TTA	AAT	TCC	TGG	AAG	AAA	ACA	CCT	TTA	AGT	TTG	CGA	AAT	480
Val	Asn	Ala	Leu	Asn	Ser	Trp	Lys	Lys	Thr	Pro	Leu	Ser	Leu	Arg	Asn	
145					150					155					160	
CCA	CAC	AGC	CAA	GGT	CGA	ATA	AGG	GAA	CTT	TTT	TCT	CAA	GCA	GAA	AGT	528
Pro	His	Ser	Gln	Gly	Arg	Ile	Arg	Glu	Leu	Phe	Ser	Gln	Ala	Glu	Ser	
				165					170					175		
CAT	TTT	CGT	AAT	TCC	ATG	CCG	TCA	TTT	GCA	GTT	TCC	AAA	TTC	GAA	GTG	576
His	Phe	Arg	Asn	Ser	Met	Pro	Ser	Phe	Ala	Val	Ser	Lys	Phe	Glu	Val	
			180					185					190			
CTG	TTT	CTA	CCA	ACA	TAT	GCA	CAA	GCT	GCA	AAT	ACA	CAT	TTA	TTG	CTA	624
Leu	Phe	Leu	Pro	Thr	Tyr	Ala	Gln	Ala	Ala	Asn	Thr	His	Leu	Leu	Leu	
		195					200					205				
TTA	AAA	GAT	GCT	CAA	GTT	TTT	GGA	GAA	GAA	TGG	GGA	TAT	TCT	TCA	GAA	672
Leu	Lys	Asp	Ala	Gln	Val	Phe	Gly	Glu	Glu	Trp	Gly	Tyr	Ser	Ser	Glu	
	210					215					220					
GAT	GTT	GCT	GAA	TTT	TAT	CAT	AGA	CAA	TTA	AAA	CTT	ACA	CAA	CAA	TAC	720

Asp	Val	Ala	Glu	Phe	Tyr	His	Arg	Gln	Leu	Lys	Leu	Thr	Gln	Gln	Tyr	
225					230					235					240	
ACT	GAC	CAT	TGT	GTT	AAT	TGG	TAT	AAT	GTT	GGA	TTA	AAT	GGT	TTA	AGA	768
Thr	Asp	His	Cys	Val	Asn	Trp	Tyr	Asn	Val	Gly	Leu	Asn	Gly	Leu	Arg	
				245					250					255		
GGT	TCA	ACT	TAT	GAT	GCA	TGG	GTC	AAA	TTT	AAC	CGT	TTT	CGC	AGA	GAA	816
Gly	Ser	Thr	Tyr	Asp	Ala	Trp	Val	Lys	Phe	Asn	Arg	Phe	Arg	Arg	Glu	
			260					265				270				
ATG	ACT	TTA	ACT	GTA	TTA	GAT	CTA	ATT	GTA	CTT	TTC	CCA	TTT	TAT	GAT	864
Met	Thr	Leu	Thr	Val	Leu	Asp	Leu	Ile	Val	Leu	Phe	Pro	Phe	Tyr	Asp	
		275					280					285				
ATT	CGG	TTA	TAC	TCA	AAA	GGG	GTT	AAA	ACA	GAA	CTA	ACA	AGA	GAC	ATT	912
Ile	Arg	Leu	Tyr	Ser	Lys	Gly	Val	Lys	Thr	Glu	Leu	Thr	Arg	Asp	Ile	
	290					295					300					
TTT	ACG	GAT	CCA	ATT	TTT	TCA	CTT	AAT	ACT	CTT	CAG	GAG	TAT	GGA	CCA	960
Phe	Thr	Asp	Pro	Ile	Phe	Ser	Leu	Asn	Thr	Leu	Gln	Glu	Tyr	Gly	Pro	
305				310						315					320	
ACT	TTT	TTG	AGT	ATA	GAA	AAC	TCT	ATT	CGA	AAA	CCT	CAT	TTA	TTT	GAT	1008
Thr	Phe	Leu	Ser	Ile	Glu	Asn	Ser	Ile	Arg	Lys	Pro	His	Leu	Phe	Asp	
				325					330				335			
TAT	TTA	CAG	GGG	ATT	GAA	TTT	CAT	ACG	CGT	CTT	CAA	CCT	GGT	TAC	TTT	1056
Tyr	Leu	Gln	Gly	Ile	Glu	Phe	His	Thr	Arg	Leu	Gln	Pro	Gly	Tyr	Phe	
		340						345				350				
GGG	AAA	GAT	TCT	TTC	AAT	TAT	TGG	TCT	GGT	AAT	TAT	GTA	GAA	ACT	AGA	1104
Gly	Lys	Asp	Ser	Phe	Asn	Tyr	Trp	Ser	Gly	Asn	Tyr	Val	Glu	Thr	Arg	
	355						360					365				
CCT	AGT	ATA	GGA	TCT	AGT	AAG	ACA	ATT	ACT	TCC	CCA	TTT	TAT	GGA	GAT	1152
Pro	Ser	Ile	Gly	Ser	Ser	Lys	Thr	Ile	Thr	Ser	Pro	Phe	Tyr	Gly	Asp	
	370					375					380					
AAA	TCT	ACT	GAA	CCT	GTA	CAA	AAG	CTA	AGC	TTT	GAT	GGA	CAA	AAA	GTT	1200
Lys	Ser	Thr	Glu	Pro	Val	Gln	Lys	Leu	Ser	Phe	Asp	Gly	Gln	Lys	Val	
385				390					395					400		
TAT	CGA	ACT	ATA	GCT	AAT	ACA	GAC	GTA	GCG	GCT	TGG	CCG	AAT	GGT	AAG	1248
Tyr	Arg	Thr	Ile	Ala	Asn	Thr	Asp	Val	Ala	Ala	Trp	Pro	Asn	Gly	Lys	
			405					410					415			
GTA	TAT	TTA	GGT	GTT	ACG	AAA	GTT	GAT	TTT	AGT	CAA	TAT	GAT	GAT	CAA	1296
Val	Tyr	Leu	Gly	Val	Thr	Lys	Val	Asp	Phe	Ser	Gln	Tyr	Asp	Asp	Gln	
		420					425				430					
AAA	AAT	GAA	ACT	AGT	ACA	CAA	ACA	TAT	GAT	TCA	AAA	AGA	AAC	AAT	GGC	1344
Lys	Asn	Glu	Thr	Ser	Thr	Gln	Thr	Tyr	Asp	Ser	Lys	Arg	Asn	Asn	Gly	
	435					440					445					
CAT	GTA	AGT	GCA	CAG	GAT	TCT	ATT	GAC	CAA	TTA	CCG	CCA	GAA	ACA	ACA	1392

His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr	
450 455 460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA	1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu	
465 470 475 480	
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG	1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	
485 490 495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT	1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	
500 505 510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC	1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile	
625 630 635 640	
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 46:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

130

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 46:

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Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
 1             5             10             15

Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
      20             25             30

Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
      35             40             45

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
      50             55             60

Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
      65             70             75             80

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
      85             90             95

Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
      100            105            110

Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
      115            120            125

Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
      130            135            140

Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Asn
      145            150            155            160

Pro His Ser Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
      165            170            175

His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
      180            185            190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
      195            200            205

Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
      210            215            220

Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
      225            230            235            240

Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
      245            250            255

Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
      260            265            270

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Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285

Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300

Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320

Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335

Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350

Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365

Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380

Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400

Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415

Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430

Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445

His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460

Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480

Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495

Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510

Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525

Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540

Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560

Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr

132

	565		570		575
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile					
	580		585		590
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr					
	595		600		605
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys					
	610		615		620
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile					
	625		630		640
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu					
	645		650		

(2) INFORMATION FOR SEQ ID NO: 47:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 47:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	

133

85	90	95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala 100 105 110			336
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser 115 120 125			384
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr 130 135 140			432
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser 145 150 155 160			480
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser 165 170 175			528
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val 180 185 190			576
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu 195 200 205			624
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220			672
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240			720
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255			768
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 265 270			816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 275 280 285			864
GTT CGG TTA TAC CCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Val Arg Leu Tyr Pro Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300			912
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro			960

305	310	315	320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT				1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	325	330	335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT				1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe	340	345	350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA				1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg	355	360	365	
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT				1152
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp	370	375	380	
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT				1200
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val	385	390	395	400
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG				1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys	405	410	415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA				1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln	420	425	430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC				1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly	435	440	445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA				1392
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr	450	455	460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA				1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu	465	470	475	480
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG				1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	485	490	495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT				1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	500	505	510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC				1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	515	520	525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA				1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys				

530	535	540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA			1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala			
545	550	555	560
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT			1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr			
	565	570	575
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC			1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile			
	580	585	590
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA			1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr			
	595	600	605
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG			1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys			
	610	615	620
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC			1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile			
	625	630	635
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA			1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu			
	645	650	

(2) INFORMATION FOR SEQ ID NO: 48:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 48:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro			
1	5	10	15
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn			
	20	25	30
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met			
	35	40	45
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp			
	50	55	60
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val			
	65	70	75
			80

136

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95

Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110

Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125

Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140

Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
 145 150 155 160

Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175

His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205

Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220

Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240

Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255

Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270

Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285

Val Arg Leu Tyr Pro Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300

Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320

Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335

Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350

Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365

Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380

Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575
 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590
 Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605
 Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 49:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1959 base pairs

(B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 49:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AAT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Asn	
145 150 155 160	
CCA CAC AGC CAA GGT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Pro His Ser Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	

CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	
195 200 205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA	672
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	
210 215 220	
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC	720
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	
225 230 235 240	
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA	768
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	
245 250 255	
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA	816
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	
260 265 270	
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT	864
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	
275 280 285	
GTT CGG TTA TAC CCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT	912
Val Arg Leu Tyr Pro Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	
290 295 300	
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA	960
Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro	
305 310 315 320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT	1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	
325 330 335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT	1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe	
340 345 350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA	1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg	
355 360 365	
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT	1152
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp	
370 375 380	
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT	1200
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val	
385 390 395 400	

140

TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG	1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys	
405 410 415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA	1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln	
420 425 430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC	1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly	
435 440 445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA	1392
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr	
450 455 460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA	1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu	
465 470 475 480	
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG	1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	
485 490 495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT	1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	
500 505 510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	

AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC 1920
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640

TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA 1959
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 50:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 50:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
 1 5 10 15
 Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
 20 25 30
 Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
 35 40 45
 Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50 55 60
 Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65 70 75 80
 Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95
 Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110
 Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125
 Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140
 Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Asn
 145 150 155 160
 Pro His Ser Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Val Arg Leu Tyr Pro Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495


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Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
      500                      505                      510

Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
      515                      520                      525

Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
      530                      535                      540

Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
545                      550                      555                      560

Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
      565                      570                      575

Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
      580                      585                      590

Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
      595                      600                      605

Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
      610                      615                      620

Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
625                      630                      635                      640

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
      645                      650

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(2) INFORMATION FOR SEQ ID NO: 51:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1956 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1953

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 51:

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ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT      48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
  1                      5                      10                      15

AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT      96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
  20                      25                      30

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CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAA GAC CCA TGG AAG GCT TTT ATG GCA CAA	336
Asn Thr Ile Trp Pro Ser Glu Asp Pro Trp Lys Ala Phe Met Ala Gln	
100 105 110	
GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT AAA	384
Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser Lys	
115 120 125	
GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT GTT	432
Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr Val	
130 135 140	
AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT AAA	480
Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser Lys	
145 150 155 160	
AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT CAT	528
Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser His	
165 170 175	
TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG CTG	576
Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val Leu	
180 185 190	
TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA TTA	624
Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu Leu	
195 200 205	
AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA GAT	672
Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu Asp	
210 215 220	
GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC ACT	720
Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr Thr	
225 230 235 240	
GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA GGT	768
Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg Gly	
245 250 255	

TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA ATG	816
Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu Met	
260 265 270	
ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT ATT	864
Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp Ile	
275 280 285	
CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT TTT	912
Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile Phe	
290 295 300	
ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA ACT	960
Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro Thr	
305 310 315 320	
TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT TAT	1008
Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp Tyr	
325 330 335	
TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT GGG	1056
Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe Gly	
340 345 350	
AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA CCT	1104
Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg Pro	
355 360 365	
AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT AAA	1152
Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp Lys	
370 375 380	
TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT TAT	1200
Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val Tyr	
385 390 395 400	
CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG GTA	1248
Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys Val	
405 410 415	
TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA AAA	1296
Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln Lys	
420 425 430	
AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC CAT	1344
Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly His	
435 440 445	
GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA GAT	1392
Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr Asp	
450 455 460	
GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA TGT	1440
Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu Cys	
465 470 475 480	

146

TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG ACA	1488
Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp Thr	
485 490 495	
CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT ACT	1536
His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile Thr	
500 505 510	
CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC ATT	1584
Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser Ile	
515 520 525	
ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA GAA	1632
Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys Glu	
530 535 540	
TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA GCC	1680
Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala Ala	
545 550 555 560	
TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT AAC	1728
Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr Asn	
565 570 575	
TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC TAC	1776
Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile Tyr	
580 585 590	
ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA TTT	1824
Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Glu Thr Phe	
595 600 605	
GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG AAT	1872
Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys Asn	
610 615 620	
GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC TAT	1920
Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile Tyr	
625 630 635 640	
ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1956
Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 52:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 651 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 52:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro

147

1	5	10	15
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	20	25	30
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	35	40	45
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	50	55	60
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	65	70	75
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	85	90	95
Asn Thr Ile Trp Pro Ser Glu Asp Pro Trp Lys Ala Phe Met Ala Gln	100	105	110
Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser Lys	115	120	125
Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr Val	130	135	140
Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser Lys	145	150	155
Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser His	165	170	175
Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val Leu	180	185	190
Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu Leu	195	200	205
Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu Asp	210	215	220
Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr Thr	225	230	235
Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg Gly	245	250	255
Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu Met	260	265	270
Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp Ile	275	280	285
Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile Phe	290	295	300

Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro Thr
 305 310 315 320
 Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp Tyr
 325 330 335
 Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe Gly
 340 345 350
 Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg Pro
 355 360 365
 Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp Lys
 370 375 380
 Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val Tyr
 385 390 395 400
 Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys Val
 405 410 415
 Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln Lys
 420 425 430
 Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly His
 435 440 445
 Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr Asp
 450 455 460
 Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu Cys
 465 470 475 480
 Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp Thr
 485 490 495
 His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile Thr
 500 505 510
 Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser Ile
 515 520 525
 Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys Glu
 530 535 540
 Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala Ala
 545 550 555 560
 Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr Asn
 565 570 575
 Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile Tyr
 580 585 590
 Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr Phe
 595 600 605

Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys Asn
 610 615 620

Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile Tyr
 625 630 635 640

Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 53:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 53:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384

150

Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC GGA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Gly Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	
195 200 205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA	672
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	
210 215 220	
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC	720
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	
225 230 235 240	
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA	768
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	
245 250 255	
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA	816
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	
260 265 270	
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT	864
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	
275 280 285	
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT	912
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	
290 295 300	
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA	960
Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro	
305 310 315 320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT	1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	
325 330 335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT	1056

Tyr	Leu	Gln	Gly	Ile	Glu	Phe	His	Thr	Arg	Leu	Gln	Pro	Gly	Tyr	Phe	
			340					345					350			
GGG	AAA	GAT	TCT	TTC	AAT	TAT	TGG	TCT	GGT	AAT	TAT	GTA	GAA	ACT	AGA	1104
Gly	Lys	Asp	Ser	Phe	Asn	Tyr	Trp	Ser	Gly	Asn	Tyr	Val	Glu	Thr	Arg	
		355					360					365				
CCT	AGT	ATA	GGA	TCT	AGT	AAG	ACA	ATT	ACT	TCC	CCA	TTT	TAT	GGA	GAT	1152
Pro	Ser	Ile	Gly	Ser	Ser	Lys	Thr	Ile	Thr	Ser	Pro	Phe	Tyr	Gly	Asp	
		370				375					380					
AAA	TCT	ACT	GAA	CCT	GTA	CAA	AAG	CTA	AGC	TTT	GAT	GGA	CAA	AAA	GTT	1200
Lys	Ser	Thr	Glu	Pro	Val	Gln	Lys	Leu	Ser	Phe	Asp	Gly	Gln	Lys	Val	
385					390					395					400	
TAT	CGA	ACT	ATA	GCT	AAT	ACA	GAC	GTA	GCG	GCT	TGG	CCG	AAT	GGT	AAG	1248
Tyr	Arg	Thr	Ile	Ala	Asn	Thr	Asp	Val	Ala	Ala	Trp	Pro	Asn	Gly	Lys	
				405					410					415		
GTA	TAT	TTA	GGT	GTT	ACG	AAA	GTT	GAT	TTT	AGT	CAA	TAT	GAT	GAT	CAA	1296
Val	Tyr	Leu	Gly	Val	Thr	Lys	Val	Asp	Phe	Ser	Gln	Tyr	Asp	Asp	Gln	
			420					425					430			
AAA	AAT	GAA	ACT	AGT	ACA	CAA	ACA	TAT	GAT	TCA	AAA	AGA	AAC	AAT	GGC	1344
Lys	Asn	Glu	Thr	Ser	Thr	Gln	Thr	Tyr	Asp	Ser	Lys	Arg	Asn	Asn	Gly	
		435					440					445				
CAT	GTA	AGT	GCA	CAG	GAT	TCT	ATT	GAC	CAA	TTA	CCG	CCA	GAA	ACA	ACA	1392
His	Val	Ser	Ala	Gln	Asp	Ser	Ile	Asp	Gln	Leu	Pro	Pro	Glu	Thr	Thr	
		450				455					460					
GAT	GAA	CCA	CTT	GAA	AAA	GCA	TAT	AGT	CAT	CAG	CTT	AAT	TAC	GCG	GAA	1440
Asp	Glu	Pro	Leu	Glu	Lys	Ala	Tyr	Ser	His	Gln	Leu	Asn	Tyr	Ala	Glu	
465					470					475					480	
TGT	TTC	TTA	ATG	CAG	GAC	CGT	CGT	GGA	ACA	ATT	CCA	TTT	TTT	ACT	TGG	1488
Cys	Phe	Leu	Met	Gln	Asp	Arg	Arg	Gly	Thr	Ile	Pro	Phe	Phe	Thr	Trp	
				485				490						495		
ACA	CAT	AGA	AGT	GTA	GAC	TTT	TTT	AAT	ACA	ATT	GAT	GCT	GAA	AAG	ATT	1536
Thr	His	Arg	Ser	Val	Asp	Phe	Phe	Asn	Thr	Ile	Asp	Ala	Glu	Lys	Ile	
			500					505				510				
ACT	CAA	CTT	CCA	GTA	GTG	AAA	GCA	TAT	GCC	TTG	TCT	TCA	GGT	GCT	TCC	1584
Thr	Gln	Leu	Pro	Val	Val	Lys	Ala	Tyr	Ala	Leu	Ser	Ser	Gly	Ala	Ser	
		515					520					525				
ATT	ATT	GAA	GGT	CCA	GGA	TTC	ACA	GGA	GGA	AAT	TTA	CTA	TTC	CTA	AAA	1632
Ile	Ile	Glu	Gly	Pro	Gly	Phe	Thr	Gly	Gly	Asn	Leu	Leu	Phe	Leu	Lys	
		530				535					540					
GAA	TCT	AGT	AAT	TCA	ATT	GCT	AAA	TTT	AAA	GTT	ACA	TTA	AAT	TCA	GCA	1680
Glu	Ser	Ser	Asn	Ser	Ile	Ala	Lys	Phe	Lys	Val	Thr	Leu	Asn	Ser	Ala	
545					550					555					560	
GCC	TTG	TTA	CAA	CGA	TAT	CGT	GTA	AGA	ATA	CGC	TAT	GCT	TCT	ACC	ACT	1728

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Ala	Leu	Leu	Gln	Arg	Tyr	Arg	Val	Arg	Ile	Arg	Tyr	Ala	Ser	Thr	Thr		
				565					570					575			
AAC	TTA	CGA	CTT	TTT	GTG	CAA	AAT	TCA	AAC	AAT	GAT	TTT	CTT	GTC	ATC	1776	
Asn	Leu	Arg	Leu	Phe	Val	Gln	Asn	Ser	Asn	Asn	Asp	Phe	Leu	Val	Ile		
			580					585					590				
TAC	ATT	AAT	AAA	ACT	ATG	AAT	AAA	GAT	GAT	GAT	TTA	ACA	TAT	CAA	ACA	1824	
Tyr	Ile	Asn	Lys	Thr	Met	Asn	Lys	Asp	Asp	Asp	Leu	Thr	Tyr	Gln	Thr		
			595				600					605					
TTT	GAT	CTC	GCA	ACT	ACT	AAT	TCT	AAT	ATG	GGG	TTC	TCG	GGT	GAT	AAG	1872	
Phe	Asp	Leu	Ala	Thr	Thr	Asn	Ser	Asn	Met	Gly	Phe	Ser	Gly	Asp	Lys		
			610				615					620					
AAT	GAA	CTT	ATA	ATA	GGA	GCA	GAA	TCT	TTC	GTT	TCT	AAT	GAA	AAA	ATC	1920	
Asn	Glu	Leu	Ile	Ile	Gly	Ala	Glu	Ser	Phe	Val	Ser	Asn	Glu	Lys	Ile		
			625			630				635				640			
TAT	ATA	GAT	AAG	ATA	GAA	TTT	ATC	CCA	GTA	CAA	TTG	TAA				1959	
Tyr	Ile	Asp	Lys	Ile	Glu	Phe	Ile	Pro	Val	Gln	Leu						
				645				650									

(2) INFORMATION FOR SEQ ID NO: 54:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 652 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 54:

Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu	His	Asp	Thr	Ile	Lys	Val	Thr	Pro		
1				5					10					15			
Asn	Ser	Glu	Leu	Gln	Thr	Asn	His	Asn	Gln	Tyr	Pro	Leu	Ala	Asp	Asn		
			20					25					30				
Pro	Asn	Ser	Thr	Leu	Glu	Glu	Leu	Asn	Tyr	Lys	Glu	Phe	Leu	Arg	Met		
			35				40					45					
Thr	Glu	Asp	Ser	Ser	Thr	Glu	Val	Leu	Asp	Asn	Ser	Thr	Val	Lys	Asp		
			50				55				60						
Ala	Val	Gly	Thr	Gly	Ile	Ser	Val	Val	Gly	Gln	Ile	Leu	Gly	Val	Val		
			65			70				75				80			
Gly	Val	Pro	Phe	Ala	Gly	Ala	Leu	Thr	Ser	Phe	Tyr	Gln	Ser	Phe	Leu		
				85				90						95			
Asn	Thr	Ile	Trp	Pro	Ser	Asp	Ala	Asp	Pro	Trp	Lys	Ala	Phe	Met	Ala		
			100					105					110				
Gln	Val	Glu	Val	Leu	Ile	Asp	Lys	Lys	Ile	Glu	Glu	Tyr	Ala	Lys	Ser		

115	120	125
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr		
130	135	140
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser		
145	150	155
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser		
	165	170
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Gly Phe Glu Val		
	180	185
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu		
	195	200
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu		
	210	215
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr		
	225	230
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg		
	245	250
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu		
	260	265
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp		
	275	280
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile		
	290	295
Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro		
	305	310
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp		
	325	330
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe		
	340	345
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg		
	355	360
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp		
	370	375
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val		
	385	390
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys		
	405	410
		415

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Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
      420                      425                      430

Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
      435      _              440                      445 _

His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
      450                      455                      460

Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
465                      470                      475                      480

Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
      485                      490                      495

Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
      500                      505                      510

Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
      515                      520                      525

Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
      530                      535                      540

Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
545                      550                      555                      560

Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
      565                      570                      575

Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
      580                      585                      590

Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
      595                      600                      605

Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
      610                      615                      620

Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
625                      630                      635                      640

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
      645                      650

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(2) INFORMATION FOR SEQ ID NO: 55:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1956 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION:1..1953

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 55:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAA GAC CCA TGG AAG GCT TTT ATG GCA CAA	336
Asn Thr Ile Trp Pro Ser Glu Asp Pro Trp Lys Ala Phe Met Ala Gln	
100 105 110	
GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT AAA	384
Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser Lys	
115 120 125	
GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT GTT	432
Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr Val	
130 135 140	
AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AAT CCA	480
Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Asn Pro	
145 150 155 160	
CAC AGC CAA GGT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT CAT	528
His Ser Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser His	
165 170 175	
TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG CTG	576
Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val Leu	
180 185 190	
TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA TTA	624
Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu Leu	

195	200	205	
AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA GAT Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu Asp 210 215 220			672
GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC ACT Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr Thr 225 230 235 240			720
GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA GGT Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg Gly 245 250 255			768
TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA ATG Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu Met 260 265 270			816
ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT ATT Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp Ile 275 280 285			864
CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT TTT Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile Phe 290 295 300			912
ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA ACT Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro Thr 305 310 315 320			960
TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT TAT Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp Tyr 325 330 335			1008
TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT GGG Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe Gly 340 345 350			1056
AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA CCT Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg Pro 355 360 365			1104
AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT AAA Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp Lys 370 375 380			1152
TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT TAT Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val Tyr 385 390 395 400			1200
CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG GTA Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys Val 405 410 415			1248
TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA AAA Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln Lys			1296

420	425	430	
AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC CAT			1344
Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly His			
435	440	445	
GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA GAT			1392
Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr Asp			
450	455	460	
GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA TGT			1440
Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu Cys			
465	470	475	480
TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG ACA			1488
Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp Thr			
485	490	495	
CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT ACT			1536
His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile Thr			
500	505	510	
CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC ATT			1584
Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser Ile			
515	520	525	
ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA GAA			1632
Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys Glu			
530	535	540	
TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA GCC			1680
Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala Ala			
545	550	555	560
TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT AAC			1728
Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr Asn			
565	570	575	
TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC TAC			1776
Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile Tyr			
580	585	590	
ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA TTT			1824
Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr Phe			
595	600	605	
GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG AAT			1872
Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys Asn			
610	615	620	
GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC TAT			1920
Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile Tyr			
625	630	635	640
ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA			1956
Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu			

158

645

650

(2) INFORMATION FOR SEQ ID NO: 56:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 651 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 56:

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Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
 1             5             10             15

Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
      20             25             30

Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
      35             40             45

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
      50             55             60

Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
      65             70             75             80

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
      85             90             95

Asn Thr Ile Trp Pro Ser Glu Asp Pro Trp Lys Ala Phe Met Ala Gln
      100            105            110

Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser Lys
      115            120            125

Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr Val
      130            135            140

Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Asn Pro
      145            150            155            160

His Ser Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser His
      165            170            175

Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val Leu
      180            185            190

Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu Leu
      195            200            205

Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu Asp
      210            215            220

Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr Thr

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225 230 235 240
 Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg Gly
 245 250 255
 Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu Met
 260 265 270
 Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp Ile
 275 280 285
 Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile Phe
 290 295 300
 Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro Thr
 305 310 315 320
 Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp Tyr
 325 330 335
 Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe Gly
 340 345 350
 Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg Pro
 355 360 365
 Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp Lys
 370 375 380
 Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val Tyr
 385 390 395 400
 Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys Val
 405 410 415
 Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln Lys
 420 425 430
 Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly His
 435 440 445
 Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr Asp
 450 455 460
 Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu Cys
 465 470 475 480
 Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp Thr
 485 490 495
 His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile Thr
 500 505 510
 Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser Ile
 515 520 525

Ile	Glu	Gly	Pro	Gly	Phe	Thr	Gly	Gly	Asn	Leu	Leu	Phe	Leu	Lys	Glu
530						535				540					
Ser	Ser	Asn	Ser	Ile	Ala	Lys	Phe	Lys	Val	Thr	Leu	Asn	Ser	Ala	Ala
545					550					555					560
Leu	Leu	Gln	Arg	Tyr	Arg	Val	Arg	Ile	Arg	Tyr	Ala	Ser	Thr	Thr	Asn
				565					570					575	
Leu	Arg	Leu	Phe	Val	Gln	Asn	Ser	Asn	Asn	Asp	Phe	Leu	Val	Ile	Tyr
			580					585					590		
Ile	Asn	Lys	Thr	Met	Asn	Lys	Asp	Asp	Asp	Leu	Thr	Tyr	Gln	Thr	Phe
		595					600					605			
Asp	Leu	Ala	Thr	Thr	Asn	Ser	Asn	Met	Gly	Phe	Ser	Gly	Asp	Lys	Asn
	610					615					620				
Glu	Leu	Ile	Ile	Gly	Ala	Glu	Ser	Phe	Val	Ser	Asn	Glu	Lys	Ile	Tyr
625					630					635					640
Ile	Asp	Lys	Ile	Glu	Phe	Ile	Pro	Val	Gln	Leu					
				645					650						

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1956 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(A) NAME/KEY: CDS
(B) LOCATION: 1..1953

ATG	AAT	CCA	AAC	AAT	CGA	AGT	GAA	CAT	GAT	ACG	ATA	AAG	GTT	ACA	CCT	48
Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu	His	Asp	Thr	Ile	Lys	Val	Thr	Pro	
1				5				10				15				
AAC	AGT	GAA	TTG	CAA	ACT	AAC	CAT	AAT	CAA	TAT	CCT	TTA	GCT	GAC	AAT	96
Asn	Ser	Glu	Leu	Gln	Thr	Asn	His	Asn	Gln	Tyr	Pro	Leu	Ala	Asp	Asn	
20				25				30								
CCA	AAT	TCA	ACA	CTA	GAA	GAA	TTA	AAT	TAT	AAA	GAA	TTT	TTA	AGA	ATG	144
Pro	Asn	Ser	Thr	Leu	Glu	Glu	Leu	Asn	Tyr	Lys	Glu	Phe	Leu	Arg	Met	
35				40				45								
ACT	GAA	GAC	AGT	TCT	ACG	GAA	GTG	CTA	GAC	AAC	TCT	ACA	GTA	AAA	GAT	192
Thr	Glu	Asp	Ser	Ser	Thr	Glu	Val	Leu	Asp	Asn	Ser	Thr	Val	Lys	Asp	
50				55				60								

GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAA GAC CCA TGG AAG GCT TTT ATG GCA CAA	336
Asn Thr Ile Trp Pro Ser Glu Asp Pro Trp Lys Ala Phe Met Ala Gln	
100 105 110	
GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT AAA	384
Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser Lys	
115 120 125	
GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT GTT	432
Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr Val	
130 135 140	
AAT GCG TTA AAT TCC TGG AAG AAA TTT CAC CAT TCT CGT CGT TCT AAA	480
Asn Ala Leu Asn Ser Trp Lys Lys Phe His His Ser Arg Arg Ser Lys	
145 150 155 160	
AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT CAT	528
Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser His	
165 170 175	
TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG CTG	576
Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val Leu	
180 185 190	
TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA TTA	624
Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu Leu	
195 200 205	
AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA GAT	672
Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu Asp	
210 215 220	
GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC ACT	720
Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr Thr	
225 230 235 240	
GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA GGT	768
Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg Gly	
245 250 255	
TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA ATG	816
Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu Met	
260 265 270	
ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT ATT	864
Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp Ile	
275 280 285	

CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT TTT Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile Phe 290 295 300	912
ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA ACT Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro Thr 305 310 315 320	960
TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT TAT Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp Tyr 325 330 335	1008
TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT GGG Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe Gly 340 345 350	1056
AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA CCT Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg Pro 355 360 365	1104
AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT AAA Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp Lys 370 375 380	1152
TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT TAT Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val Tyr 385 390 395 400	1200
CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG GTA Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys Val 405 410 415	1248
TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA AAA Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln Lys 420 425 430	1296
AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC CAT Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly His 435 440 445	1344
GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA GAT Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr Asp 450 455 460	1392
GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA TGT Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu Cys 465 470 475 480	1440
TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG ACA Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp Thr 485 490 495	1488
CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT ACT His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile Thr 500 505 510	1536

CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC ATT	1584
Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser Ile	
515 520 525	
ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA GAA	1632
Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys Glu	
530 535 540	
TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA GCC	1680
Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala Ala	
545 550 555 560	
TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT AAC	1728
Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr Asn	
565 570 575	
TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC TAC	1776
Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile Tyr	
580 585 590	
ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA TTT	1824
Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr Phe	
595 600 605	
GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG AAT	1872
Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys Asn	
610 615 620	
GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC TAT	1920
Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile Tyr	
625 630 635 640	
ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1956
Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 58:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 651 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 58:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
1 5 10 15
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
20 25 30
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
35 40 45

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50 55 60
 Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65 70 75 80
 Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95
 Asn Thr Ile Trp Pro Ser Glu Asp Pro Trp Lys Ala Phe Met Ala Gln
 100 105 110
 Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser Lys
 115 120 125
 Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr Val
 130 135 140
 Asn Ala Leu Asn Ser Trp Lys Lys Phe His His Ser Arg Arg Ser Lys
 145 150 155 160
 Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser His
 165 170 175
 Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val Leu
 180 185 190
 Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu Leu
 195 200 205
 Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu Asp
 210 215 220
 Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr Thr
 225 230 235 240
 Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg Gly
 245 250 255
 Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu Met
 260 265 270
 Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp Ile
 275 280 285
 Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile Phe
 290 295 300
 Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro Thr
 305 310 315 320
 Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp Tyr
 325 330 335
 Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe Gly

340	345	350
Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg Pro		
355	360	365
Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp Lys		
370	375	380
Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val Tyr		
385	390	400
Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys Val		
405	410	415
Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln Lys		
420	425	430
Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly His		
435	440	445
Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr Asp		
450	455	460
Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu Cys		
465	470	475
Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp Thr		
485	490	495
His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile Thr		
500	505	510
Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser Ile		
515	520	525
Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys Glu		
530	535	540
Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala Ala		
545	550	555
Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr Asn		
565	570	575
Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile Tyr		
580	585	590
Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr Phe		
595	600	605
Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys Asn		
610	615	620
Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile Tyr		
625	630	635
		640

Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
645 650

(2) INFORMATION FOR SEQ ID NO: 59:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 59:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	

GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser 145 150 155 160	480
AAA AGA AGC CAA GGT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT Lys Arg Ser Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser 165 170 175	528
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val 180 185 190	576
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu 195 200 205	624
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220	672
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240	720
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255	768
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 265 270	816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 275 280 285	864
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300	912
TTT ACG GAT CCA ATT TTT ACC CTT AAT ACA CTA CAG AAG TAC GGA CCA Phe Thr Asp Pro Ile Phe Thr Leu Asn Thr Leu Gln Lys Tyr Gly Pro 305 310 315 320	960
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp 325 330 335	1008
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe 340 345 350	1056
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg 355 360 365	1104

CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp 370 375 380	1152
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val 385 390 395 400	1200
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys 405 410 415	1248
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln 420 425 430	1296
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly 435 440 445	1344
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr 450 455 460	1392
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu 465 470 475 480	1440
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp 485 490 495	1488
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile 500 505 510	1536
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser 515 520 525	1584
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys 530 535 540	1632
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala 545 550 555 560	1680
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr 565 570 575	1728
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile 580 585 590	1776

TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC	1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile	
625 630 635 640	
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 60:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 60:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	

170

Lys Arg Ser Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190
 Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Thr Leu Asn Thr Leu Gln Lys Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr

171

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      450              455              460
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
465              470              475              480

Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
      485              490              495

Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
      500              505              510

Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
      515              520              525

Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
      530              535              540

Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
545              550              555              560

Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
      565              570              575

Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
      580              585              590

Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
      595              600              605

Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
      610              615              620

Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
625              630              635              640

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
      645              650

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(2) INFORMATION FOR SEQ ID NO: 61:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 61:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT

48

Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu	His	Asp	Thr	Ile	Lys	Val	Thr	Pro	
1				5					10					15		
AAC	AGT	GAA	TTG	CAA	ACT	AAC	CAT	AAT	CAA	TAT	CCT	TTA	GCT	GAC	AAT	96
Asn	Ser	Glu	Leu	Gln	Thr	Asn	His	Asn	Gln	Tyr	Pro	Leu	Ala	Asp	Asn	
			20					25					30			
CCA	AAT	TCA	ACA	CTA	GAA	GAA	TTA	AAT	TAT	AAA	GAA	TTT	TTA	AGA	ATG	144
Pro	Asn	Ser	Thr	Leu	Glu	Glu	Leu	Asn	Tyr	Lys	Glu	Phe	Leu	Arg	Met	
			35					40					45			
ACT	GAA	GAC	AGT	TCT	ACG	GAA	GTG	CTA	GAC	AAC	TCT	ACA	GTA	AAA	GAT	192
Thr	Glu	Asp	Ser	Ser	Thr	Glu	Val	Leu	Asp	Asn	Ser	Thr	Val	Lys	Asp	
	50						55				60					
GCA	GTT	GGG	ACA	GGA	ATT	TCT	GTT	GTA	GGG	CAG	ATT	TTA	GGT	GTT	GTA	240
Ala	Val	Gly	Thr	Gly	Ile	Ser	Val	Val	Gly	Gln	Ile	Leu	Gly	Val	Val	
	65				70					75					80	
GGA	GTT	CCA	TTT	GCT	GGG	GCA	CTC	ACT	TCA	TTT	TAT	CAA	TCA	TTT	CTT	288
Gly	Val	Pro	Phe	Ala	Gly	Ala	Leu	Thr	Ser	Phe	Tyr	Gln	Ser	Phe	Leu	
				85					90					95		
AAC	ACT	ATA	TGG	CCA	AGT	GAT	GCT	GAC	CCA	TGG	AAG	GCT	TTT	ATG	GCA	336
Asn	Thr	Ile	Trp	Pro	Ser	Asp	Ala	Asp	Pro	Trp	Lys	Ala	Phe	Met	Ala	
			100					105					110			
CAA	GTT	GAA	GTA	CTG	ATA	GAT	AAG	AAA	ATA	GAG	GAG	TAT	GCT	AAA	AGT	384
Gln	Val	Glu	Val	Leu	Ile	Asp	Lys	Lys	Ile	Glu	Glu	Tyr	Ala	Lys	Ser	
		115					120					125				
AAA	GCT	CTT	GCA	GAG	TTA	CAG	GGT	CTT	CAA	AAT	AAT	TTC	GAA	GAT	TAT	432
Lys	Ala	Leu	Ala	Glu	Leu	Gln	Gly	Leu	Gln	Asn	Asn	Phe	Glu	Asp	Tyr	
	130					135					140					
GTT	AAT	GCG	TTA	AAT	TCC	TGG	AAG	AAA	ACA	CCT	TTA	AGT	TTG	CGA	AGT	480
Val	Asn	Ala	Leu	Asn	Ser	Trp	Lys	Lys	Thr	Pro	Leu	Ser	Leu	Arg	Ser	
	145				150					155				160		
AAA	AGA	AGC	CAA	GGT	CGA	ATA	AGG	GAA	CTT	TTT	TCT	CAA	GCA	GAA	AGT	528
Lys	Arg	Ser	Gln	Gly	Arg	Ile	Arg	Glu	Leu	Phe	Ser	Gln	Ala	Glu	Ser	
				165				170					175			
CAT	TTT	CGT	AAT	TCC	ATG	CCG	TCA	TTT	GCA	GTT	TCC	AAA	TTC	GAA	GTG	576
His	Phe	Arg	Asn	Ser	Met	Pro	Ser	Phe	Ala	Val	Ser	Lys	Phe	Glu	Val	
			180					185					190			
CTG	TTT	CTA	CCA	ACA	TAT	GCA	CAA	GCT	GCA	AAT	ACA	CAT	TTA	TTG	CTA	624
Leu	Phe	Leu	Pro	Thr	Tyr	Ala	Gln	Ala	Ala	Asn	Thr	His	Leu	Leu	Leu	
		195					200					205				
TTA	AAA	GAT	GCT	CAA	GTT	TTT	GGA	GAA	GAA	TGG	GGA	TAT	TCT	TCA	GAA	672
Leu	Lys	Asp	Ala	Gln	Val	Phe	Gly	Glu	Glu	Trp	Gly	Tyr	Ser	Ser	Glu	
	210					215					220					
GAT	GTT	GCT	GAA	TTT	TAT	CAT	AGA	CAA	TTA	AAA	CTT	ACA	CAA	CAA	TAC	720

Asp	Val	Ala	Glu	Phe	Tyr	His	Arg	Gln	Leu	Lys	Leu	Thr	Gln	Gln	Tyr	
225					230					235					240	
ACT	GAC	CAT	TGT	GTT	AAT	TGG	TAT	AAT	GTT	GGA	TTA	AAT	GGT	TTA	AGA	768
Thr	Asp	His	Cys	Val	Asn	Trp	Tyr	Asn	Val	Gly	Leu	Asn	Gly	Leu	Arg	
				245					250					255		
GGT	TCA	ACT	TAT	GAT	GCA	TGG	GTC	AAA	TTT	AAC	CGT	TTT	CGC	AGA	GAA	816
Gly	Ser	Thr	Tyr	Asp	Ala	Trp	Val	Lys	Phe	Asn	Arg	Phe	Arg	Arg	Glu	
			260					265					270			
ATG	ACT	TTA	ACT	GTA	TTA	GAT	CTA	ATT	GTA	CTT	TTC	CCA	TTT	TAT	GAT	864
Met	Thr	Leu	Thr	Val	Leu	Asp	Leu	Ile	Val	Leu	Phe	Pro	Phe	Tyr	Asp	
		275					280					285				
GTT	CGG	TTA	TAC	CCA	AAA	GGG	GTT	AAA	ACA	GAA	CTA	ACA	AGA	GAC	ATT	912
Val	Arg	Leu	Tyr	Pro	Lys	Gly	Val	Lys	Thr	Glu	Leu	Thr	Arg	Asp	Ile	
	290					295					300					
TCT	ACG	GAT	CCA	ATT	TTT	GCC	GTT	AAT	ACT	CTG	TGG	GAA	TAC	GGA	CCA	960
Ser	Thr	Asp	Pro	Ile	Phe	Ala	Val	Asn	Thr	Leu	Trp	Glu	Tyr	Gly	Pro	
305					310					315					320	
ACT	TTT	TTG	AGT	ATA	GAA	AAC	TCT	ATT	CGA	AAA	CCT	CAT	TTA	TTT	GAT	1008
Thr	Phe	Leu	Ser	Ile	Glu	Asn	Ser	Ile	Arg	Lys	Pro	His	Leu	Phe	Asp	
				325					330					335		
TAT	TTA	CAG	GGG	ATT	GAA	TTT	CAT	ACG	CGT	CTT	CGA	CCT	GGT	TAC	TTT	1056
Tyr	Leu	Gln	Gly	Ile	Glu	Phe	His	Thr	Arg	Leu	Arg	Pro	Gly	Tyr	Phe	
			340						345				350			
GGG	AAA	GAT	TCT	TTC	AAT	TAT	TGG	TCT	GGT	AAT	TAT	GCA	GAA	ACT	AGA	1104
Gly	Lys	Asp	Ser	Phe	Asn	Tyr	Trp	Ser	Gly	Asn	Tyr	Ala	Glu	Thr	Arg	
		355					360					365				
CCT	AGT	ATA	GGA	TCT	AGT	AAG	ACA	ATT	ACT	TCC	CCA	TTT	TAT	GGA	GAT	1152
Pro	Ser	Ile	Gly	Ser	Ser	Lys	Thr	Ile	Thr	Ser	Pro	Phe	Tyr	Gly	Asp	
		370				375					380					
AAA	TCT	ACT	GAA	CCT	GTA	CAA	AAG	CTA	AGC	TTT	GAT	GGA	CAA	AAA	GTT	1200
Lys	Ser	Thr	Glu	Pro	Val	Gln	Lys	Leu	Ser	Phe	Asp	Gly	Gln	Lys	Val	
385					390				395						400	
TAT	CGA	ACT	ATA	GCT	AAT	ACA	GAC	GTA	GCG	GCT	TGG	CCG	AAT	GGT	AAG	1248
Tyr	Arg	Thr	Ile	Ala	Asn	Thr	Asp	Val	Ala	Ala	Trp	Pro	Asn	Gly	Lys	
				405					410					415		
GTA	TAT	TTA	GGT	GTT	ACG	AAA	GTT	GAT	TTT	AGT	CAA	TAT	GAT	GAT	CAA	1296
Val	Tyr	Leu	Gly	Val	Thr	Lys	Val	Asp	Phe	Ser	Gln	Tyr	Asp	Asp	Gln	
			420					425					430			
AAA	AAT	GAA	ACT	AGT	ACA	CAA	ACA	TAT	GAT	TCA	AAA	AGA	AAC	AAT	GGC	1344
Lys	Asn	Glu	Thr	Ser	Thr	Gln	Thr	Tyr	Asp	Ser	Lys	Arg	Asn	Asn	Gly	
		435					440					445				
CAT	GTA	AGT	GCA	CAG	GAT	TCT	ATT	GAC	CAA	TTA	CCG	CCA	GAA	ACA	ACA	1392

His	Val	Ser	Ala	Gln	Asp	Ser	Ile	Asp	Gln	Leu	Pro	Pro	Glu	Thr	Thr	
450						455					460					
GAT	GAA	CCA	CTT	GAA	AAA	GCA	TAT	AGT	CAT	CAG	CTT	AAT	TAC	GCG	GAA	1440
Asp	Glu	Pro	Leu	Glu	Lys	Ala	Tyr	Ser	His	Gln	Leu	Asn	Tyr	Ala	Glu	
465					470					475					480	
TGT	TTC	TTA	ATG	CAG	GAC	CGT	CGT	GGA	ACA	ATT	CCA	TTT	TTT	ACT	TGG	1488
Cys	Phe	Leu	Met	Gln	Asp	Arg	Arg	Gly	Thr	Ile	Pro	Phe	Phe	Thr	Trp	
				485				490						495		
ACA	CAT	AGA	AGT	GTA	GAC	TTT	TTT	AAT	ACA	ATT	GAT	GCT	GAA	AAG	ATT	1536
Thr	His	Arg	Ser	Val	Asp	Phe	Phe	Asn	Thr	Ile	Asp	Ala	Glu	Lys	Ile	
			500					505					510			
ACT	CAA	CTT	CCA	GTA	GTG	AAA	GCA	TAT	GCC	TTG	TCT	TCA	GGT	GCT	TCC	1584
Thr	Gln	Leu	Pro	Val	Val	Lys	Ala	Tyr	Ala	Leu	Ser	Ser	Gly	Ala	Ser	
	515						520					525				
ATT	ATT	GAA	GGT	CCA	GGA	TTC	ACA	GGA	GGA	AAT	TTA	CTA	TTC	CTA	AAA	1632
Ile	Ile	Glu	Gly	Pro	Gly	Phe	Thr	Gly	Gly	Asn	Leu	Leu	Phe	Leu	Lys	
	530					535					540					
GAA	TCT	AGT	AAT	TCA	ATT	GCT	AAA	TTT	AAA	GTT	ACA	TTA	AAT	TCA	GCA	1680
Glu	Ser	Ser	Asn	Ser	Ile	Ala	Lys	Phe	Lys	Val	Thr	Leu	Asn	Ser	Ala	
545					550					555					560	
GCC	TTG	TTA	CAA	CGA	TAT	CGT	GTA	AGA	ATA	CGC	TAT	GCT	TCT	ACC	ACT	1728
Ala	Leu	Leu	Gln	Arg	Tyr	Arg	Val	Arg	Ile	Arg	Tyr	Ala	Ser	Thr	Thr	
				565					570					575		
AAC	TTA	CGA	CTT	TTT	GTG	CAA	AAT	TCA	AAC	AAT	GAT	TTT	CTT	GTC	ATC	1776
Asn	Leu	Arg	Leu	Phe	Val	Gln	Asn	Ser	Asn	Asn	Asp	Phe	Leu	Val	Ile	
			580					585					590			
TAC	ATT	AAT	AAA	ACT	ATG	AAT	AAA	GAT	GAT	GAT	TTA	ACA	TAT	CAA	ACA	1824
Tyr	Ile	Asn	Lys	Thr	Met	Asn	Lys	Asp	Asp	Asp	Leu	Thr	Tyr	Gln	Thr	
		595					600					605				
TTT	GAT	CTC	GCA	ACT	ACT	AAT	TCT	AAT	ATG	GGG	TTC	TCG	GGT	GAT	AAG	1872
Phe	Asp	Leu	Ala	Thr	Thr	Asn	Ser	Asn	Met	Gly	Phe	Ser	Gly	Asp	Lys	
	610					615					620					
AAT	GAA	CTT	ATA	ATA	GGA	GCA	GAA	TCT	TTC	GTT	TCT	AAT	GAA	AAA	ATC	1920
Asn	Glu	Leu	Ile	Ile	Gly	Ala	Glu	Ser	Phe	Val	Ser	Asn	Glu	Lys	Ile	
625					630					635					640	
TAT	ATA	GAT	AAG	ATA	GAA	TTT	ATC	CCA	GTA	CAA	TTG	TAA				1959
Tyr	Ile	Asp	Lys	Ile	Glu	Phe	Ile	Pro	Val	Gln	Leu					
				645						650						

(2) INFORMATION FOR SEQ ID NO: 62:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 652 amino acids

175

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 62:

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Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
 1             5             10             15

Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
 20             25             30

Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
 35             40             45

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50             55             60

Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65             70             75             80

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85             90             95

Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
100             105             110

Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
115             120             125

Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
130             135             140

Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
145             150             155             160

Lys Arg Ser Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
165             170             175

His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
180             185             190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
195             200             205

Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
210             215             220

Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
225             230             235             240

Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
245             250             255

Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
260             265             270

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Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Val Arg Leu Tyr Pro Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Ser Thr Asp Pro Ile Phe Ala Val Asn Thr Leu Trp Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Arg Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Ala Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr

177

	565		570		575
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile					
580		585		590	
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr					
595		600		605	
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys					
610		615		620	
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile					
625		630		635	640
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu					
645		650			

(2) INFORMATION FOR SEQ ID NO: 63:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 63:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	

178

85	90	95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala _100 105 _ 110			336
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser 115 120 125			384
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr 130 135 140			432
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser 145 150 155 160			480
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser 165 170 175			528
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val 180 185 190			576
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu 195 200 205			624
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220			672
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240			720
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255			768
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu _260 265 270			816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 275 280 285			864
GTT CGG TTA TAC CCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Val Arg Leu Tyr Pro Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300			912
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro			960

305	310	315	320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT				1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp				
325		330	335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CGA CCT GGT TAC TTT				1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Arg Pro Gly Tyr Phe				
340		345	350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA				1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg				
355		360	365	
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT				1152
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp				
370		375	380	
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT				1200
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val				
385		390	395	400
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG				1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys				
405		410	415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA				1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln				
420		425	430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC				1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly				
435		440	445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA				1392
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr				
450		455	460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA				1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu				
465		470	475	480
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG				1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp				
485		490	495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT				1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile				
500		505	510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC				1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser				
515		520	525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA				1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys				

180

530	535	540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA			1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala			
545	550	555	560
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT			1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr			
	565	570	575
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC			1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile			
	580	585	590
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA			1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr			
	595	600	605
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG			1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys			
	610	615	620
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC			1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile			
	625	630	640
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA			1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu			
	645	650	

(2) INFORMATION FOR SEQ ID NO: 64:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 652 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 64:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro			
1	5	10	15
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn			
	20	25	30
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met			
	35	40	45
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp			
	50	55	60
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val			
	65	70	75
			80

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95
 Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110
 Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125
 Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140
 Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
 145 150 155 160
 Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190
 Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Val Arg Leu Tyr Pro Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Arg Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380

182

Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575
 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590
 Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605
 Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 65:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1959 base pairs

- (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 65:

ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT	48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro	
1 5 10 15	
AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT	96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	
20 25 30	
CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GGT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	

CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val 180 185 190	576
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu 195 200 205	624
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu 210 215 220	672
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr 225 230 235 240	720
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg 245 250 255	768
GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu 260 265 270	816
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp 275 280 285	864
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile 290 295 300	912
TTT ACG GAT CCA ATT TTT TTA CTT AAT ACT CTT CAG GAG TAT GGA CCA Phe Thr Asp Pro Ile Phe Leu Leu Asn Thr Leu Gln Glu Tyr Gly Pro 305 310 315 320	960
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp 325 330 335	1008
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe 340 345 350	1056
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg 355 360 365	1104
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp 370 375 380	1152
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val 385 390 395 400	1200

TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG	1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys	
405 410 415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA	1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln	
420 425 430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC	1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly	
435 440 445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA	1392
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr	
450 455 460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA	1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu	
465 470 475 480	
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG	1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	
485 490 495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT	1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	
500 505 510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	

AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC 1920
 Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640

TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA 1959
 Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 66:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 66:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
 1 5 10 15
 Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
 20 25 30
 Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
 35 40 45
 Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
 50 55 60
 Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65 70 75 80
 Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95
 Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110
 Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125
 Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140
 Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
 145 150 155 160
 Lys Arg Ser Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190

Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Leu Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495

Thr	His	Arg	Ser	Val	Asp	Phe	Phe	Asn	Thr	Ile	Asp	Ala	Glu	Lys	Ile
			500				505						510		
Thr	Gln	Leu	Pro	Val	Val	Lys	Ala	Tyr	Ala	Leu	Ser	Ser	Gly	Ala	Ser
			515				520						525		
Ile	Ile	Glu	Gly	Pro	Gly	Phe	Thr	Gly	Gly	Asn	Leu	Leu	Phe	Leu	Lys
			530				535						540		
Glu	Ser	Ser	Asn	Ser	Ile	Ala	Lys	Phe	Lys	Val	Thr	Leu	Asn	Ser	Ala
			545				550						555		
Ala	Leu	Leu	Gln	Arg	Tyr	Arg	Val	Arg	Ile	Arg	Tyr	Ala	Ser	Thr	Thr
			565				570						575		
Asn	Leu	Arg	Leu	Phe	Val	Gln	Asn	Ser	Asn	Asn	Asp	Phe	Leu	Val	Ile
			580				585						590		
Tyr	Ile	Asn	Lys	Thr	Met	Asn	Lys	Asp	Asp	Asp	Leu	Thr	Tyr	Gln	Thr
			595				600						605		
Phe	Asp	Leu	Ala	Thr	Thr	Asn	Ser	Asn	Met	Gly	Phe	Ser	Gly	Asp	Lys
			610				615						620		
Asn	Glu	Leu	Ile	Ile	Gly	Ala	Glu	Ser	Phe	Val	Ser	Asn	Glu	Lys	Ile
			625				630						635		
Tyr	Ile	Asp	Lys	Ile	Glu	Phe	Ile	Pro	Val	Gln	Leu				
			645				650								

(2) INFORMATION FOR SEQ ID NO: 67:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
(B) LOCATION: 1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 67:

ATG	AAT	CCA	AAC	AAT	CGA	AGT	GAA	CAT	GAT	ACG	ATA	AAG	GTT	ACA	CCT	48
Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu	His	Asp	Thr	Ile	Lys	Val	Thr	Pro	
1				5					10					15		
AAC	AGT	GAA	TTG	CAA	ACT	AAC	CAT	AAT	CAA	TAT	CCT	TTA	GCT	GAC	AAT	96
Asn	Ser	Glu	Leu	Gln	Thr	Asn	His	Asn	Gln	Tyr	Pro	Leu	Ala	Asp	Asn	
			20					25					30			

CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG	144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	
35 40 45	
ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT	192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	
50 55 60	
GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA	240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	
65 70 75 80	
GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT	288
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	
85 90 95	
AAC ACT ATA TGG CCA AGT GAT GCT GAC CCA TGG AAG GCT TTT ATG GCA	336
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	
100 105 110	
CAA GTT GAA GTA CTG ATA GAT AAG AAA ATA GAG GAG TAT GCT AAA AGT	384
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	
115 120 125	
AAA GCT CTT GCA GAG TTA CAG GGT CTT CAA AAT AAT TTC GAA GAT TAT	432
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	
130 135 140	
GTT AAT GCG TTA AAT TCC TGG AAG AAA ACA CCT TTA AGT TTG CGA AGT	480
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	
145 150 155 160	
AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA AGT	528
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	
165 170 175	
CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA GTG	576
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	
180 185 190	
CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG CTA	624
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	
195 200 205	
TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA GAA	672
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	
210 215 220	
GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA TAC	720
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	
225 230 235 240	
ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA AGA	768
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	
245 250 255	

190

GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA GAA	816
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	
260 265 270	
ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT GAT	864
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	
275 280 285	
ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC ATT	912
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	
290 295 300	
TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA CCA	960
Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro	
305 310 315 320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT	1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	
325 330 335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CGA CCT GGT TAC TTT	1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Arg Pro Gly Tyr Phe	
340 345 350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA	1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg	
355 360 365	
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT	1152
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp	
370 375 380	
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT	1200
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val	
385 390 395 400	
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG	1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys	
405 410 415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA	1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln	
420 425 430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC	1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly	
435 440 445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA	1392
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr	
450 455 460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA	1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu	
465 470 475 480	

191

TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG	1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	
485 490 495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT	1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	
500 505 510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys	
530 535 540	
GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA GCA	1680
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala	
545 550 555 560	
GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC ACT	1728
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr	
565 570 575	
AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC ATC	1776
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile	
580 585 590	
TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA ACA	1824
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr	
595 600 605	
TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT AAG	1872
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys	
610 615 620	
AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA ATC	1920
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile	
625 630 635 640	
TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1959
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
645 650	

(2) INFORMATION FOR SEQ ID NO: 68:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 68:

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro

192

1	5	10	15
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn	20	25	30
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met	35	40	45
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp	50	55	60
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val	65	70	75
Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu	85	90	95
Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala	100	105	110
Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser	115	120	125
Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr	130	135	140
Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser	145	150	155
Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser	165	170	175
His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val	180	185	190
Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu	195	200	205
Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu	210	215	220
Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr	225	230	235
Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg	245	250	255
Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu	260	265	270
Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp	275	280	285
Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile	290	295	300

Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Arg Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400
 Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415
 Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430
 Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445
 His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460
 Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480
 Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495
 Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510
 Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525
 Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540
 Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560
 Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr
 565 570 575
 Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile
 580 585 590
 Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr
 595 600 605

Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys
 610 615 620

Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile
 625 630 635 640

Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
 645 650

(2) INFORMATION FOR SEQ ID NO: 69:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1482 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION:1..1479

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 69:

AGT AAA AGA AGC CAA GAT CGA ATA AGG GAA CTT TTT TCT CAA GCA GAA	48
Ser Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu	
1 5 10 15	
AGT CAT TTT CGT AAT TCC ATG CCG TCA TTT GCA GTT TCC AAA TTC GAA	96
Ser His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu	
20 25 30	
GTG CTG TTT CTA CCA ACA TAT GCA CAA GCT GCA AAT ACA CAT TTA TTG	144
Val Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu	
35 40 45	
CTA TTA AAA GAT GCT CAA GTT TTT GGA GAA GAA TGG GGA TAT TCT TCA	192
Leu Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser	
50 55 60	
GAA GAT GTT GCT GAA TTT TAT CAT AGA CAA TTA AAA CTT ACA CAA CAA	240
Glu Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln	
65 70 75 80	
TAC ACT GAC CAT TGT GTT AAT TGG TAT AAT GTT GGA TTA AAT GGT TTA	288
Tyr Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu	
85 90 95	
AGA GGT TCA ACT TAT GAT GCA TGG GTC AAA TTT AAC CGT TTT CGC AGA	336
Arg Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg	
100 105 110	

GAA ATG ACT TTA ACT GTA TTA GAT CTA ATT GTA CTT TTC CCA TTT TAT Glu Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr 115 120 125	384
GAT ATT CGG TTA TAC TCA AAA GGG GTT AAA ACA GAA CTA ACA AGA GAC Asp Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp 130 135 140	432
ATT TTT ACG GAT CCA ATT TTT TCA CTT AAT ACT CTT CAG GAG TAT GGA Ile Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly 145 150 155 160	480
CCA ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT Pro Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe 165 170 175	528
GAT TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC Asp Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr 180 185 190	576
TTT GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT Phe Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr 195 200 205	624
AGA CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA Arg Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly 210 215 220	672
GAT AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA Asp Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys 225 230 235 240	720
GTT TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT Val Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly 245 250 255	768
AAG GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT Lys Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp 260 265 270	816
CAA AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT Gln Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn 275 280 285	864
GGC CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA Gly His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr 290 295 300	912
ACA GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG Thr Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala 305 310 315 320	960
GAA TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT Glu Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr 325 330 335	1008

TGG ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG	1056
Trp Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys	
340 345 350	
ATT ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT	1104
Ile Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala	
355 360 365	
TCC ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA	1152
Ser Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu	
370 375 380	
AAA GAA TCT AGT AAT TCA ATT GCT AAA TTT AAA GTT ACA TTA AAT TCA	1200
Lys Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser	
385 390 395 400	
GCA GCC TTG TTA CAA CGA TAT CGT GTA AGA ATA CGC TAT GCT TCT ACC	1248
Ala Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr	
405 410 415	
ACT AAC TTA CGA CTT TTT GTG CAA AAT TCA AAC AAT GAT TTT CTT GTC	1296
Thr Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val	
420 425 430	
ATC TAC ATT AAT AAA ACT ATG AAT AAA GAT GAT GAT TTA ACA TAT CAA	1344
Ile Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln	
435 440 445	
ACA TTT GAT CTC GCA ACT ACT AAT TCT AAT ATG GGG TTC TCG GGT GAT	1392
Thr Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp	
450 455 460	
AAG AAT GAA CTT ATA ATA GGA GCA GAA TCT TTC GTT TCT AAT GAA AAA	1440
Lys Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys	
465 470 475 480	
ATC TAT ATA GAT AAG ATA GAA TTT ATC CCA GTA CAA TTG TAA	1482
Ile Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu	
485 490	

(2) INFORMATION FOR SEQ ID NO: 70:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 493 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 70:

Ser Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu
1 5 10 15
Ser His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu
20 25 30

Val Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu
 35 40 45
 Leu Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser
 50 55 60
 Glu Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln
 65 70 75 80
 Tyr Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu
 85 90 95
 Arg Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg
 100 105 110
 Glu Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr
 115 120 125
 Asp Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp
 130 135 140
 Ile Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly
 145 150 155 160
 Pro Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe
 165 170 175
 Asp Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr
 180 185 190
 Phe Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr
 195 200 205
 Arg Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly
 210 215 220
 Asp Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys
 225 230 235 240
 Val Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly
 245 250 255
 Lys Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp
 260 265 270
 Gln Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn
 275 280 285
 Gly His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr
 290 295 300
 Thr Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala
 305 310 315 320
 Glu Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr

198

	325		330		335
Trp Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys					
	340		345		350
Ile Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala					
	355		360		365
Ser Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu					
	370		375		380
Lys Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser					
	385		390		395
Ala Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr					
	405		410		415
Thr Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val					
	420		425		430
Ile Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln					
	435		440		445
Thr Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp					
	450		455		460
Lys Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys					
	465		470		475
Ile Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu					
	485		490		

(2) INFORMATION FOR SEQ ID NO: 71:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 23 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 71:

AGACAACTCT ACAGTAAAAG ATG

23

(2) INFORMATION FOR SEQ ID NO: 72:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 72:

GGTAATTGGT CAATAGAATC

20

(2) INFORMATION FOR SEQ ID NO: 73:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 39 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:21..23
- (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 73:

CAGAAGATGT TGCTGAATTC NNNCATAGAC AATTAAAAC

39

(2) INFORMATION FOR SEQ ID NO: 74:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 34 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:19..21
- (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 74:

GATGTTGCTG AATTCTATNN NAGACAATTA AAAC

34

(2) INFORMATION FOR SEQ ID NO: 75:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 33 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:17
 (D) OTHER INFORMATION:/note= "N = A, T, C or G"

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:18
 (D) OTHER INFORMATION:/note= "N = T, G, C or A"

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:19
 (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 75:

CCCATTTTAT GATATTNNNT TATACTCAAA AGG

33

(2) INFORMATION FOR SEQ ID NO: 76:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 64 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:24
 (D) OTHER INFORMATION:/note= "N = T, G, C or A"

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:one-of(25, 27, 28, 30, 34, 36, 39, 43)
 (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:one-of(31, 33, 35, 37, 42, 44)
 (D) OTHER INFORMATION:/note= "N = A, G, C or T"

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:40
 (D) OTHER INFORMATION:/note= "N = A, T, C or G"

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:one-of(26, 29, 32, 38, 41)
 (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 76:

AGCTATGCTG GTCTCGGAAG AAANNNNNNNN NNNNNNNNNN NNNNAAAAGA AGCCAAGATC 60
GAAT 64

(2) INFORMATION FOR SEQ ID NO: 77:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 40 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 77:

GGTCACCTAG GTCTCTCTTC CAGGAATTTA ACGCATTAAC 40

(2) INFORMATION FOR SEQ ID NO: 78:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 65 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: modified_base
(B) LOCATION:one-of(22, 27, 29, 30, 37, 42)
(D) OTHER INFORMATION:/note= "N = A, G, C or T"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
(B) LOCATION:one-of(23, 26, 28, 31, 38, 40, 43, 44)
(D) OTHER INFORMATION:/note= "N = T, G, C or A"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
(B) LOCATION:one-of(24, 39)
(D) OTHER INFORMATION:/note= "N = A, T, G or C"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
(B) LOCATION:one-of(25, 32, 33, 41, 46, 47, 48)
(D) OTHER INFORMATION:/note= "N = A, T, C or G"

(ix) FEATURE:

- (A) NAME/KEY: modified_base

- (B) LOCATION:34
- (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:45
- (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:35..36
- (D) OTHER INFORMATION:/note= "N = A, G, C or T"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 78:

AGCTATGCTG GTCTCCCAT TNNNNNNNNN NNNNNNNNNN NNNNNNNNGT TAAAACAGAA 60
CTAAC 65

(2) INFORMATION FOR SEQ ID NO: 79:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 36 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 79:

ATCCAGTGGG GTCTCAAATG GGAAAAGTAC AATTAG 36

(2) INFORMATION FOR SEQ ID NO: 80:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 63 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:one-of(23, 27, 31, 36, 44)
- (D) OTHER INFORMATION:/note= "N = A, G, C or T"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:one-of(24, 25, 26, 33, 35, 38)
- (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:one-of(28, 34, 37)
 (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:one-of(29, 30, 32, 39, 42, 45)
 (D) OTHER INFORMATION:/note= "N = T, G, C or A"

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:one-of(40, 43)
 (D) OTHER INFORMATION:/note= "N = A, T, C or G"

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:41
 (D) OTHER INFORMATION:/note= "N = A, C, T or G"

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:46
 (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 80:

CATTTTACG GATCCAATTT TTNNNNNNNN NNNNNNNNNN NNNNNNGGAC CAACTTTTTT 60

GAG 63

(2) INFORMATION FOR SEQ ID NO: 81:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 62 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:one-of(28, 31, 32, 33, 42)
 (D) OTHER INFORMATION:/note= "N = A, G, C or T"

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:one-of(29, 38, 39, 41)
 (D) OTHER INFORMATION:/note= "N = T, G, C or A"

(ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:30
 (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:one-of(34, 35, 40)
- (D) OTHER INFORMATION:/note= "N = A, T, C or G"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:36
- (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:37
- (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 81:

GAATTTTCATA CGCGTCTTCA ACCTGGTNNN NNNNNNNNNN NNTCTTTCAA TTATTGGTCT 60
GG 62

(2) INFORMATION FOR SEQ ID NO: 82:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 73 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:one-of(41, 49, 52)
- (D) OTHER INFORMATION:/note= "N = A, G, C or T"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:42..43
- (D) OTHER INFORMATION:/note= "N = A, T, C or G"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:44..45
- (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:46
- (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:one-of(47, 48, 53, 54)

(D) OTHER INFORMATION:/note= "N = T, G, C or A"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:one-of(50, 51, 55)
- (D) OTHER INFORMATION:/note= "N = A, T, C or G"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 82:

AAAAGTTTAT CGAACTATAG CTAATACAGA CGTAGCGGCT NNNNNNNNNN NNNNNGTATA 60

TTTAGGTGTT ACG 73

(2) INFORMATION FOR SEQ ID NO: 83:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 83:

GGAGTTCCAT TTGCTGGGGC 20

(2) INFORMATION FOR SEQ ID NO: 84:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 17 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 84:

ATCTCCATAA AATGGGG 17

(2) INFORMATION FOR SEQ ID NO: 85:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 32 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 85:

GCGAAGTAAA AGAAGCCAAG GTCGAATAAG GG

32

(2) INFORMATION FOR SEQ ID NO: 86:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 43 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 86:

CCTTTAAGTT TGCGAAATCC ACACAGCCAA GGTCGAATAA GGG

43

(2) INFORMATION FOR SEQ ID NO: 87:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 35 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 87:

CCCATTTTAT GATGTTCGGT TATACCCAAA AGGGG

35

(2) INFORMATION FOR SEQ ID NO: 88:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 88:

GGCCAAGTGA AGACCCATGG AAGGC

25

(2) INFORMATION FOR SEQ ID NO: 89:

207

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 22 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 89:

GCAGTTTCCG GATTCGAAGT GC

22

(2) INFORMATION FOR SEQ ID NO: 90:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 90:

CCGCTACGTC TGTATTA

17

(2) INFORMATION FOR SEQ ID NO: 91:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 91:

ATAATGGAAG CACCTGA

17

(2) INFORMATION FOR SEQ ID NO: 92:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 60 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

208

- (ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:one-of(22, 26, 29)
 (D) OTHER INFORMATION:/note= "N = T, G, C or A"

- (ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:one-of(23, 33, 36)
 (D) OTHER INFORMATION:/note= "N = A, G, C or T"

- (ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:one-of(24, 27, 28, 32, 35, 37, 38)
 (D) OTHER INFORMATION:/note= "N = A, T, C or G"

- (ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:one-of(25, 30, 31, 34)
 (D) OTHER INFORMATION:/note= "N = A, T, G or C"

- (ix) FEATURE:
 (A) NAME/KEY: modified_base
 (B) LOCATION:39
 (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 92:

AGCTATGCTG GTCTCTTCTT ANNNNNNNNNN NNNNNNNNNA CAATTCCATT TTTTACTTGG 60

(2) INFORMATION FOR SEQ ID NO: 93:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 40 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 93:

ATCCAGTTGG GTCTCTAAGA AACAAACCGC GTAATTAAGC 40

(2) INFORMATION FOR SEQ ID NO: 94:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 94:

CCTCAAGGGT TATAACATCC

20

(2) INFORMATION FOR SEQ ID NO: 95:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 55 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:one-of(19, 22, 23, 31)
- (D) OTHER INFORMATION:/note= "N = A, T, C or G"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:one-of(20, 26, 27, 29, 30, 35)
- (D) OTHER INFORMATION:/note= "N = T, G, C or A"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:one-of(21, 32, 34)
- (D) OTHER INFORMATION:/note= "N = A, G, C or T"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:one-of(24, 33)
- (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:25
- (D) OTHER INFORMATION:/note= "N = A, G, T or C"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:28
- (D) OTHER INFORMATION:/note= "N = A, T, G or C"

(ix) FEATURE:

- (A) NAME/KEY: modified_base
- (B) LOCATION:36
- (D) OTHER INFORMATION:/note= "N = A, G, C or T"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 95:

GTACAAAAGC TAAGCTTTNN NNNNNNNNNN NNNNNNCGAA CTATAGCTAA TACAG

55

(2) INFORMATION FOR SEQ ID NO: 96:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 7 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS:
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 96:

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Ser Lys Arg Ser Gln Asp Arg
1             5

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(2) INFORMATION FOR SEQ ID NO: 97:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 1959 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION:1..1956

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 97:

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ATG AAT CCA AAC AAT CGA AGT GAA CAT GAT ACG ATA AAG GTT ACA CCT      48
Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
1             5             10             15

AAC AGT GAA TTG CAA ACT AAC CAT AAT CAA TAT CCT TTA GCT GAC AAT      96
Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
20             25             30

CCA AAT TCA ACA CTA GAA GAA TTA AAT TAT AAA GAA TTT TTA AGA ATG      144
Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
35             40             45

ACT GAA GAC AGT TCT ACG GAA GTG CTA GAC AAC TCT ACA GTA AAA GAT      192
Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
50             55             60

GCA GTT GGG ACA GGA ATT TCT GTT GTA GGG CAG ATT TTA GGT GTT GTA      240
Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
65             70             75             80

GGA GTT CCA TTT GCT GGG GCA CTC ACT TCA TTT TAT CAA TCA TTT CTT      288

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211

Gly	Val	Pro	Phe	Ala	Gly	Ala	Leu	Thr	Ser	Phe	Tyr	Gln	Ser	Phe	Leu	
				85					90					95		
AAC	ACT	ATA	TGG	CCA	AGT	GAT	GCT	GAC	CCA	TGG	AAG	GCT	TTT	ATG	GCA	336
Asn	Thr	Ile	Trp	Pro	Ser	Asp	Ala	Asp	Pro	Trp	Lys	Ala	Phe	Met	Ala	
			100					105					110			
CAA	GTT	GAA	GTA	CTG	ATA	GAT	AAG	AAA	ATA	GAG	GAG	TAT	GCT	AAA	AGT	384
Gln	Val	Glu	Val	Leu	Ile	Asp	Lys	Lys	Ile	Glu	Glu	Tyr	Ala	Lys	Ser	
		115					120					125				
AAA	GCT	CTT	GCA	GAG	TTA	CAG	GGT	CTT	CAA	AAT	AAT	TTC	GAA	GAT	TAT	432
Lys	Ala	Leu	Ala	Glu	Leu	Gln	Gly	Leu	Gln	Asn	Asn	Phe	Glu	Asp	Tyr	
	130					135				140						
GTT	AAT	GCG	TTA	AAT	TCC	TGG	AAG	AAA	ACA	CCT	TTA	AGT	TTG	CGA	AGT	480
Val	Asn	Ala	Leu	Asn	Ser	Trp	Lys	Lys	Thr	Pro	Leu	Ser	Leu	Arg	Ser	
145					150					155				160		
AAA	AGA	AGC	CAA	GAT	CGA	ATA	AGG	GAA	CTT	TTT	TCT	CAA	GCA	GAA	AGT	528
Lys	Arg	Ser	Gln	Asp	Arg	Ile	Arg	Glu	Leu	Phe	Ser	Gln	Ala	Glu	Ser	
			165					170						175		
CAT	TTT	CGT	AAT	TCC	ATG	CCG	TCA	TTT	GCA	GTT	TCC	AAA	TTC	GAA	GTG	576
His	Phe	Arg	Asn	Ser	Met	Pro	Ser	Phe	Ala	Val	Ser	Lys	Phe	Glu	Val	
			180					185					190			
CTG	TTT	CTA	CCA	ACA	TAT	GCA	CAA	GCT	GCA	AAT	ACA	CAT	TTA	TTG	CTA	624
Leu	Phe	Leu	Pro	Thr	Tyr	Ala	Gln	Ala	Ala	Asn	Thr	His	Leu	Leu	Leu	
		195					200					205				
TTA	AAA	GAT	GCT	CAA	GTT	TTT	GGA	GAA	GAA	TGG	GGA	TAT	TCT	TCA	GAA	672
Leu	Lys	Asp	Ala	Gln	Val	Phe	Gly	Glu	Glu	Trp	Gly	Tyr	Ser	Ser	Glu	
	210					215					220					
GAT	GTT	GCT	GAA	TTT	TAT	CAT	AGA	CAA	TTA	AAA	CTT	ACA	CAA	CAA	TAC	720
Asp	Val	Ala	Glu	Phe	Tyr	His	Arg	Gln	Leu	Lys	Leu	Thr	Gln	Gln	Tyr	
225				230				235						240		
ACT	GAC	CAT	TGT	GTT	AAT	TGG	TAT	AAT	GTT	GGA	TTA	AAT	GGT	TTA	AGA	768
Thr	Asp	His	Cys	Val	Asn	Trp	Tyr	Asn	Val	Gly	Leu	Asn	Gly	Leu	Arg	
			245					250					255			
GGT	TCA	ACT	TAT	GAT	GCA	TGG	GTC	AAA	TTT	AAC	CGT	TTT	CGC	AGA	GAA	816
Gly	Ser	Thr	Tyr	Asp	Ala	Trp	Val	Lys	Phe	Asn	Arg	Phe	Arg	Arg	Glu	
			260					265					270			
ATG	ACT	TTA	ACT	GTA	TTA	GAT	CTA	ATT	GTA	CTT	TTC	CCA	TTT	TAT	GAT	864
Met	Thr	Leu	Thr	Val	Leu	Asp	Leu	Ile	Val	Leu	Phe	Pro	Phe	Tyr	Asp	
		275					280					285				
ATT	CGG	TTA	TAC	TCA	AAA	GGG	GTT	AAA	ACA	GAA	CTA	ACA	AGA	GAC	ATT	912
Ile	Arg	Leu	Tyr	Ser	Lys	Gly	Val	Lys	Thr	Glu	Leu	Thr	Arg	Asp	Ile	
	290					295					300					
TTT	ACG	GAT	CCA	ATT	TTT	TCA	CTT	AAT	ACT	CTT	CAG	GAG	TAT	GGA	CCA	960

Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro	
305 310 315 320	
ACT TTT TTG AGT ATA GAA AAC TCT ATT CGA AAA CCT CAT TTA TTT GAT	1008
Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp	
325 330 335	
TAT TTA CAG GGG ATT GAA TTT CAT ACG CGT CTT CAA CCT GGT TAC TTT	1056
Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe	
340 345 350	
GGG AAA GAT TCT TTC AAT TAT TGG TCT GGT AAT TAT GTA GAA ACT AGA	1104
Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg	
355 360 365	
CCT AGT ATA GGA TCT AGT AAG ACA ATT ACT TCC CCA TTT TAT GGA GAT	1152
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp	
370 375 380	
AAA TCT ACT GAA CCT GTA CAA AAG CTA AGC TTT GAT GGA CAA AAA GTT	1200
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val	
385 390 395 400	
TAT CGA ACT ATA GCT AAT ACA GAC GTA GCG GCT TGG CCG AAT GGT AAG	1248
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys	
405 410 415	
GTA TAT TTA GGT GTT ACG AAA GTT GAT TTT AGT CAA TAT GAT GAT CAA	1296
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln	
420 425 430	
AAA AAT GAA ACT AGT ACA CAA ACA TAT GAT TCA AAA AGA AAC AAT GGC	1344
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly	
435 440 445	
CAT GTA AGT GCA CAG GAT TCT ATT GAC CAA TTA CCG CCA GAA ACA ACA	1392
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr	
450 455 460	
GAT GAA CCA CTT GAA AAA GCA TAT AGT CAT CAG CTT AAT TAC GCG GAA	1440
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu	
465 470 475 480	
TGT TTC TTA ATG CAG GAC CGT CGT GGA ACA ATT CCA TTT TTT ACT TGG	1488
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp	
485 490 495	
ACA CAT AGA AGT GTA GAC TTT TTT AAT ACA ATT GAT GCT GAA AAG ATT	1536
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile	
500 505 510	
ACT CAA CTT CCA GTA GTG AAA GCA TAT GCC TTG TCT TCA GGT GCT TCC	1584
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser	
515 520 525	
ATT ATT GAA GGT CCA GGA TTC ACA GGA GGA AAT TTA CTA TTC CTA AAA	1632

213

Ile	Ile	Glu	Gly	Pro	Gly	Phe	Thr	Gly	Gly	Asn	Leu	Leu	Phe	Leu	Lys		
530						535					540						
GAA	TCT	AGT	AAT	TCA	ATT	GCT	AAA	TTT	AAA	GTT	ACA	TTA	AAT	TCA	GCA	1680	
Glu	Ser	Ser	Asn	Ser	Ile	Ala	Lys	Phe	Lys	Val	Thr	Leu	Asn	Ser	Ala		
545					550					555					560		
GCC	TTG	TTA	CAA	CGA	TAT	CGT	GTA	AGA	ATA	CGC	TAT	GCT	TCT	ACC	ACT	1728	
Ala	Leu	Leu	Gln	Arg	Tyr	Arg	Val	Arg	Ile	Arg	Tyr	Ala	Ser	Thr	Thr		
				565					570					575			
AAC	TTA	CGA	CTT	TTT	GTG	CAA	AAT	TCA	AAC	AAT	GAT	TTT	CTT	GTC	ATC	1776	
Asn	Leu	Arg	Leu	Phe	Val	Gln	Asn	Ser	Asn	Asn	Asp	Phe	Leu	Val	Ile		
			580					585					590				
TAC	ATT	AAT	AAA	ACT	ATG	AAT	AAA	GAT	GAT	GAT	TTA	ACA	TAT	CAA	ACA	1824	
Tyr	Ile	Asn	Lys	Thr	Met	Asn	Lys	Asp	Asp	Asp	Leu	Thr	Tyr	Gln	Thr		
		595					600					605					
TTT	GAT	CTC	GCA	ACT	ACT	AAT	TCT	AAT	ATG	GGG	TTC	TCG	GGT	GAT	AAG	1872	
Phe	Asp	Leu	Ala	Thr	Thr	Asn	Ser	Asn	Met	Gly	Phe	Ser	Gly	Asp	Lys		
	610						615					620					
AAT	GAA	CTT	ATA	ATA	GGA	GCA	GAA	TCT	TTC	GTT	TCT	AAT	GAA	AAA	ATC	1920	
Asn	Glu	Leu	Ile	Ile	Gly	Ala	Glu	Ser	Phe	Val	Ser	Asn	Glu	Lys	Ile		
625						630					635				640		
TAT	ATA	GAT	AAG	ATA	GAA	TTT	ATC	CCA	GTA	CAA	TTG	TAA				1959	
Tyr	Ile	Asp	Lys	Ile	Glu	Phe	Ile	Pro	Val	Gln	Leu						
				645						650							

(2) INFORMATION FOR SEQ ID NO: 98:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 98:

Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu	His	Asp	Thr	Ile	Lys	Val	Thr	Pro
1				5					10					15	
Asn	Ser	Glu	Leu	Gln	Thr	Asn	His	Asn	Gln	Tyr	Pro	Leu	Ala	Asp	Asn
			20					25					30		
Pro	Asn	Ser	Thr	Leu	Glu	Glu	Leu	Asn	Tyr	Lys	Glu	Phe	Leu	Arg	Met
			35				40					45			
Thr	Glu	Asp	Ser	Ser	Thr	Glu	Val	Leu	Asp	Asn	Ser	Thr	Val	Lys	Asp
	50					55				60					
Ala	Val	Gly	Thr	Gly	Ile	Ser	Val	Val	Gly	Gln	Ile	Leu	Gly	Val	Val
65					70					75				80	

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95
 Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110
 Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125
 Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140
 Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
 145 150 155 160
 Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190
 Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp

215

370	375	380
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val		
385	390	395 400
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys		
	405	410 415
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln		
	420	425 430
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly		
	435	440 445
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr		
	450	455 460
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu		
	465	470 475 480
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp		
	485	490 495
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile		
	500	505 510
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser		
	515	520 525
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys		
	530	535 540
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala		
	545	550 555 560
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr		
	565	570 575
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile		
	580	585 590
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr		
	595	600 605
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys		
	610	615 620
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile		
	625	630 635 640
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu		
	645	650

(2) INFORMATION FOR SEQ ID NO: 99:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2000 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 99:

CCATCCATGG CAAACCCTAA CAATCGTTCC GAACACGACA CCATCAAGGT TACTCCAAAC	60
TCTGAGTTGC AAATAATCA CAACCAGTAC CCATTGGCTG ACAATCCTAA CAGTACTCTT	120
GAGGAACTTA ACTACAAGGA GTTTCTCCGG ATGACCGAAG ATAGCTCCAC TGAGGTCTC	180
GATAACTCTA CAGTGAAGGA CGCTGTTGGA ACTGGCATTG GCGTTGTGGG ACAGATTCTT	240
GGAGTGGTTG GTGTTCCATT CGCTGGAGCT TTGACCAGCT TCTACCAGTC CTTTCTCAAC	300
ACCATCTGGC CTTCAGATGC TGATCCCTGG AAGGCTTTCA TGGCCCAAGT GGAAGTCTTG	360
ATCGATAAGA AGATCGAAGA GTATGCCAAG TCTAAAGCCT TGGCTGAGTT GCAAGGTTTG	420
CAGAACAAC TCGAGGATTA CGTCAACGCA CTCAACAGCT GGAAGAAAAC TCCCTTGAGT	480
CTCAGGTCTA AGCGTTCCCA GGACCGTATT CGTGAACCTT TCAGCCAAGC CGAATCCCAC	540
TTCAGAAACT CCATGCCTAG CTTTGCCGTT TCTAAGTTCG AGGTGCTCTT CTTGCCAACA	600
TACGCACAAG CTGCCAACAC TCATCTCTTG CTTCTCAAAG ACGCTCAGGT GTTTGGTGAG	660
GAATGGGGTT ACTCCAGTGA AGATGTTGCC GAGTTCTACC GTAGGCAGCT CAAGTTGACT	720
CAACAGTACA CAGACCACTG CGTCAACTGG TACAACGTTG GGCTCAATGG TCTTAGAGGA	780
TCTACCTACG ACGCATGGGT GAAGTTCAAC AGGTTTCGTA GAGAGATGAC CTTGACTGTG	840
CTCGATCTTA TCGTTCTCTT TCCATTCTAC GACATTCGTC TTTACTCCAA AGGCGTTAAG	900
ACAGAGCTGA CCAGAGACAT CTTACCGAT CCCATCTTCC TACTTACGAC CCTGCAGAAA	960
TACGGTCCAA CTTTTCTCTC CATTGAGAAC AGCATCAGGA AGCCTCACCT CTTCGACTAT	1020
CTGCAAGGCA TTGAGTTTCA CACCAGGTTG CAACCTGGTT ACTTCGGTAA GGATTCCTTC	1080
AACTACTGGA GCGGAACTA CGTTGAAACC AGACCATCCA TCGGATCTAG CAAGACCATC	1140
ACTTCTCCAT TCTACGGTGA CAAGAGCACT GAGCCAGTGC AGAAGTTGAG CTTCGATGGG	1200
CAGAAGGTGT ATAGAACCAT CGCCAATACC GATGTTGCAG CTTGGCCTAA TGGCAAGGTC	1260
TACCTTGAG TTTACTAAAGT GGACTTCTCC CAATACGACG ATCAGAAGAA CGAGACATCT	1320
ACTCAAACCT ACGATAGTAA GAGGAACAAT GGCCATGTTT CCGCACAAGA CTCCATTGAC	1380

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CAACTTCCAC CTGAAACCAC TGATGAACCA TTGGAGAAGG CTTACAGTCA CCAACTTAAC      1440
TACGCCGAAT GCTTTCTCAT GCAAGACAGG CGTGGCACCA TTCCGTTCTT TACATGGACT      1500
CACAGGTCTG TCGACTTCTT TAACACTATC GACGCTGAGA AGATTACCCA ACTTCCCCTG      1560
GTCAAGGCTT ATGCCTTGTC CAGCGGAGCT TCCATCATTG AAGGTCCAGG CTTCAACCGGT      1620
GGCAACTTGC TCTTCCTTAA GGAGTCCAGC AACTCCATCG CCAAGTTCAA AGTGACACTT      1680
AACTCAGCAG CCTTGCTCCA ACGTTACAGG GTTCGTATCA GATACGCAAG CACTACCAAT      1740
CTTCGCCTCT TTGTCCAGAA CAGCAACAAT GATTTCCTTG TCATCTACAT CAACAAGACT      1800
ATGAACAAAG ACGATGACCT CACCTACCAA ACATTCGATC TTGCCACTAC CAATAGTAAC      1860
ATGGGATTCT CTGGTGACAA GAACGAGCTG ATCATAGGTG CTGAGAGCTT TGTCTCTAAT      1920
GAGAAGATTT ACATAGACAA GATCGAGTTC ATTCCAGTTC AACTCTAATA GATCCCCCGG      1980
GCTGCAGGAA TTCGATATCA                                     2000

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(2) INFORMATION FOR SEQ ID NO: 100:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 653 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 100:

```

Met Ala Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr
1           5           10           15

Pro Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp
20           25           30

Asn Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg
35           40           45

Met Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys
50           55           60

Asp Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val
65           70           75           80

Val Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe
85           90           95

Leu Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met

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218

100	105	110
Ala Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys		
115	120	125
Ser Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp		
130	135	140
Tyr Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg		
145	150	155
Ser Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu		
165	170	175
Ser His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu		
180	185	190
Val Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu		
195	200	205
Leu Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser		
210	215	220
Glu Asp Val Ala Glu Phe Tyr Arg Arg Gln Leu Lys Leu Thr Gln Gln		
225	230	235
Tyr Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu		
245	250	255
Arg Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg		
260	265	270
Glu Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr		
275	280	285
Asp Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp		
290	295	300
Ile Phe Thr Asp Pro Ile Phe Leu Leu Thr Thr Leu Gln Lys Tyr Gly		
305	310	315
Pro Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe		
325	330	335
Asp Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr		
340	345	350
Phe Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr		
355	360	365
Arg Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly		
370	375	380
Asp Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys		
385	390	395
		400

219

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Val Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly
      405                      410          415

Lys Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp
      420                      425          430

Gln Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn
      435                      440          445

Gly His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr
      450                      455          460

Thr Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala
      465                      470          475          480

Glu Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr
      485                      490          495

Trp Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys
      500                      505          510

Ile Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala
      515                      520          525

Ser Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu
      530                      535          540

Lys Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser
      545                      550          555          560

Ala Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr
      565                      570          575

Thr Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val
      580                      585          590

Ile Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln
      595                      600          605

Thr Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp
      610                      615          620

Lys Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys
      625                      630          635          640

Ile Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu
      645                      650

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(2) INFORMATION FOR SEQ ID NO: 101:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 2050 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 101:

TGGAGCTCCA CCGCGGTGGC GGCCGCTCTA GAACTAGTGG ATCTAGGCCT CCATATGAAC	60
CCTAACAAATC GTTCCGAACA CGACACCATC AAGGTTACTC CAAACTCTGA GTTGCAAAC	120
AATCACAACC AGTACCCATT GGCTGACAAT CCTAACAGTA CTCTTGAGGA ACTTAACTAC	180
AAGGAGTTTC TCCGGATGAC CGAAGATAGC TCCACTGAGG TTCTCGATAA CTCTACAGTG	240
AAGGACGCTG TTGGAAC	300
TTG CATTAGCGTT GTGGGACAGA TTCTTGAGT GGTGGTGT	360
CTGCTTCA	420
GATGCTGATC CCTGGAAGGC TTTCATGGCC CAAGTGGAAG TCTTGATCGA TAAGAAGATC	480
GAAGAGTATG CCAAGTCTAA AGCCTTGGCT GAGTTGCAAG GTTTGCAGAA CAACTTCGAG	540
GATTACGTCA ACGCACTCAA CAGCTGGAAG AAAACTCCCT TGAGTCTCAG GTCTAAGCGT	600
TCCCAGGACC GTATTTCGTGA ACTTTTCAGC CAAGCCGAAT CCCACTTCAG AA	660
CTAGCTTTG CCGTTTCTAA GTTCGAGGTG CTCTTCTTGC CAACATACGC ACAAGCTGCC	720
AACACTCATC TCTTGCTTCT CAAAGACGCT CAGGTGTTTG GTGAGGAATG GGGTTACTCC	780
AGTGAAGATG TTGCCGAGTT CTACCATAGG CAGCTCAAGT TGA	840
CTACGACGCA	900
TGGGTGAAGT TCAACAGGTT TCGTAGAGAG ATGACCTTGA CTGTGCTCGA TCTTATCGTT	960
CTCTTTCCAT TCTACGACAT TCGTCTTTAC TCCAAAGGCG TTAAGACAGA GCTGACCAGA	1020
GACATCTTCA CCGATCCCAT CTTCTCACTT AACACCCTGC AGGAATACGG TCCA	1080
CTCTCCATTG AGAACAGCAT CAGGAAGCCT CACCTCTTCG ACTATCTGCA AGGCATTGAG	1140
TTTCACACCA GGTGCAACC TGGTTACTTC GGTAAGGATT CCTTCAACTA CTGGAGCGGA	1200
AACTACGTTG AAACCAGACC ATCCATCGGA TCTAGCAAGA CCATCACTTC TCCATTCTAC	1260
GGTGACAAGA GCACTGAGCC AGTGCAGAAG TTGAGCTTCG ATGGGCAGAA GGTGTATAGA	1320
ACCATCGCCA ATACCGATGT TGCAGCTTGG CCTAATGGCA AGGTCTACCT TGGAGTTACT	1380
AAAGTGGACT TCTCCCAATA CGACGATCAG AAGAACGAGA CATCTACTCA AACCTACGAT	1440
AGTAAGAGGA ACAATGGCCA TGTTTCCGCA CAAGACTCCA TTGACCAACT TCCACCTGAA	1500
ACCACTGATG AACCATTGGA GAAGGCTTAC AGTCACCAAC TTA	

CTCATGCAAG ACAGGCGTGG CACCATTCCG TTCTTTACAT GGACTCACAG GTCTGTGCGAC	1560
TTCTTTAACA CTATCGACGC TGAGAAGATT ACCCAACTTC CCGTGGTCAA GGCTTATGCC	1620
TTGTCCAGCG GAGCTTCCAT CATTGAAGGT CCAGGCTTCA CCGGTGGCAA CTTGCTCTTC	1680
CTTAAGGAGT CCAGCAACTC CATCGCCAAG TTCAAAGTGA CACTTAACTC AGCAGCCTTG	1740
CTCCAACGTT ACAGGGTTCG TATCAGATAC GCAAGCACTA CCAATCTTCG CCTCTTTGTC	1800
CAGAACAGCA ACAATGATTT CTTGTGCATC TACATCAACA AGACTATGAA CAAAGACGAT	1860
GACCTCACCT ACAACACATT CGATCTTGCC ACTACCAATA GTAACATGGG ATTCTCTGGT	1920
GACAAGAACG AGCTGATCAT AGGTGCTGAG AGCTTTGTCT CTAATGAGAA GATTTACATA	1980
GACAAGATCG AGTTCATTCC AGTTCAACTC TAATAGATCC CCCGGGCTGC AGGAATTCGA	2040
TATCAAGCTT	2050

(2) INFORMATION FOR SEQ ID NO: 102:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2280 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 102:

TTAAAATTAA TTTTGTATAC TTTTCATTGT AATAATATGA TTTTAAAAC GAAAAAGTGC	60
ATATACAAC TATCAGGAGG GGGGGGATGC ACAAAGAAGA AAAGAATAAG AAGTGAATGT	120
TTATAATGTT CAATAGTTTT ATGGAAGGC ATTTTATCAG GTAGAAAGTT ATGTATTATG	180
ATAAGAATGG GAGGAAGAAA AATGAATCCA AACAATCGAA GTGAACATGA TACGATAAAG	240
GTTACACCTA ACAGTGAATT GCAAATAAC CATAATCAAT ATCCTTTAGC TGACAATCCA	300
AATTCAACAC TAGAAGAATT AAATTATAAA GAATTTTAA GAATGACTGA AGACAGTTCT	360
ACGGAAGTGC TAGACAAC TACAGTAAAA GATGCAGTTG GGACAGGAAT TTCTGTTGTA	420
GGGCAGATTT TAGGTGTTGT AGGAGTTCCA TTTGCTGGGG CACTCACTTC ATTTTATCAA	480
TCATTTCTTA AACTATATG GCCAAGTGAT GCTGACCCAT GGAAGGCTTT TATGGCACAA	540
GTTGAAGTAC TGATAGATAA GAAAATAGAG GAGTATGCTA AAAGTAAAGC TCTTGCAGAG	600
TTACAGGGTC TTCAAATAA TTTCGAAGAT TATGTTAATG CGTTAAATTC CTGGAAGAAA	660

ACACCTTTAA GTTTGC GAAG TAAAAGAAGC CAAGATCGAA TAAGGGA ACT TTTTCTCAA	720
GCAGAAAGTC ATTTTCGTAA TTCCATGCCG TCATTTGCAG TTTCCAAATT CGAAGTGCTG	780
TTTCTACCAA CATATGCACA AGCTGCAAAT ACACATTTAT TGCTATTAAA AGATGCTCAA	840
GTTTTTGGAG AAGAATGGGG ATATTCTTCA GAAGATGTTG CTGAATTTTA TCATAGACAA	900
TTAAA ACTTA CACAACAATA CACTGACCAT TGTGTTAATT GGTATAATGT TGGATTAAAT	960
GGTTTAAGAG GTTCAACTTA TGATGCATGG GTCAAATTTA ACCGTTTTTCG CAGAGAAATG	1020
ACTTTAACTG TATTAGATCT AATTGTACTT TTCCCATTTT ATGATATTCG GTTATACTCA	1080
AAAGGGGTTA AAACAGAACT AACAAGAGAC ATTTTACGG ATCCAATTTT TTCACTTAAT	1140
ACTCTTCAGG AGTATGGACC AACTTTTTTTG AGTATAGAAA ACTCTATTCG AAAACCTCAT	1200
TTATTTGATT ATTTACAGGG GATTGAATTT CATACGCGTC TTCAACCTGG TTACTTTGGG	1260
AAAGATTCTT TCAATTATTG GTCTGGTAAT TATGTAGAAA CTAGACCTAG TATAGGATCT	1320
AGTAAGACAA TTACTTCCCC ATTTTATGGA GATAAATCTA CTGAACCTGT ACAAAGCTA	1380
AGCTTTGATG GACAAAAAGT TTATCGAACT ATAGCTAATA CAGACGTAGC GGCTTGGCCG	1440
AATGGTAAGG TATATTTAGG TGTTACGAAA GTTGATTTTA GTCAATATGA TGATCAAAAA	1500
AATGAACTA GTACACAAAC ATATGATTCA AAAAGAAACA ATGGCCATGT AAGTGCACAG	1560
GATTCTATTG ACCAATTACC GCCAGAAACA ACAGATGAAC CACTTGAAAA AGCATATAGT	1620
CATCAGCTTA ATTACGCGGA ATGTTTCTTA ATGCAGGACC GTCGTGGAAC AATTCCATTT	1680
TTTACTTGGA CACATAGAAG TGTAGACTTT TTTAATACAA TTGATGCTGA AAAGATTACT	1740
CAACTTCCAG TAGTGAAAGC ATATGCCTTG TCTTCAGGTG CTTCCATTAT TGAAGGTCCA	1800
GGATTACAG GAGGAAATTT ACTATTCCTA AAAGAATCTA GTAATTCAAT TGCTAAATTT	1860
AAAGTTACAT TAAATTCAGC AGCCTTGTTA CAACGATATC GTGTAAGAAT ACGCTATGCT	1920
TCTACCACTA ACTTACGACT TTTTGTGCAA AATTCAAACA ATGATTTTCT TGTCATCTAC	1980
ATTAATAAAA CTATGAATAA AGATGATGAT TTAACATATC AAACATTTGA TCTCGCAACT	2040
ACTAATTCTA ATATGGGGTT CTCGGGTGAT AAGAATGAAC TTATAATAGG AGCAGAATCT	2100
TTCGTTTCTA ATGAAAAAAT CTATATAGAT AAGATAGAAT TTATCCCAGT ACAATTGTAA	2160
GGAGATTTTA AAATGTTGGG TGATGGTCAA AATGAAAGAA TAGGAAGGTG AATTTTGATG	2220
GTTAGGAAAG ATTCTTTTAA CAAAAGCAAC ATGGAAAAGT ATACAGTACA AATATTAACC	2280

(2) INFORMATION FOR SEQ ID NO: 103:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 32 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 103:

TAGGCCTCCA TCCATGGCAA ACCCTAACAA TC

32

(2) INFORMATION FOR SEQ ID NO: 104:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 42 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 104:

TCCCATCTTC CTACTTACGA CCCTGCAGAA ATACGGTCCA AC

42

(2) INFORMATION FOR SEQ ID NO: 105:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 28 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 105:

GACCTCACCT ACCAAACATT CGATCTTG

28

(2) INFORMATION FOR SEQ ID NO: 106:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 25 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 106:

CGAGTTCTAC CGTAGGCAGC TCAAG

25

(2) INFORMATION FOR SEQ ID NO: 107:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1959 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 107:

ATGAATCCAA ACAATCGAAG TGAACATGAT ACGATAAAGG TTACACCTAA CAGTGAATTG	60
CAAACCTAACC ATAATCAATA TCCTTTAGCT GACAATCCAA ATTCAACACT AGAAGAATTA	120
AATTATAAAG AATTTTTTAAG AATGACTGAA GACAGTTCTA CGGAAGTGCT AGACAACTCT	180
ACAGTAAAAG ATGCAGTTGG GACAGGAATT TCTGTTGTAG GGCAGATTTT AGGTGTTGTA	240
GGAGTTCCAT TTGCTGGGGC ACTCACTTCA TTTTATCAAT CATTCTCTAA CACTATATGG	300
CCAAGTGATG CTGACCCATG GAAGGCTTTT ATGGCACAAG TTGAAGTACT GATAGATAAG	360
AAAATAGAGG AGTATGCTAA AAGTAAAGCT CTTGCAGAGT TACAGGGTCT TCAAAATAAT	420
TTCGAAGATT ATGTTAATGC GTTAAATTCC TGGAAGAAAA CACCTTTAAG TTTGCGAAGT	480
AAAAGAAGCC AAGGTCGAAT AAGGGAACCT TTTTCTCAAG CAGAAAGTCA TTTTCGTAAT	540
TCCATGCCGT CATTTGCAGT TTCCAAATTC GAAGTGCTGT TTCTACCAAC ATATGCACAA	600
GCTGCAAATA CACATTTATT GCTATTAAAA GATGCTCAAG TTTTGGAGA AGAATGGGGA	660
TATTCTTCAG AAGATGTTGC TGAATTCTAT CGTAGACAAT TAAAACTTAC ACAACAATAC	720
ACTGACCATT GTGTTAATTG GTATAATGTT GGATTAAATG GTTTAAGAGG TTCAACTTAT	780
GATGCATGGG TCAAATTTAA CCGTTTTTCGC AGAGAAATGA CTTTAACTGT ATTAGATCTA	840
ATTGTACTTT TCCCATTTTA TGATATTCGG TTATACTCAA AAGGGGTAA AACAGAACTA	900
ACAAGAGACA TTTTACGGA TCCAATTTTT TTAATTACTA CGCTTCAGAA GTACGGACCA	960
ACTTTTTTGA GTATAGAAAA CTCTATTCGA AAACCTCATT TATTTGATTA TTTACAGGGG	1020
ATTGAATTTT ATACGCGTCT TCAACCTGGT TACTTTGGGA AAGATTCTTT CAATTATTGG	1080

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TCTGGTAATT ATGTAGAAAC TAGACCTAGT ATAGGATCTA GTAAGACAAT TACTTCCCCA 1140
TTTTATGGAG ATAAATCTAC TGAACCTGTA CAAAAGCTAA GCTTTGATGG ACAAAAAGTT 1200
TATCGAACTA TAGCTAATAC AGACGTAGCG GCTTGGCCGA ATGGTAAGGT ATATTTAGGT 1260
GTTACGAAAG TTGATTTTAG TCAATATGAT GATCAAAAAA ATGAAACTAG TACACAAACA 1320
TATGATTCAA AAAGAAACAA TGGCCATGTA AGTGCACAGG ATTCTATTGA CCAATTACCG 1380
CCAGAAACAA CAGATGAACC ACTTGAAAAA GCATATAGTC ATCAGCTTAA TTACGCGGAA 1440
TGTTTCTTAA TGCAGGACCG TCGTGGAACA ATTCCATTTT TACTTGGAC ACATAGAAGT 1500
GTAGACTTTT TTAATACAAT TGATGCTGAA AAGATTACTC AACTTCCAGT AGTGAAAGCA 1560
TATGCCTTGT CTTCAGGTGC TTCCATTATT GAAGGTCCAG GATTCACAGG AGGAAATTTA 1620
CTATTCCTAA AAGAATCTAG TAATTCAATT GCTAAATTTA AAGTTACATT AAATTCAGCA 1680
GCCTTGTTAC AACGATATCG TGTAAGAATA CGCTATGCTT CTACCACTAA CTTACGACTT 1740
TTTGTGCAAA ATTCAAACAA TGATTTTCTT GTCATCTACA TTAATAAAAC TATGAATAAA 1800
GATGATGATT TAACATATCA AACATTTGAT CTCGCAACTA CTAATTCTAA TATGGGGTTC 1860
TCGGGTGATA AGAATGAACT TATAATAGGA GCAGAATCTT TCGTTTCTAA TGAAAAAATC 1920
TATATAGATA AGATAGAATT TATCCCAGTA CAATTGTAA 1959

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(2) INFORMATION FOR SEQ ID NO: 108:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 108:

```

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
1           5           10           15

Asn Ser Glu Leu Gln Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
          20           25           30

Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
          35           40           45

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
50           55           60

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Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
 65 70 75 80
 Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
 85 90 95
 Asn Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110
 Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125
 Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140
 Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
 145 150 155 160
 Lys Arg Ser Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190
 Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr Arg Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Leu Leu Thr Thr Leu Gln Lys Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg

227

355	360	365
Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp		
370	375	380
Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val		
385	390	395
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys		
	405	410
		415
Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln		
	420	425
		430
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly		
	435	440
		445
His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr		
	450	455
		460
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu		
465	470	475
		480
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp		
	485	490
		495
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile		
	500	505
		510
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser		
	515	520
		525
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys		
	530	535
		540
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala		
545	550	555
		560
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr		
	565	570
		575
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Leu Val Ile		
	580	585
		590
Tyr Ile Asn Lys Thr Met Asn Lys Asp Asp Asp Leu Thr Tyr Gln Thr		
	595	600
		605
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Lys		
	610	615
		620
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile		
625	630	635
		640
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu		
	645	650

(2) INFORMATION FOR SEQ ID NO: 109:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 649 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 109:

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Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Ala Thr Glu
1           5           10           15

Asn Asn Glu Val Ser Asn Asn His Ala Gln Tyr Pro Leu Ala Asp Thr
          20           25           30

Pro Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Arg Thr Thr
          35           40           45

Asp Asn Asn Val Glu Ala Leu Asp Ser Ser Thr Thr Lys Asp Ala Ile
50           55           60

Gln Lys Gly Ile Ser Ile Ile Gly Asp Leu Leu Gly Val Val Gly Phe
65           70           75           80

Pro Tyr Gly Gly Ala Leu Val Ser Phe Tyr Thr Asn Leu Leu Asn Thr
          85           90           95

Ile Trp Pro Gly Glu Asp Pro Leu Lys Ala Phe Met Gln Gln Val Glu
          100          105          110

Ala Leu Ile Asp Gln Lys Ile Ala Asp Tyr Ala Lys Asp Lys Ala Thr
          115          120          125

Ala Glu Leu Gln Gly Leu Lys Asn Val Phe Lys Asp Tyr Val Ser Ala
130           135           140

Leu Asp Ser Trp Asp Lys Thr Pro Leu Thr Leu Arg Asp Gly Arg Ser
145           150           155           160

Gln Gly Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser His Phe Arg
          165          170          175

Arg Ser Met Pro Ser Phe Ala Val Ser Gly Tyr Glu Val Leu Phe Leu
          180          185          190

Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu Leu Lys Asp
          195          200          205

Ala Gln Ile Tyr Gly Thr Asp Trp Gly Tyr Ser Thr Asp Asp Leu Asn
210           215           220

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229

Glu Phe His Thr Lys Gln Lys Asp Leu Thr Ile Glu Tyr Thr Asn His
 225 230 235 240
 Cys Ala Lys Trp Tyr Lys Ala Gly Leu Asp Lys Leu Arg Gly Ser Thr
 245 250 255
 Tyr Glu Glu Trp Val Lys Phe Asn Arg Tyr Arg Arg Glu Met Thr Leu
 260 265 270
 Thr Val Leu Asp Leu Ile Thr Leu Phe Pro Leu Tyr Asp Val Arg Thr
 275 280 285
 Tyr Thr Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Val Leu Thr Asp
 290 295 300
 Pro Ile Val Ala Val Asn Asn Met Asn Gly Tyr Gly Thr Thr Phe Ser
 305 310 315 320
 Asn Ile Glu Asn Tyr Ile Arg Lys Pro His Leu Phe Asp Tyr Leu His
 325 330 335
 Ala Ile Gln Phe His Ser Arg Leu Gln Pro Gly Tyr Phe Gly Thr Asp
 340 345 350
 Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Ser Thr Arg Ser Ser Ile
 355 360 365
 Gly Ser Asp Glu Ile Ile Arg Ser Pro Phe Tyr Gly Asn Lys Ser Thr
 370 375 380
 Leu Asp Val Gln Asn Leu Glu Phe Asn Gly Glu Lys Val Phe Arg Ala
 385 390 395 400
 Val Ala Asn Gly Asn Leu Ala Val Trp Pro Val Gly Thr Gly Gly Thr
 405 410 415
 Lys Ile His Ser Gly Val Thr Lys Val Gln Phe Ser Gln Tyr Asn Asp
 420 425 430
 Arg Lys Asp Glu Val Arg Thr Gln Thr Tyr Asp Ser Lys Arg Asn Val
 435 440 445
 Gly Gly Ile Val Phe Asp Ser Ile Asp Gln Leu Pro Pro Ile Thr Thr
 450 455 460
 Asp Glu Ser Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Val Arg
 465 470 475 480
 Cys Phe Leu Leu Gln Gly Gly Arg Gly Ile Ile Pro Val Phe Thr Trp
 485 490 495
 Thr His Lys Ser Val Asp Phe Tyr Asn Thr Leu Asp Ser Glu Lys Ile
 500 505 510
 Thr Gln Ile Pro Phe Val Lys Ala Phe Ile Leu Val Asn Ser Thr Ser
 515 520 525

230

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Val Val Ala Gly Pro Gly Phe Thr Gly Gly Asp Ile Ile Lys Cys Thr
  530                      535                      540

Asn Gly Ser Gly Leu Thr Leu Tyr Val Thr Pro Ala Pro Asp Leu Thr
  545                      550                      555                      560

Tyr Ser Lys Thr Tyr Lys Ile Arg Ile Arg Tyr Ala Ser Thr Ser Gln
                      565                      570                      575

Val Arg Phe Gly Ile Asp Leu Gly Ser Tyr Thr His Ser Ile Ser Tyr
                      580                      585                      590

Phe Asp Lys Thr Met Asp Lys Gly Asn Thr Leu Thr Tyr Asn Ser Phe
  595                      600                      605

Asn Leu Ser Ser Val Ser Arg Pro Ile Glu Ile Ser Gly Gly Asn Lys
  610                      615                      620

Ile Gly Val Ser Val Gly Gly Ile Gly Ser Gly Asp Glu Val Tyr Ile
  625                      630                      635                      640

Asp Lys Ile Glu Phe Ile Pro Met Asp
                      645

```

(2) INFORMATION FOR SEQ ID NO: 110:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 652 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS:
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 110:

```

Met Asn Pro Asn Asn Arg Ser Glu His Asp Thr Ile Lys Val Thr Pro
  1                      5                      10                      15

Asn Ser Glu Leu Pro Thr Asn His Asn Gln Tyr Pro Leu Ala Asp Asn
                      20                      25                      30

Pro Asn Ser Thr Leu Glu Glu Leu Asn Tyr Lys Glu Phe Leu Arg Met
                      35                      40                      45

Thr Glu Asp Ser Ser Thr Glu Val Leu Asp Asn Ser Thr Val Lys Asp
                      50                      55                      60

Ala Val Gly Thr Gly Ile Ser Val Val Gly Gln Ile Leu Gly Val Val
  65                      70                      75                      80

Gly Val Pro Phe Ala Gly Ala Leu Thr Ser Phe Tyr Gln Ser Phe Leu
                      85                      90                      95

```


231

Asp Thr Ile Trp Pro Ser Asp Ala Asp Pro Trp Lys Ala Phe Met Ala
 100 105 110
 Gln Val Glu Val Leu Ile Asp Lys Lys Ile Glu Glu Tyr Ala Lys Ser
 115 120 125
 Lys Ala Leu Ala Glu Leu Gln Gly Leu Gln Asn Asn Phe Glu Asp Tyr
 130 135 140
 Val Asn Ala Leu Asn Ser Trp Lys Lys Thr Pro Leu Ser Leu Arg Ser
 145 150 155 160
 Lys Arg Ser Gln Asp Arg Ile Arg Glu Leu Phe Ser Gln Ala Glu Ser
 165 170 175
 His Phe Arg Asn Ser Met Pro Ser Phe Ala Val Ser Lys Phe Glu Val
 180 185 190
 Leu Phe Leu Pro Thr Tyr Ala Gln Ala Ala Asn Thr His Leu Leu Leu
 195 200 205
 Leu Lys Asp Ala Gln Val Phe Gly Glu Glu Trp Gly Tyr Ser Ser Glu
 210 215 220
 Asp Val Ala Glu Phe Tyr His Arg Gln Leu Lys Leu Thr Gln Gln Tyr
 225 230 235 240
 Thr Asp His Cys Val Asn Trp Tyr Asn Val Gly Leu Asn Gly Leu Arg
 245 250 255
 Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270
 Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285
 Val Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300
 Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320
 Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335
 Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Ser
 340 345 350
 Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365
 Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380
 Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val

232

385		390		395		400
Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys						
	405			410		415
Ile Tyr Phe Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln						
	420		425			430
Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly						
	435		440			445
His Val Gly Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr						
	450		455			460
Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu						
	465		470		475	480
Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp						
	485		490			495
Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile						
	500		505			510
Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser						
	515		520			525
Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys						
	530		535			540
Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala						
	545		550		555	560
Ala Leu Leu Gln Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr						
	565		570			575
Asn Leu Arg Leu Phe Val Gln Asn Ser Asn Asn Asp Phe Ile Val Ile						
	580		585			590
Tyr Ile Asn Lys Thr Met Asn Ile Asp Asp Asp Leu Thr Tyr Gln Thr						
	595		600			605
Phe Asp Leu Ala Thr Thr Asn Ser Asn Met Gly Phe Ser Gly Asp Thr						
	610		615			620
Asn Glu Leu Ile Ile Gly Ala Glu Ser Phe Val Ser Asn Glu Lys Ile						
	625		630		635	640
Tyr Ile Asp Lys Ile Glu Phe Ile Pro Val Gln Leu						
	645		650			

(2) INFORMATION FOR SEQ ID NO: 111:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:

233

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 111:

Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu	His	Asp	Thr	Ile	Lys	Val	Thr	Pro	1	5	10	15
Asn	Ser	Glu	Leu	Gln	Thr	Asn	His	Asn	Gln	Tyr	Pro	Leu	Ala	Asp	Asn	20	25	30	
Pro	Asn	Ser	Thr	Leu	Glu	Glu	Leu	Asn	Tyr	Lys	Glu	Phe	Leu	Arg	Met	35	40	45	
Thr	Glu	Asp	Ser	Ser	Thr	Glu	Val	Leu	Asp	Asn	Ser	Thr	Val	Lys	Asp	50	55	60	
Ala	Val	Gly	Thr	Gly	Ile	Ser	Val	Val	Gly	Gln	Ile	Leu	Gly	Val	Val	65	70	75	80
Gly	Val	Pro	Phe	Ala	Gly	Ala	Leu	Thr	Ser	Phe	Tyr	Gln	Ser	Phe	Leu	85	90	95	
Asn	Thr	Ile	Trp	Pro	Ser	Asp	Ala	Asp	Pro	Trp	Lys	Ala	Phe	Met	Ala	100	105	110	
Gln	Val	Glu	Val	Leu	Ile	Asp	Lys	Lys	Ile	Glu	Glu	Tyr	Ala	Lys	Ser	115	120	125	
Lys	Ala	Leu	Ala	Glu	Leu	Gln	Gly	Leu	Gln	Asn	Asn	Phe	Glu	Asp	Tyr	130	135	140	
Val	Asn	Ala	Leu	Asn	Ser	Trp	Lys	Lys	Thr	Pro	Leu	Ser	Leu	Arg	Ser	145	150	155	160
Lys	Arg	Ser	Gln	Asp	Arg	Ile	Arg	Glu	Leu	Phe	Ser	Gln	Ala	Glu	Ser	165	170	175	
His	Phe	Arg	Asn	Ser	Met	Pro	Ser	Phe	Ala	Val	Ser	Lys	Phe	Glu	Val	180	185	190	
Leu	Phe	Leu	Pro	Thr	Tyr	Ala	Gln	Ala	Ala	Asn	Thr	His	Leu	Leu	Leu	195	200	205	
Leu	Lys	Asp	Ala	Gln	Val	Phe	Gly	Glu	Glu	Trp	Gly	Tyr	Ser	Ser	Glu	210	215	220	
Asp	Val	Ala	Glu	Phe	Tyr	His	Arg	Gln	Leu	Lys	Leu	Thr	Gln	Gln	Tyr	225	230	235	240
Thr	Asp	His	Cys	Val	Asn	Trp	Tyr	Asn	Val	Gly	Leu	Asn	Gly	Leu	Arg	245	250	255	

234

Gly Ser Thr Tyr Asp Ala Trp Val Lys Phe Asn Arg Phe Arg Arg Glu
 260 265 270

Met Thr Leu Thr Val Leu Asp Leu Ile Val Leu Phe Pro Phe Tyr Asp
 275 280 285

Ile Arg Leu Tyr Ser Lys Gly Val Lys Thr Glu Leu Thr Arg Asp Ile
 290 295 300

Phe Thr Asp Pro Ile Phe Ser Leu Asn Thr Leu Gln Glu Tyr Gly Pro
 305 310 315 320

Thr Phe Leu Ser Ile Glu Asn Ser Ile Arg Lys Pro His Leu Phe Asp
 325 330 335

Tyr Leu Gln Gly Ile Glu Phe His Thr Arg Leu Gln Pro Gly Tyr Phe
 340 345 350

Gly Lys Asp Ser Phe Asn Tyr Trp Ser Gly Asn Tyr Val Glu Thr Arg
 355 360 365

Pro Ser Ile Gly Ser Ser Lys Thr Ile Thr Ser Pro Phe Tyr Gly Asp
 370 375 380

Lys Ser Thr Glu Pro Val Gln Lys Leu Ser Phe Asp Gly Gln Lys Val
 385 390 395 400

Tyr Arg Thr Ile Ala Asn Thr Asp Val Ala Ala Trp Pro Asn Gly Lys
 405 410 415

Val Tyr Leu Gly Val Thr Lys Val Asp Phe Ser Gln Tyr Asp Asp Gln
 420 425 430

Lys Asn Glu Thr Ser Thr Gln Thr Tyr Asp Ser Lys Arg Asn Asn Gly
 435 440 445

His Val Ser Ala Gln Asp Ser Ile Asp Gln Leu Pro Pro Glu Thr Thr
 450 455 460

Asp Glu Pro Leu Glu Lys Ala Tyr Ser His Gln Leu Asn Tyr Ala Glu
 465 470 475 480

Cys Phe Leu Met Gln Asp Arg Arg Gly Thr Ile Pro Phe Phe Thr Trp
 485 490 495

Thr His Arg Ser Val Asp Phe Phe Asn Thr Ile Asp Ala Glu Lys Ile
 500 505 510

Thr Gln Leu Pro Val Val Lys Ala Tyr Ala Leu Ser Ser Gly Ala Ser
 515 520 525

Ile Ile Glu Gly Pro Gly Phe Thr Gly Gly Asn Leu Leu Phe Leu Lys
 530 535 540

Glu Ser Ser Asn Ser Ile Ala Lys Phe Lys Val Thr Leu Asn Ser Ala
 545 550 555 560

Ala	Leu	Leu	Gln	Arg	Tyr	Arg	Val	Arg	Ile	Arg	Tyr	Ala	Ser	Thr	Thr
				565					570					575	
Asn	Leu	Arg	Leu	Phe	Val	Gln	Asn	Ser	Asn	Asn	Asp	Phe	Leu	Val	Ile
			580					585					590		
Tyr	Ile	Asn	Lys	Thr	Met	Asn	Lys	Asp	Asp	Asp	Leu	Thr	Tyr	Gln	Thr
		595					600					605			
Phe	Asp	Leu	Ala	Thr	Thr	Asn	Ser	Asn	Met	Gly	Phe	Ser	Gly	Asp	Lys
	610					615					620				
Asn	Glu	Leu	Ile	Ile	Gly	Ala	Glu	Ser	Phe	Val	Ser	Asn	Glu	Lys	Ile
625					630					635					640
Tyr	Ile	Asp	Lys	Ile	Glu	Phe	Ile	Pro	Val	Gln	Leu				
				645					650						

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 659 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

Met	Ile	Arg	Met	Gly	Gly	Arg	Lys	Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu
1				5					10					15	
Tyr	Asp	Thr	Ile	Lys	Val	Thr	Pro	Asn	Ser	Glu	Leu	Pro	Thr	Asn	His
			20					25					30		
Asn	Gln	Tyr	Pro	Leu	Ala	Asp	Asn	Pro	Asn	Ser	Thr	Leu	Glu	Glu	Leu
		35					40					45			
Asn	Tyr	Lys	Glu	Phe	Leu	Arg	Met	Thr	Ala	Asp	Asn	Ser	Thr	Glu	Val
	50					55					60				
Leu	Asp	Ser	Ser	Thr	Val	Lys	Asp	Ala	Val	Gly	Thr	Gly	Ile	Ser	Val
65					70					75					80
Val	Gly	Gln	Ile	Leu	Gly	Val	Val	Gly	Val	Pro	Phe	Ala	Gly	Ala	Leu
				85					90					95	
Thr	Ser	Phe	Tyr	Gln	Ser	Phe	Leu	Asn	Ala	Ile	Trp	Pro	Ser	Asp	Ala
			100					105					110		
Asp	Pro	Trp	Lys	Ala	Phe	Met	Ala	Gln	Val	Glu	Val	Leu	Ile	Asp	Lys
		115					120					125			

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Lys Ile Glu Glu Tyr Ala Lys Ser Lys Ala Leu Ala Glu Leu Gln Gly
 130 135 140
 Leu Gln Asn Asn Phe Glu Asp Tyr Val Asn Ala Leu Asp Ser Trp Lys
 145 150 155 160
 Lys Ala Pro Val Asn Leu Arg Ser Arg Arg Ser Gln Asp Arg Ile Arg
 165 170 175
 Glu Leu Phe Ser Gln Ala Glu Ser His Phe Arg Asn Ser Met Pro Ser
 180 185 190
 Phe Ala Val Ser Lys Phe Glu Val Leu Phe Leu Pro Thr Tyr Ala Gln
 195 200 205
 Ala Ala Asn Thr His Leu Leu Leu Leu Lys Asp Ala Gln Val Phe Gly
 210 215 220
 Glu Glu Trp Gly Tyr Ser Ser Glu Asp Ile Ala Glu Phe Tyr Gln Arg
 225 230 235 240
 Gln Leu Lys Leu Thr Gln Gln Tyr Thr Asp His Cys Val Asn Trp Tyr
 245 250 255
 Asn Val Gly Leu Asn Ser Leu Arg Gly Ser Thr Tyr Asp Ala Trp Val
 260 265 270
 Lys Phe Asn Arg Phe Arg Arg Glu Met Thr Leu Thr Val Leu Asp Leu
 275 280 285
 Ile Val Leu Phe Pro Phe Tyr Asp Val Arg Leu Tyr Ser Lys Gly Val
 290 295 300
 Lys Thr Glu Leu Thr Arg Asp Ile Phe Thr Asp Pro Ile Phe Thr Leu
 305 310 315 320
 Asn Ala Leu Gln Glu Tyr Gly Pro Thr Phe Ser Ser Ile Glu Asn Ser
 325 330 335
 Ile Arg Lys Pro His Leu Phe Asp Tyr Leu Arg Gly Ile Glu Phe His
 340 345 350
 Thr Arg Leu Arg Pro Gly Tyr Ser Gly Lys Asp Ser Phe Asn Tyr Trp
 355 360 365
 Ser Gly Asn Tyr Val Glu Thr Arg Pro Ser Ile Gly Ser Asn Asp Thr
 370 375 380
 Ile Thr Ser Pro Phe Tyr Gly Asp Lys Ser Ile Glu Pro Ile Gln Lys
 385 390 395 400
 Leu Ser Phe Asp Gly Gln Lys Val Tyr Arg Thr Ile Ala Asn Thr Asp
 405 410 415
 Ile Ala Ala Phe Pro Asp Gly Lys Ile Tyr Phe Gly Val Thr Lys Val

237

420	425	430
Asp Phe Ser Gln Tyr Asp Asp Gln Lys Asn Glu Thr Ser Thr Gln Thr		
435	440	445
Tyr Asp Ser Lys Arg Tyr Asn Gly Tyr Leu Gly Ala Gln Asp Ser Ile		
450	455	460
Asp Gln Leu Pro Pro Glu Thr Thr Asp Glu Pro Leu Glu Lys Ala Tyr		
465	470	475
Ser His Gln Leu Asn Tyr Ala Glu Cys Phe Leu Met Gln Asp Arg Arg		
485	490	495
Gly Thr Ile Pro Phe Phe Thr Trp Thr His Arg Ser Val Asp Phe Phe		
500	505	510
Asn Thr Ile Asp Ala Glu Lys Ile Thr Gln Leu Pro Val Val Lys Ala		
515	520	525
Tyr Ala Leu Ser Ser Gly Ala Ser Ile Ile Glu Gly Pro Gly Phe Thr		
530	535	540
Gly Gly Asn Leu Leu Phe Leu Lys Glu Ser Ser Asn Ser Ile Ala Lys		
545	550	555
Phe Lys Val Thr Leu Asn Ser Ala Ala Leu Leu Gln Arg Tyr Arg Val		
565	570	575
Arg Ile Arg Tyr Ala Ser Thr Thr Asn Leu Arg Leu Phe Val Gln Asn		
580	585	590
Ser Asn Asn Asp Phe Leu Val Ile Tyr Ile Asn Lys Thr Met Asn Ile		
595	600	605
Asp Gly Asp Leu Thr Tyr Gln Thr Phe Asp Phe Ala Thr Ser Asn Ser		
610	615	620
Asn Met Gly Phe Ser Gly Asp Thr Asn Asp Phe Ile Ile Gly Ala Glu		
625	630	635
Ser Phe Val Ser Asn Glu Lys Ile Tyr Ile Asp Lys Ile Glu Phe Ile		
645	650	655
Pro Val Gln		

(2) INFORMATION FOR SEQ ID NO: 113:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 652 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 113:

Met	Ile	Arg	Lys	Gly	Gly	Arg	Lys	Met	Asn	Pro	Asn	Asn	Arg	Ser	Glu	1	5	10	15
His	Asp	Thr	Ile	Lys	Thr	Thr	Glu	Asn	Asn	Glu	Val	Pro	Thr	Asn	His	20	25	30	
Val	Gln	Tyr	Pro	Leu	Ala	Glu	Thr	Pro	Asn	Pro	Thr	Leu	Glu	Asp	Leu	35	40	45	
Asn	Tyr	Lys	Glu	Phe	Leu	Arg	Met	Thr	Ala	Asp	Asn	Asn	Thr	Glu	Ala	50	55	60	
Leu	Asp	Ser	Ser	Thr	Thr	Lys	Asp	Val	Ile	Gln	Lys	Gly	Ile	Ser	Val	65	70	75	80
Val	Gly	Asp	Leu	Leu	Gly	Val	Val	Gly	Phe	Pro	Phe	Gly	Gly	Ala	Leu	85	90	95	
Val	Ser	Phe	Tyr	Thr	Asn	Phe	Leu	Asn	Thr	Ile	Trp	Pro	Ser	Glu	Asp	100	105	110	
Pro	Trp	Lys	Ala	Phe	Met	Glu	Gln	Val	Glu	Ala	Leu	Met	Asp	Gln	Lys	115	120	125	
Ile	Ala	Asp	Tyr	Ala	Lys	Asn	Lys	Ala	Leu	Ala	Glu	Leu	Gln	Gly	Leu	130	135	140	
Gln	Asn	Asn	Val	Glu	Asp	Tyr	Val	Ser	Ala	Leu	Ser	Ser	Trp	Gln	Lys	145	150	155	160
Asn	Pro	Val	Ser	Ser	Arg	Asn	Pro	His	Ser	Gln	Gly	Arg	Ile	Arg	Glu	165	170	175	
Leu	Phe	Ser	Gln	Ala	Glu	Ser	His	Phe	Arg	Asn	Ser	Met	Pro	Ser	Phe	180	185	190	
Ala	Ile	Ser	Gly	Tyr	Glu	Val	Leu	Phe	Leu	Thr	Thr	Tyr	Ala	Gln	Ala	195	200	205	
Ala	Asn	Thr	His	Leu	Phe	Leu	Leu	Lys	Asp	Ala	Gln	Ile	Tyr	Gly	Glu	210	215	220	
Glu	Trp	Gly	Tyr	Glu	Lys	Glu	Asp	Ile	Ala	Glu	Phe	Tyr	Lys	Arg	Gln	225	230	235	240
Leu	Lys	Leu	Thr	Gln	Glu	Tyr	Thr	Asp	His	Cys	Val	Lys	Trp	Tyr	Asn	245	250	255	
Val	Gly	Leu	Asp	Lys	Leu	Arg	Gly	Ser	Ser	Tyr	Glu	Ser	Trp	Val	Asn	260	265	270	

239

Phe Asn Arg Tyr Arg Arg Glu Met Thr Leu Thr Val Leu Asp Leu Ile
 275 280 285

Ala Leu Phe Pro Leu Tyr Asp Val Arg Leu Tyr Pro Lys Glu Val Lys
 290 - 295 300-

Thr Glu Leu Thr Arg Asp Val Leu Thr Asp Pro Ile Val Gly Val Asn
 305 310 315 320

Asn Leu Arg Gly Tyr Gly Thr Thr Phe Ser Asn Ile Glu Asn Tyr Ile
 325 330 335

Arg Lys Pro His Leu Phe Asp Tyr Leu His Arg Ile Gln Phe His Thr
 340 345 350

Arg Phe Gln Pro Gly Tyr Tyr Gly Asn Asp Ser Phe Asn Tyr Trp Ser
 355 360 365

Gly Asn Tyr Val Ser Thr Arg Pro Ser Ile Gly Ser Asn Asp Ile Ile
 370 375 380

Thr Ser Pro Phe Tyr Gly Asn Lys Ser Ser Glu Pro Val Gln Asn Leu
 385 390 395 400

Glu Phe Asn Gly Glu Lys Val Tyr Arg Ala Val Ala Asn Thr Asn Leu
 405 410 415

Ala Val Trp Pro Ser Ala Val Tyr Ser Gly Val Thr Lys Val Glu Phe
 420 425 430

Ser Gln Tyr Asn Asp Gln Thr Asp Glu Ala Ser Thr Gln Thr Tyr Asp
 435 440 445

Ser Lys Arg Asn Val Gly Ala Val Ser Trp Asp Ser Ile Asp Gln Leu
 450 455 460

Pro Pro Glu Thr Thr Asp Glu Pro Leu Glu Lys Gly Tyr Ser His Gln
 465 470 475 480

Leu Asn Tyr Val Met Cys Phe Leu Met Gln Gly Ser Arg Gly Thr Ile
 485 490 495

Pro Val Leu Thr Trp Thr His Lys Ser Val Asp Phe Phe Asn Met Ile
 500 505 510

Asp Ser Lys Lys Ile Thr Gln Leu Pro Leu Val Lys Ala Tyr Lys Leu
 515 520 525

Gln Ser Gly Ala Ser Val Val Ala Gly Pro Arg Phe Thr Gly Gly Asp
 530 535 540

Ile Ile Gln Cys Thr Glu Asn Gly Ser Ala Ala Thr Ile Tyr Val Thr
 545 550 555 560

Pro Asp Val Ser Tyr Ser Gln Lys Tyr Arg Ala Arg Ile His Tyr Ala
 565 570 575

240

Ser	Thr	Ser	Gln	Ile	Thr	Phe	Thr	Leu	Ser	Leu	Asp	Gly	Ala	Pro	Phe	
			580					585					590			
Asn	Gln	Tyr	Tyr	Phe	Asp	Lys	Thr	Ile	Asn	Lys	Gly	Asp	Thr	Leu	Thr	
		595					600					605				
Tyr	Asn	Ser	Phe	Asn	Leu	Ala	Ser	Phe	Ser	Thr	Pro	Phe	Glu	Leu	Ser	
	610					615					620					
Gly	Asn	Asn	Leu	Gln	Ile	Gly	Val	Thr	Gly	Leu	Ser	Ala	Gly	Asp	Lys	
625					630					635					640	
Val	Tyr	Ile	Asp	Lys	Ile	Glu	Phe	Ile	Pro	Val	Asn					
				645					650							

INTERNATIONAL SEARCH REPORT

International Application No.

PCT/US 98/26852

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/32 C07K14/325 A01H5/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C07K A01H

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	CODY V ET AL: "Purification and crystallization of insecticidal delta-endotoxin CryIIIB2 from <i>Bacillus thuringiensis</i> ." PROTEINS, (1992 OCT) 14 (2) 324., XP002103659 cited in the application see the whole document	94-116
Y	P GROCHULSKI ET AL: " <i>Bacillus thuringiensis</i> CryIA(a) insecticidal toxin: Crystal structure and channel formation" JOURNAL OF MOLECULAR BIOLOGY, vol. 254, no. 3, 1 January 1995, pages 447-464, XP002095820 see the whole document --- -/-	94-116



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

26 May 1999

Date of mailing of the international search report

10/06/1999

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+31-70) 340-3016

Authorized officer

Andres, S

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 98/26852

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 92 13954 A (ECOGEN INC) 20 August 1992 see page 70 - page 72 see page 15, line 26 - page 16, line 4 see page 19, line 5 - page 24 see examples 6-12	1,2,41, 42, 48-50, 53-67, 69-73, 75-82, 84-87
A	see claims	26,27, 33,34
A	--- WO 91 14778 A (ECOGEN INC) 3 October 1991 see claims; examples	1-93
X	--- EP 0 382 990 A (PLANT GENETIC SYSTEMS NV) 22 August 1990 see page 3, line 43 - page 5, line 22 see examples 3,4,8 see claims	1,2,37, 41,42, 50, 53-67, 69-73, 75-82, 84-87
A	see figure 1	3,6,8, 11,14, 17,18, 26,34, 39,40, 43-49
X	--- WO 93 15206 A (MYCOGEN CORP) 5 August 1993 see page 40 - page 42 see page 10 - page 14, line 7 see examples 3,8,9	1,2,37, 41,42, 50, 53-67, 69-73, 75-82, 84-87
A	--- VON TERSCH M A ET AL: "Membrane-permeabilizing activities of Bacillus thuringiensis coleopteran-active toxin CryIIIB2 and CryIIIB2 domain I peptide." APPLIED AND ENVIRONMENTAL MICROBIOLOGY, (1994 OCT) 60 (10) 3711-7., XP002103660 cited in the application ---	
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INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 98/26852

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>SMEDLEY, D. & ELLAR, D.: "Mutagenesis of three surface-exposed loops of a Bacillus thuringiensis insecticidal toxin reveals residues important for toxicity, receptor recognition and possibly membrane insertion." MICROBIOLOGY, vol. 142, July 1996, pages 1617-1624, XP002103661 cited in the application -----</p>	
A	<p>DEAN D H ET AL: "Probing the mechanism of action of Bacillus thuringiensis insecticidal proteins by site-directed mutagenesis -- a minireview" GENE, vol. 179, no. 1, 7 November 1996, page 111-117 XP004071972 -----</p>	

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 98/26852

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9213954 A	20-08-1992	AU 649785 B	02-06-1994
		AU 1192692 A	07-09-1992
		CA 2101338 A	01-08-1992
		EP 0569438 A	18-11-1993
		JP 2531913 B	04-09-1996
		JP 6502077 T	10-03-1994
		US 5264364 A	23-11-1993
		US 5378625 A	03-01-1995
WO 9114778 A	03-10-1991	US 5187091 A	16-02-1993
		AT 125569 T	15-08-1995
		AU 645080 B	06-01-1994
		AU 7555291 A	21-10-1991
		CA 2078571 A	21-09-1991
		DE 69111600 D	31-08-1995
		DE 69111600 T	29-02-1996
		EP 0522036 A	13-01-1993
		EP 0606110 A	13-07-1994
		ES 2077223 T	16-11-1995
		HU 62036 A	29-03-1993
		JP 5505109 T	05-08-1993
		PL 165919 B	31-03-1995
		US 5382429 A	17-01-1995
EP 0382990 A	22-08-1990	AT 100144 T	15-01-1994
		AU 633303 B	28-01-1993
		AU 5047690 A	05-09-1990
		CA 2046646 A	16-08-1990
		DE 69006015 D	24-02-1994
		DE 69006015 T	26-05-1994
		DK 458819 T	07-03-1994
		WO 9009445 A	23-08-1990
		EP 0458819 A	04-12-1991
		ES 2062512 T	16-12-1994
		US 5683691 A	04-11-1997
WO 9315206 A	05-08-1993	US 5185148 A	09-02-1993
		AU 684712 B	08-01-1998
		AU 3608693 A	01-09-1993
		CA 2129106 A	05-08-1993
		EP 0633936 A	18-01-1995
		EP 0816500 A	07-01-1998
		JP 7503846 T	27-04-1995
		US 5554534 A	10-09-1996